

● Rapid Oligonucleotides QC with Automated High Resolution ESI-LCMS and BioPharma Compass

Oligonucleotides – sequences of nucleotides (RNA and DNA) – are vital research tools that have a wide range of applications in genetic testing, research, and forensics.

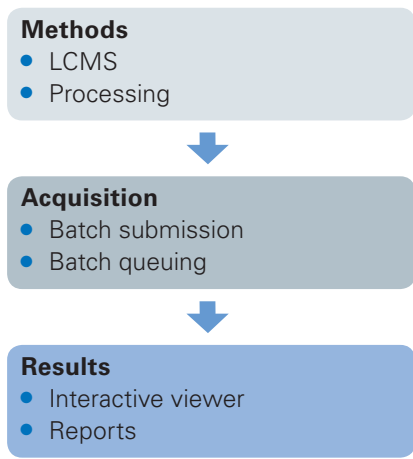
Abstract

Common applications include artificial gene synthesis, primers for polymerase chain reaction (PCR), DNA sequencing, molecular cloning and as molecular probes. Oligonucleotides are typically made in the laboratory by

a solid-phase chemical synthesis process using phosphoramidite chemistry. Modern synthesis platforms and high-throughput automation systems support robust oligonucleotide manufacturing capabilities. Despite these advances, oligonucleotide synthesis is a complex process,

involving hundreds of sequential chemical reactions. Quality control of precise and accurate synthesis is of high importance as it is ensuring that synthesis by-products or process contaminants are within acceptable limits.

Keywords:
Oligo QC, maXis II
QTOF, BioPharma
Compass



Steps involved in setting up the automated workflow

Mass Spectrometry (MS) is the technology of choice for analyzing oligonucleotides. It provides confirmation that the correct sequence has been synthesized with high mass accuracy and enables sensitive detection of low levels of by-products generated during synthesis process such as truncated or deleted sequences, depurination and incomplete removal of protecting groups. Mass spectrometry techniques and integrated software solutions facilitate oligonucleotide quality control by assessing oligonucleotide identity in a high-throughput manner.

Here we describe an automated UPLC-MS method for the quality control of synthetic oligonucleotides. The automated workflow is implemented using BioPharma Compass® software which encompasses data acquisition, processing and reporting using a seamless workflow solution.

Introduction

Dedicated software, such as BioPharma Compass (Bruker Daltonics), enables the routine analysis of biopharmaceuticals including oligonucleotides and offers LC-MS workflows with automated measurements and report generation. BioPharma Compass comprises a

