

# An Analytically Validated Bioanalytical Clinical Research Method for the Quantification of Acylcarnitines in Urine

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# **APPLICATION BENEFITS**

- Direct analysis of carnitine and acyl carnitines in urine using gradient reversed-phase UltraPerformance Chromatography (UPLC®) combined with tandem quadrupole MS
- Rapid analysis
- Minimal sample preparation, without the need for derivatization prior to analysis

### INTRODUCTION

The amino acid derivative carnitine is involved in lipid metabolism, transporting long-chain acyl groups from fatty acids to the mitochondrial matrix, where they are broken down to form usable energy via the citric acid cycle. The analysis of these acylcarnitines in blood, urine, and plasma for clinical research studies is typically performed by tandem quadrupole MS coupled to gas chromatography, high-performance liquid chromatography, or capillary electrophoresis.<sup>2</sup> In this application note, we describe an analytically validated bioanalytical clinical research method for the quantification of low concentration acylcarnitines in urine.<sup>3,4</sup>

# WATERS SOLUTIONS

Xevo® TQ-S

Open Architecture UPLC System

**ACQUITY UPLC® HSS T3 Column** 

TargetLynx™ Software

### **KEYWORDS**

Acylcarnitine, urine, quantification

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### **EXPERIMENTAL**

# Sample description

Urine samples, stored at -80 °C, were left to thaw at 4 °C. A 30  $\mu$ L aliquot of urine was centrifuged at 10,300 g for 10 minutes. Then, 20  $\mu$ L of supernatant from each sample was diluted with 65  $\mu$ L of water, 5  $\mu$ L of acetonitrile, and 10  $\mu$ L of the internal standard mixture. The internal standard mixture consisted of propionyl-L-carnitine-d<sub>3</sub>; butyryl-L-carnitine-d<sub>3</sub>, hexanoyl-L-carnitine-d<sub>3</sub> and decanoyl-L-carnitine-d<sub>3</sub> at 250 ng/mL and isovaleryl-L-carnitine-d<sub>9</sub>, octanoyl-L-carnitine-d<sub>3</sub> and decanoyl-L-carnitine-d<sub>3</sub> at 125 ng/mL. Tetradecanoyl-L-carnitine-d<sub>3</sub> and hexadecanoyl-L-carnitine-d<sub>3</sub> were also included at 125 ng/mL for testing of these analytes.

# **Method conditions**

The UPLC-MS analysis was performed on an ACQUITY UPLC binary solvent manager, 2777 sample manager (CTC) and column manager (Waters, Milford, MA, USA) interfaced to a Xevo TQ-S Tandem Quadrupole Mass Spectrometer (Waters, Milford, MA, USA).

LC conditions		MS conditions	
LC system:	Open Architecture UPLC System	MS system:	Xevo TQ-S
Detection:	Xevo TQ-S	Ionization mode:	Positive ion
Vials:	Waters® certified vials	Acquisition range:	MRM mode
Column:	ACQUITY UPLC HSS T3	Capillary voltage:	2.75 kV
	1.8 μm, 2.1 mm x 150 mm	Collision energy:	14-26 eV
Column temp.:	45 °C	Cone voltage:	8-48 V
Sample temp.:	4 °C		
Injection volume:	2 μL	Data management	

Chromatography

software: MassLynx® Software
MS software: MassLynx Software
Informatics: TargetLynx™ Software

