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## NTRODUCTION

In biopharmaceutical analysis, undesired secondary analyte/surface interactions have hindered performance in HPLC. Analytes that contain electron rich functional groups are susceptible to adsorb onto surfaces along the stainless-steel flow path causing reduced resolution and recovified with a chemically resistant hybrid organic inorganic barrier called MaxPeak ${ }^{\text {TM }}$ High Performance Surfaces (HPS) Technology This technology was then used in the construction of a bio-inert system called the Alliance ${ }^{\text {TM }}$ iS Bio HPLC System.

In this poster, the Alliance is Bio HPLC System was evaluated by analysing oligonucleotides, monoclonal antibodies (mAbs), and glucagon-like-peptides (GLP-1) and compared to a legacy HPLC system for improved resolution and recovery

## METHODS

Established methods that were previously analyzed on legacy HPLC platforms and columns were scaled and transferred to the Alliance iS Bio HPLC System. Ion-pairing reversed phase chromatography (IPRPLC), size exclusion chromatograph (SEC), and RPLC were used as the prevailing techniques for analyzing the analytes on both systems.

## Oligonucleotide Analysis ${ }^{1}$

Waters MassPREP ${ }^{\text {TM }}$ Oligonucleotide Standard (OST) containing 15-35 mer oligodeoxythymidines ( 4 pmol/ $\mu \mathrm{L}$ ) and GEM91, a 25 mer fully thiolated phosphorothioate oligonucleotide $(0.5 \mathrm{mg} / \mathrm{mL})$ were injected onto both systems.

Legacy HPLC System Method Conditions:
Column: $\quad$ XBridge ${ }^{\text {TM }} \mathrm{BEH}^{\text {TM }} \mathrm{C}_{18}$ Column $5 \mu \mathrm{~m}, 130 \AA$, $4.6 \times 100 \mathrm{~mm}$ ( $\mathrm{p} / \mathrm{n}: 186003115$ ) XBridge BEH C ${ }_{18}$ Column $2.5 \mu \mathrm{~m}, 130 \AA$, $4.6 \times 100 \mathrm{~mm}(\mathrm{p} / \mathrm{n}: 186006039)$
Alliance is Bio HPLC System Method Conditions:
Column: XBridge Premier Oligonucleotide BEH C Column $2.5 \mu \mathrm{~m}, 130 \AA, 4.6 \times 100 \mathrm{~mm}$ (p/n:186009902) $6 \times 100 \mathrm{~mm}$ (p/n: 186003115 ) $5 \mathrm{~m}, 130 \AA$ $4.6 \times 100 \mathrm{~mm}$ ( $\mathrm{p} / \mathrm{n}: 186003115$ )

Shared Conditions
25 mM Hexylammonium acetate (HAA) in
Mobile Phase B: $\quad 25 \mathrm{mM}$ HAA in water/acetonitrile (ACN)
Mobile Phase C. $\quad$.
Mobil Phase D:
Mobile Phase
Injection volume
Column temp.
Wavelength:
MassPREP OST: $\quad 1.73 \% \mathrm{~B} / \mathrm{min}$ gradient
$0.5 \% \mathrm{~B} / \mathrm{min}$ gradient

## Monoclonal Antibody Analysis ${ }^{2}$

USP mAb reference standards were injected at a concentration of 10 $\mathrm{mg} / \mathrm{mL}$ in formulation buffer onto both systems

## Legacy HPLC System Method Conditions: ${ }^{3}$

Column: $\quad$ BioSuite ${ }^{\text {TM }}$ Diol ( OH ) Column, 250A $\mu$, $7.8 \mathrm{~mm} \times 300 \mathrm{~mm}$ ( $\mathrm{p} / \mathrm{n}$ : 186002165) Flow Rate. $\quad 0.500 \mathrm{~mL} / \mathrm{min}$
Run Time: $\quad 30$ minutes, isocratic
Alliance iS Bio HPLC System Method Conditions
Column: XBridge Premier Protein SEC Column 250
Injection volume: $\quad-3.5 \mu \mathrm{~m}, 78 \times 150 \mathrm{~mm}$ ( $\mathrm{p} / \mathrm{n}: 186009961$ )
$3.5 \mu \mathrm{~L}$
Run Time:
7.5 minutes, isocratic