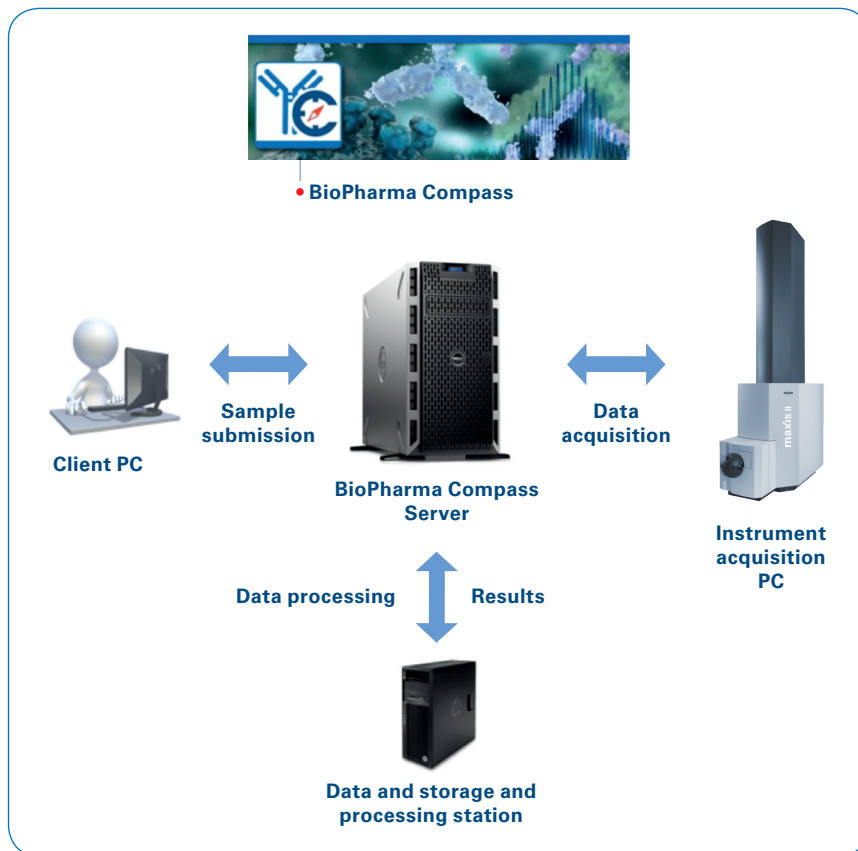


Figure 1: BioPharma Compass data flow

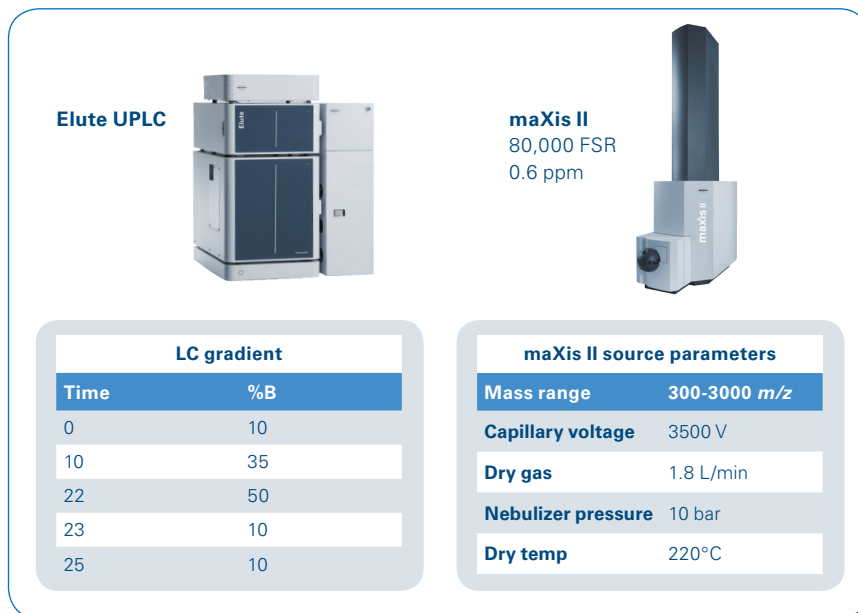


Figure 2: Elute HPLC gradient and maXis II parameters

secure server that communicates with the client PC and the instrument acquisition PC. The user can set up sample batches for acquisition; the data are acquired and can be automatically moved to the data storage

and processing station to preserve the data integrity and security. The results are accessible to the user from the BioPharma Compass server. The entire workflow is fully supported by 21 CFR Part 11 compliance features.