# **RESULTS AND DISCUSSION**

0.5 mL/min

0.1% formic acid

0.1% formic acid in acetonitrile

Gradient mode elution

The chromatographic separation was developed to provide sufficient resolution of the analytes, with a run time of 10 minutes consistent with medium to high throughput analysis, while maintaining the separation between the two pairs of isomeric species isovaleryl-L-carnitine (isoC5) and valeryl-L-carnitine (C5), and isobutyryl-L-carnitine (isoC4) and butyryl-L-carnitine (C4). Due to their relatively low concentrations in urine, compared to carnitine itself and some other high concentration acylcarnitines, the analytes measured in this method, propionyl-L-carnitine (C3), butyryl-L-carnitine (C4), isovaleryl-L-carnitine (isoC5), valeryl-L-carnitine (C5), hexanoyl-L-carnitine (C6), octanoyl-L-carnitine (C8), decanoyl-L-carnitine (C10) and dodecanoyl L-carnitine (C12) need to be quantified using a method optimized for low concentration acylcarnitines. Dodecanoyl L-carnitine (C12), tetradecanoyl-L-carnitine (C14) and hexadecanoyl-L-carnitine (C16) are too low in concentration in the control urine to be quantified using the sample preparation method for this analysis, however, the internal standards are displayed in Figure 1B for the purposes of demonstrating retention time and could allow for testing of these analytes.

### LC-MS

Flow rate:

Mobile phase A:

Mobile phase B:

The samples were analyzed using a reversed-phased gradient separation with the methodology optimized for analyte resolution and reproducibility. The analyses were detected and quantified by positive ion MRM with optimal MS conditions determined by the infusion of authentic analyte and stable isotope labelled internal standards to determine the maximum response for each transition. For quantification, one transition from each of these precursor ions was used to calculate unknown concentrations. The choice of quantification transition was determined from the ionization energy of each compound.

# [APPLICATION NOTE]

Typical chromatograms for the UPLC-MS/MS analysis of the acylcarnitine standards are shown in Figure 1A and 1B. Figure 2 shows a typical chromatogram of a urine sample diluted one part in five. All eight acylcarnitines examined were chromatographically resolved. The separation was shown to be highly reproducible, with retention time coefficients of variance for the acylcarnitine standards at the ULOQQC of <0.20% across the entire validation run (n=18).

### **LINEARITY**

The method was linear over the calibration ranges employed for the individual analytes, using a linear fit with a weighting of 1/X. The correlation coefficient (r²) values of the individual standard curves were determined to be between 0.988 and 0.999 for all analytes.

#### **INTRA-DAY VALIDATION**

The method was assessed for accuracy linearity and bias over three days for the concentration ranges 7.5–100 ng/mL for propionyl-L-carnitine (C3); 3–40 ng/mL for butyryl-L-carnitine (C4), valeryl-L-carnitine (C5), isovaleryl-L-carnitine (isoC5), hexanoyl-L-carnitine (C12); 1.5–20 ng/mL for octanoyl-L-carnitine (C8) and decanoyl-L-carnitine (C10).

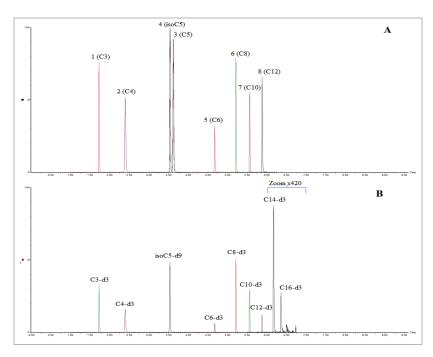


Figure 1. Chromatographic separation of the ULOQQC acylcarnitine standards (A) natural abundance species (B) SIL species. Note at the retention times of C14- $d_3$  and C16- $d_3$ , B is zoomed in 420 times to show IS peaks.

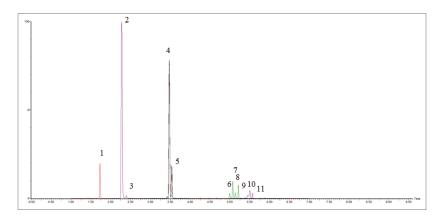


Figure 2. Typical urine sample diluted one part to five. Peak annotation;

- 1) propionyl-L-carnitine,
- 2) isobutyry-L-carnitine,
- 3) butyry-L-carnitine,
- 4) isovaleryl L-carnitine,
- 5) valeryl L-carnitine,
- 6, 7, 8) isomers of octanoyl-L-carnitine,
- 9) octanoyl-L-carnitine,
- 10) isomers of decanoyl-L-carnitine,
- 11) decanoyl-L-carnitine

# [APPLICATION NOTE]

All of the analytes monitored showed excellent accuracy and precision. As an example, the coefficient of variance and mean concentration data obtained for the method data for day three are shown in Tables 1 and 2 respectively.

