

GLYCOWORKS *RapiFluor*-MS QUICK START PROTOCOL

STREAMLINED PROTOCOL – 96 SAMPLE (4 x 24 FORMAT)

- Set heat blocks to at least 90 °C and 50 °C.
- Protocol is based on 1.5 mg/mL glycoprotein starting concentration.



STEP 1: Rapid Deglycosylation

1. Reconstitute 1 vial of the Intact mAb Mass Check Standard (1 mg/vial) in 670 μL 18.2 M Ω water to create a 1.5 mg/mL solution.

Note: For glycoproteins with a formulation buffer containing nucleophiles or anionic reagents (e.g., His, Gly, Tris, PO₄²⁻), if a Glycan C₁₈ AX Column is applied for LC separation, it is highly recommended to desalt the sample with water prior to Step 1.

2. Prepare 3% (w/v) *RapiGest*™ SF by dissolving 10 mg of *RapiGest* SF Surfactant in 200 μL of Rapid Buffer and 135 μL water, vortex.
3. Dilute PNGase F enzyme (35 μL) with 220 μL water for a total of 255 μL .
4. Add 10 μL of 1.5 mg/mL glycoprotein into the provided tube.
5. Add 10 μL of buffered 3% (w/v) *RapiGest* SF solution to above tube, aspirate to mix.
6. Heat at least to 90 °C for 3 minutes.
7. Cool at room temperature for 3 minutes.
8. Add 10 μL Rapid PNGase F and aspirate to mix.
9. Incubate at 50 °C for 5 minutes.
10. Cool at room temperature for 3 minutes.

STEP 2: Rapid Labeling of Glycosylamines

1. Add 280 μL of anhydrous DMF directly to one vial of 23 mg of *RapiFluor*-MS™ Reagent. Mix to solubilize.
2. Add 10 μL of the *RapiFluor*-MS solution to the deglycosylation mixture and aspirate to mix.
3. Allow the labeling to proceed at room temperature for 5 minutes.
4. Dilute the reaction with 360 μL of acetonitrile (ACN) and aspirate to mix.

STEP 3: HILIC Cleanup of Labeled Glycosylamines

1. Set up a GlycoWorks™ HILIC $\mu\text{Elution}$ Plate and add in shims or spacer and waste tray.
2. Condition wells by adding 200 μL of water per well.
3. Equilibrate wells by adding 200 μL 85% ACN.
4. Load ACN-diluted samples (~400 μL).
5. Wash wells with two (2) 600 μL volumes of 1% formic acid, 90% ACN.
6. Replace waste tray with sample collection tray loaded with 600 μL tubes.
7. Elute glycans with three (3) 30 μL volumes of SPE Elution Buffer into 600 μL tapered bottom inserts.
8. Dilute SPE eluate with 310 μL of the GlycoWorks SPE Diluent (DMF/ACN). Aspirate to mix.
Note: For a Glycan C₁₈ AX separation sample, either skip dilution Step 8 or dilute with 310 μL of water.
9. Cap the tubes with pre-slit cap mats.

► For the complete Care and Use Manual, visit [waters.com](https://www.waters.com) and search [715004793EN](#).

► For more details on this method, download Application Notes [720005506EN](#) and [720007038EN](#).