## Shared Conditions: ${ }^{3}$

Mobile Phase: $\quad 0.20 \mathrm{M}$ potassium phosphate and 0.25 M
Column temp.
potassium chloride, pH 6.2
Wavelength:
280 nm

## GLP-1 Analysis

Dulaglutide and glucagon stock were prepared with DMSO at $1 \mathrm{mg} / \mathrm{mL}$ Liraglutide and tirzepatide stock were prepared with DMSO at $0.5 \mathrm{mg} /$ mL . Exenatide and semaglutide stock were prepared with 10 mM ammonium formate buffer, pH 8.5 at $0.5 \mathrm{mg} / \mathrm{mL}$. The GLP-1 pane $0.5 \%$ trilus pre $1 \%$ for

Legacy HPLC System Method Conditions
Column: $\quad$ XSelect ${ }^{\text {TM }}$ Peptide CSH $^{\text {TM }} \mathrm{C}_{18}$ Column 130 $\AA$
$2.5 \mathrm{~m}, 4.6 \times 150 \mathrm{~mm}(\mathrm{p} / \mathrm{n} \cdot 186007038$ )
Alliance iS Bio HPLC System Method Conditions:
Column: XSelect Premier Peptide CSH C ${ }_{18}$ Column 130 $\AA$, $2.5 \mu \mathrm{~m}, 4.6 \times 150 \mathrm{~mm}(\mathrm{p} / \mathrm{n}: 186009909)$

## Shared Conditions:

Mobile Phase A:
Column temp.:
Wavelength:
Injection volume
Flow Rate
$.1 \%$ formic acid in ACN
$60^{\circ} \mathrm{C}$
214 nm
$10 \mu \mathrm{~L}$
$0.960 \mathrm{~mL} / \mathrm{min}$
$2.725 \% \mathrm{ACN} / \mathrm{m}$

RESULTS AND DISCUSSION


Figure 2: GEM91 was analyzed to further investigate the performance differences across systems. Despite its refinement as a drug substance, GEM91 contains impurities that necessitate monitoring. Utilizing the $5 \mu \mathrm{~m}$ XBridge BEH $\mathrm{C}_{18}$ Column on both systems, the Alliance iS Bio HPLC System showed an $\sim 40 \%$ increase in the signal-to-noise ratio for the trace impurities versus the legacy HPLC system. This improvement translates to enhanced recovery and increased accuracy when analyzing critical species for novel therapies.



CONCLUSION

- The biocompatible and bio-inert construction of the Alliance iS Bio HPLC System is well suited for biotherapeutic analysis of oligonucleotides, monoclonal antibodies, and small biologics such as GLP-1 analytes.
- The Alliance iS Bio HPLC System demonstrated increased resolution and recovery of biotherapeutics while delivering consistent performance with improved precision.
- The larger mixing volume enhances low-level impurities for improved accuracy in detection and integration.


## References

Du X, Birdsall RE, Bigos P. Han D. Nyholm K. Deploying the Alliance ${ }^{\text {Tw }}$ is Bio HPLC Syster as a modern HPLC Cor biopharmaceel 2. Bispos P PBirdsall RE, Nyholm K. Modernizing Compendial SEC Methods for Bioth 200008290EN.

Figure 4: The larger mixing volume of the Alliance is Bio HPLC System produces chromatograms with lower baseline noise and improved peak shape for the GLP-1 peptides when compared to the legacy HPLC system. This enables improved accuracy in the detection and integration of low abundant impurities and main peaks