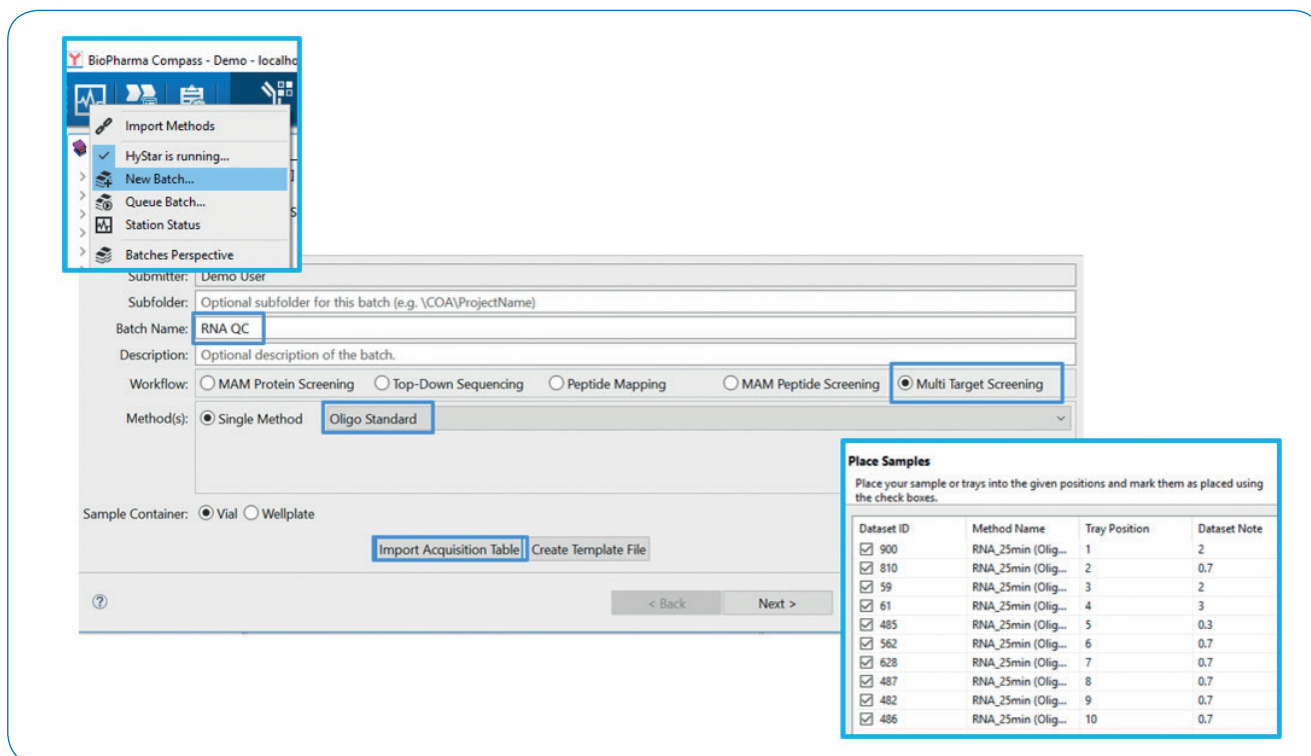


Figure 3: Automated acquisition set-up in BioPharma Compass

Experimental

A fully automated acquisition and processing workflow for rapid oligonucleotide QC was defined in BioPharma Compass. The LCMS method set included the LC separation and the detection by mass spectrometry. The separation was carried out on a Bruker Elute UPLC system (Bruker Daltonics) equipped with a BEH C4 1.7 μm 1x100 mm column at a flow rate of 500 $\mu\text{L}/\text{min}$ and a temperature of 80°C. The mobile phases were A – 1% HFIP, 0.1% Hexylamine in water and B – 50% ACN + 50% MeOH. One μg of synthetic oligonucleotide samples dissolved in water were injected on column. The detection was performed on a Bruker maXis II ETD UHR QTOF operated in negative ion mode. The LC gradient and MS source parameters are shown in Figure 2.

The Automated Acquisition button in the blue BioPharma Compass Ribbon allowed straightforward setting up of sample batches for acquisition. The

simple interface required minimal input from the user for a batch name, workflow and method as seen in Figure 3. A LIMS generated worklist can be directly imported. Once the samples are placed in the autosampler positions, the acquisition of the queued batch is initiated.

The processing view in BioPharma Compass allows monitoring of the submitted queue. As the data is

acquired, the data mover feature automatically moves it to the data storage and processing station preventing direct user access, thus preserving data integrity and maintaining security. Spectra were obtained across the chromatographic peaks and charge deconvoluted using a Maximum Entropy algorithm yielding neutral average mass molecular weights.


Type	Name	User	Proce...	Status	Start	Finis
Multi Target Screening...	RNA QC-[113]	Dem...	0 / 10	running	2020-05...	