Table 1. Summary of the coefficient of variance for day three

CV	LLOQQC1	LLOQQC2	LQC	MQC	HQC	ULOQQC
Propionyl-L-carnitine, C3	1.67	2.06	2.35	5.88	2.11	4.62
Butyryl-L-carnitine, C4	2.61	2.17	1.89	5.49	1.37	4.12
Valeryl-L-carnitine, C5	1.89	1.46	2.17	5.47	1.30	3.68
Isovaleryl-L-carnitine, isoC5	1.92	2.13	2.12	4.82	1.59	4.25
Hexanoyl-L-carnitine, C6	1.70	1.84	3.02	5.58	1.47	4.76
Octanoyl-L-carnitine, C8	2.58	1.93	2.71	6.39	2.38	4.38
Decanoyl-L-carnitine, C10	3.16	3.89	3.52	8.02	4.31	6.55
Dodecanoyl-L-carnitine, C12	4.50	4.40	10.9	12.10	9.8	14.4

Numbers rounded to three significant figures, or integers, as appropriate.

Table 2. Summary of the mean concentration data for day three

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Mean	LLOQQC1	LLOQQC2	LQC	MQC	HQC	ULOQQC
nominal concentration (ng/mL)	7.5	10	20	30	80	100
Propionyl- L-carnitine C3	7.02	9.42	19.3	29.3	77.6	96.9
nominal concentration (ng/mL)	3	4	8	12	32	40
Butyryl-L-carnitine, C4	2.88	3.85	7.78	11.7	31.1	38.8
Valeryl-L-carnitine, C5	2.73	3.75	7.70	11.6	30.6	38.0
Isovaleryl-L-carnitine, isoC5	2.85	3.83	7.75	11.7	30.8	38.3
Hexanoyl-L-carnitine, C6	3.03	4.08	8.05	11.6	29.9	36.7
Dodecanoyl-L-carnitine C12	3.03	3.80	7.90	11.7	30.0	33.7
nominal concentration (ng/mL)	1.5	2	4	6	16	20
Octanoyl-L-carnitine, C8	1.58	2.12	4.32	6.52	17.1	21.4
Decanoyl-L-carnitine, C10	1.63	2.15	4.40	6.63	17.3	21.6

Numbers rounded to three significant figures, or integers, as appropriate.

#### INTER-DAY VALIDATION

The inter-day validation results are summarized in Table 3 and Table 4. Table 3 summarizes the inter-day coefficients of variance for each QC standard and Table 4 summarizes the inter-day mean concentration for each QC standard. The precision of the back calculated standards across the inter-day validation ranged from 0.89 to 9.75%, while the mean back calculated concentrations obtained for each of the standards gave mean bias values ranging from 3.25 to 8.20%. The Lower Limit of Quantification (LLOQ) was defined as 7.5 ng/mL for propionyl-L-carnitine (C3), 3 ng/mL for butyryl-L-carnitine (C4), valeryl-L-carnitine, (C5), isovaleryl-L-carnitine (isoC5) and hexanoyl-L-carnitine (C6), and 1.5 ng/mL for octanoyl L-carnitine (C8). However, for dodecanoyl-L-carnitine the LLOQ was 4 ng/mL and decanoyl- L-carnitine (C10) was 2 ng/mL, although the standards were tested to 3 ng/mL and 1.5 ng/mL respectively. The mean bias of the method ranged from -1.7% for decanoyl-L-carnitine, (C10) to +14.7% for dodecanoyl-L-carnitine (C12).