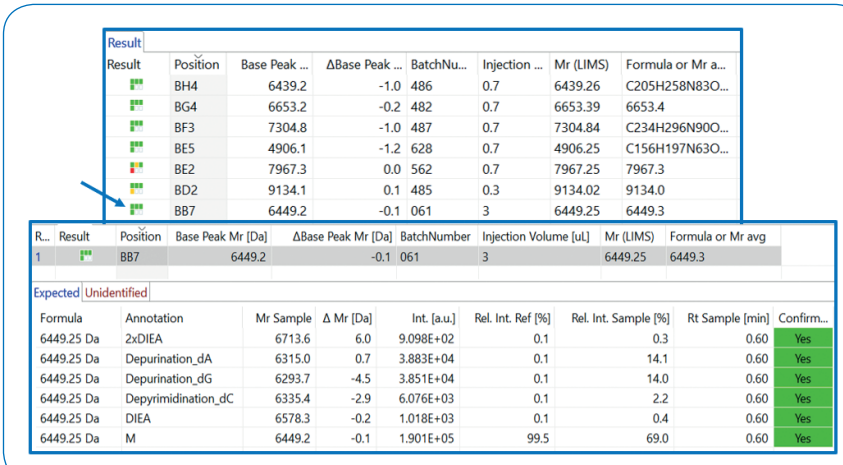
#	Algorithm / Method	Status	Pr...	Start	Finish...	Name	URL	Message
10	Oligo Standard_1.0	waiting		2020-...		486-[141]		
9	Oligo Standard_1.0	waiting		2020-...		482-[140]		
8	Oligo Standard_1.0	waiting		2020-...		487-[139]		
7	Oligo Standard_1.0	waiting		2020-...		628-[138]		
6	Oligo Standard_1.0	waiting		2020-...		562-[137]		
5	Oligo Standard_1.0	waiting		2020-...		485-[136]		
4	Oligo Standard_1.0	waiting		2020-...		61-[135]		
3	Oligo Standard_1.0	waiting		2020-...		59-[134]		
2	Oligo Standard_1.0	waiting		2020-...		810-[133]		
1	Oligo Standard_1.0	running	0 %	2020-...		900-[132]		idle








Figure 4: Processing view shows status of queued batch


Results

BioPharma Compass has an interactive user-friendly interface that allows users to see the results at-a-glance. Several views highlight different aspects of the data such as the result tables, chromatograms and spectra. Screening of batches is facilitated through similarity scores and mass accuracy attributes which are the basis for an automatic pass/fail assessment. Reports utilize color coding of results and charts for rapid reviewing.

The Result table summarizes the analyses in the sample table including the MAM reporting icon , base peak of the spectrum and mass difference to the base peak. Clicking one of the rows in the results table shows the expected table view seen in Figure 5 with the detailed modifications for that specific analysis. The relative intensities of the sample versus the reference allow comparison of the measured profile to the reference profile.

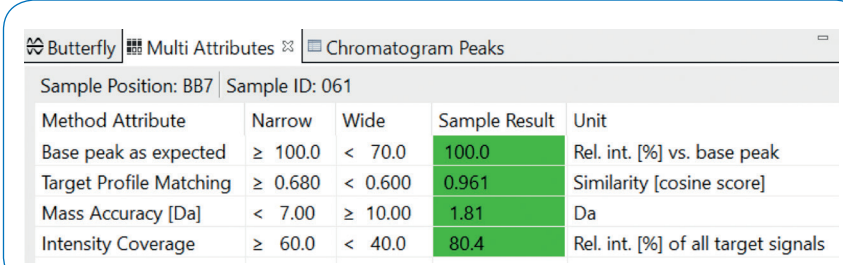


Result	Position	Base Peak ...	ΔBase Peak ...	BatchNu...	Injection ...	Mr (LIMS)	Formula or Mr a...
	BH4	6439.2	-1.0	486	0.7	6439.26	C205H258N83O...
	BG4	6653.2	-0.2	482	0.7	6653.39	6653.4
	BF3	7304.8	-1.0	487	0.7	7304.84	C234H296N90O...
	BE5	4906.1	-1.2	628	0.7	4906.25	C156H197N63O...
	BE2	7967.3	0.0	562	0.7	7967.25	7967.3
	BD2	9134.1	0.1	485	0.3	9134.02	9134.0
	BB7	6449.2	-0.1	061	3	6449.25	6449.3

R...	Result	Position	Base Peak Mr [Da]	ΔBase Peak Mr [Da]	BatchNumber	Injection Volume [uL]	Mr (LIMS)	Formula or Mr avg
1		BB7	6449.2	-0.1	061	3	6449.25	6449.3