Table 3. Summary of the inter-day coefficient of variance

Acylcarnitine	LLOQQC1	LLOQQC2	LQC	MQC	HQC	ULOQQC
Propionyl- L-carnitine, C3	3.61	3.22	2.57	3.57	2.34	3.76
Butyryl-L-carnitine, C4	5.73	6.06	5.10	7.35	6.50	7.03
Valeryl-L-carnitine, C5	3.80	2.52	1.82	3.46	2.35	3.14
Isovaleryl-L-carnitine, isoC5	6.33	5.51	5.64	5.70	4.94	5.15
Hexanoyl-L-carnitine, C6	2.53	2.47	2.49	3.87	2.44	3.92
Octanoyl-L-carnitine, C8	5.41	3.83	5.37	6.53	4.77	5.53
Decanoyl-L-carnitine, C10	7.24	4.74	6.12	7.62	7.37	8.44
Dodecanoyl-L-carnitine, C12	25.2	19.0	8.94	8.56	11.2	12.0

Numbers rounded to three significant figures, or integers, as appropriate.

Table 4. Summary of the inter-day mean concentration data

Mean	LLOQQC1	LLOQQC2	LQC	MQC	HQC	ULOQQC
nominal concentration (ng/mL)	7.5	10	20	30	80	100
Propionyl- L-carnitine, C3	6.85	9.22	18.7	28.0	74.4	92.7
nominal concentration (ng/mL)	3	4	8	12	32	40
Butyryl-L-carnitine, C4	2.84	3.81	7.62	11.5	30.5	37.9
Valeryl-L-carnitine, C5	2.68	3.64	7.39	11.1	29.4	36.5
Isovaleryl-L-carnitine, isoC5	3.07	4.11	8.25	12.4	32.7	40.6
Hexanoyl-L-carnitine, C6	2.97	3.98	7.96	11.7	30.8	38.2
Dodecanoyl-L-carnitine, C12	3.83	4.71	8.25	12.3	29.8	36.2
nominal concentration (ng/mL)	1.5	2	4	6	16	20
Octanoyl-L-carnitine, C8	1.53	2.04	4.05	6.09	16.2	20.0
Decanoyl-L-carnitine, C10	1.63	2.11	4.11	6.14	15.9	19.8

Numbers rounded to three significant figures, or integers, as appropriate.

### **CARRYOVER**

Carryover was assessed by running a double blank (water, containing no internal standard and no analytes) immediately after a upper limit of quantitation (ULOQ) calibration standard. The results obtained indicate that there was no significant carryover for the acylcarnitine standards. Carryover is the peak area of a compound found in the blank sample calculated as a percentage of the peak area of the compound in the lower limit of quantification quality control (LLOQQC). The highest percentage carry over at 14.27% was observed for dodecanoyl-L-carnitine. This is within the accepted limit of 20%. The internal standards carryover, at less than 0.98% for all compounds, was negligible.

# [APPLICATION NOTE]

### **CONCLUSIONS**

Based on the use of stable isotope labelled internal standards, a quantitative clinical research method was validated over the range 7.5–100 ng/mL (37.5–500 ng/mL in urine) for propionyl-L-carnitine (C3) and 3–40 ng/mL (15–200 ng/mL in urine) for butyryl-L-carnitine (C4), valeryl-L-carnitine, (C5), isovaleryl-L-carnitine (isoC5) and hexanoyl-L-carnitine (C6). Dodecanoyl-L-carnitine (C12) ranged from 4–40 ng/mL (20–200 ng/mL in urine) while the octanoyl-L-carnitine (C8) range is from 1.5–20 ng/mL (7.5–100 ng/mL in urine) and decanoyl-L-carnitine (C10) from 2–20 ng/mL (10–100 ng/mL in urine). The methodology required only 30 µL of urine and exhibited excellent analytical sensitivity, robustness, and reproducibility.

### References

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The analytical data presented in this application note are intended to demonstrate the robustness of a Waters research method. These data in no way substitute for independent method validation required by any applicable legal or laboratory standards.



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