Formula	Annotation	Mr Sample	Δ Mr [Da]	Int. [a.u.]	Rel. Int. Ref [%]	Rel. Int. Sample [%]	Rt Sample [min]	Confirm...
6449.25 Da	2xDI EA	6713.6	6.0	9.098E+02	0.1	0.3	0.60	Yes
6449.25 Da	Depurination_dA	6315.0	0.7	3.883E+04	0.1	14.1	0.60	Yes
6449.25 Da	Depurination_dG	6293.7	-4.5	3.851E+04	0.1	14.0	0.60	Yes
6449.25 Da	Depyrimidination_dC	6335.4	-2.9	6.076E+03	0.1	2.2	0.60	Yes
6449.25 Da	DI EA	6578.3	-0.2	1.018E+03	0.1	0.4	0.60	Yes
6449.25 Da	M	6449.2	-0.1	1.901E+05	99.5	69.0	0.60	Yes

Figure 5: Results summary table and detailed expected results table showing fragmentation products and adducts resulting from the chromatographic additives.



Method Attribute	Narrow	Wide	Sample Result	Unit
Base peak as expected	≥ 100.0	< 70.0	100.0	Rel. int. [%] vs. base peak
Target Profile Matching	≥ 0.680	< 0.600	0.961	Similarity [cosine score]
Mass Accuracy [Da]	< 7.00	≥ 10.00	1.81	Da
Intensity Coverage	≥ 60.0	< 40.0	80.4	Rel. int. [%] of all target signals

Figure 6: Multi-attribute table provides a legend to each individual sample result.

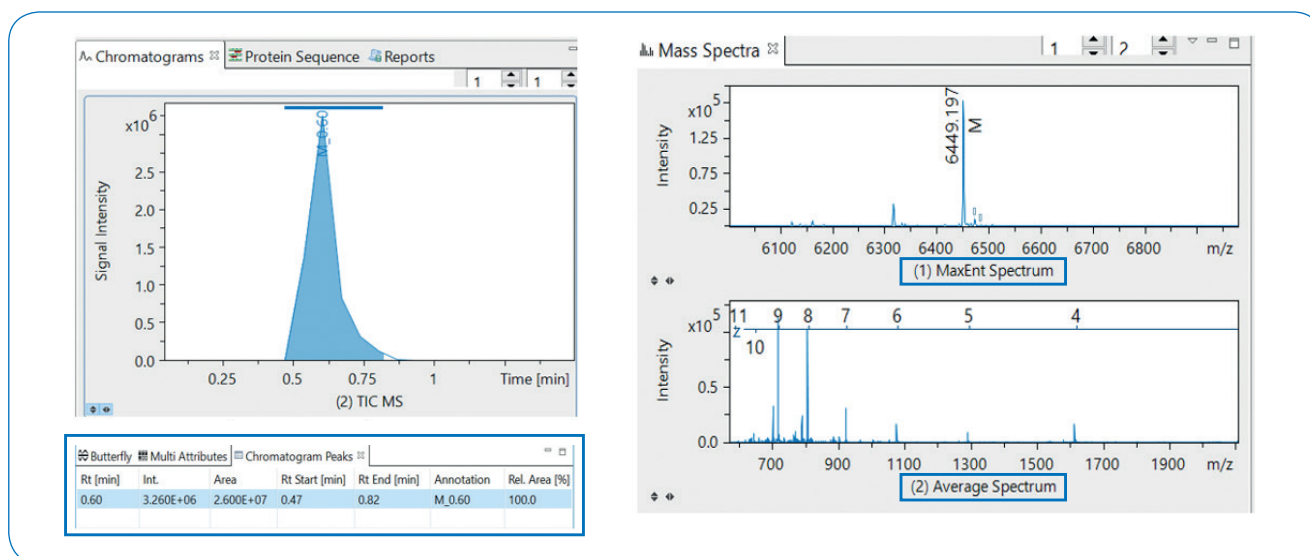


Figure 7: Chromatogram and spectrum views



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<http://www.bruker.com/massspectrometry>



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