

i-Series Cannabis Analyzer

- for Potency Testing -

CIPS
Shimadzu Europa GmbH



Outline

1. Introduction

- Overview of cannabis chemistry and potency testing

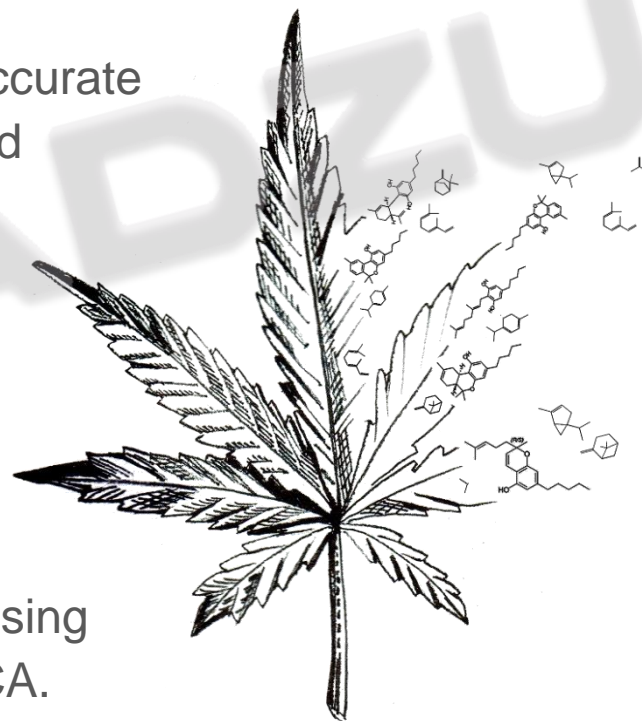
2. The Cannabis Analyzer for Potency Testing

- What is included in the package
- Goal oriented analytical methods
- Quantitative accuracy and analytical results
- Sample preparation
- Workflow in the dedicated software

Introduction

• Overview of Cannabis Testing

- Cannabis contains more than 500 unique compounds, including over 80 alkaloids known as cannabinoids.
- QC testing for cannabinoids is essential for the accurate labeling of cannabis products for both medical and recreational use.
- Cannabis “potency” is normally reserved for the quantitation of the major cannabinoids, namely THCA, THC, CBD and CBN
- HPLC has emerged as the gold standard for potency determinations because separation and detection of the cannabinoids is done without causing any decomposition of the naturally abundant THCA. This acid form undergoes decarboxylation to THC under the influence of heat and light.



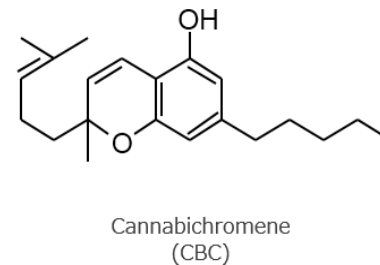
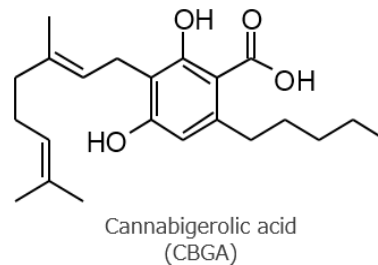
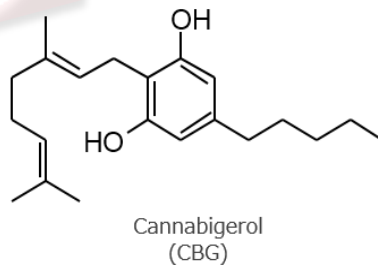
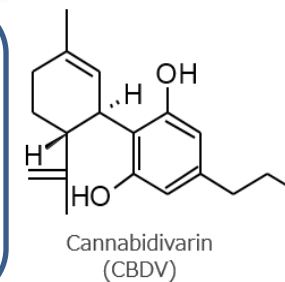
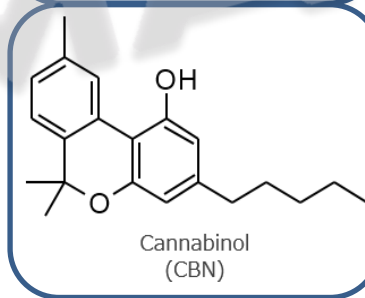
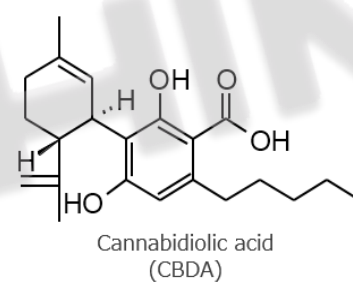
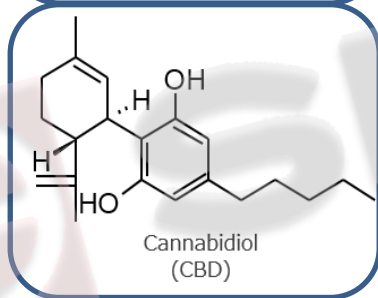
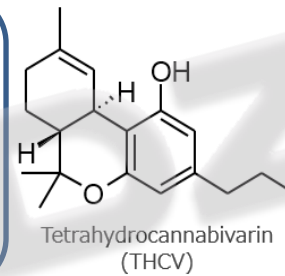
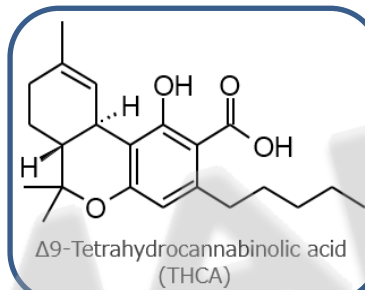
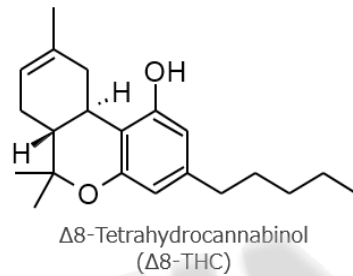
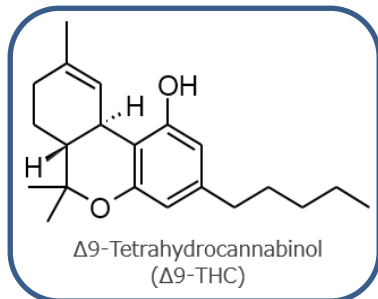
Importance of Cannabis

- THC does not occur naturally in cannabis plants. THC-A is the natural, non-psychoactive, carboxylic acid form of THC. It is rapidly converted to THC upon smoking or heating.
- The premium products sold in medicinal cannabis dispensaries are typically those with high THC concentrations, however, it's THC-A and the other non-psychoactive “CBx” cannabinoids, such as CBD that have been reported to reduce convulsions, inflammation, nausea and anxiety, and even eradicate tumors in some patients.
- CBN, only slightly psychoactive, is a degradation byproduct of THC and it is elevated when cannabis is poorly stored.



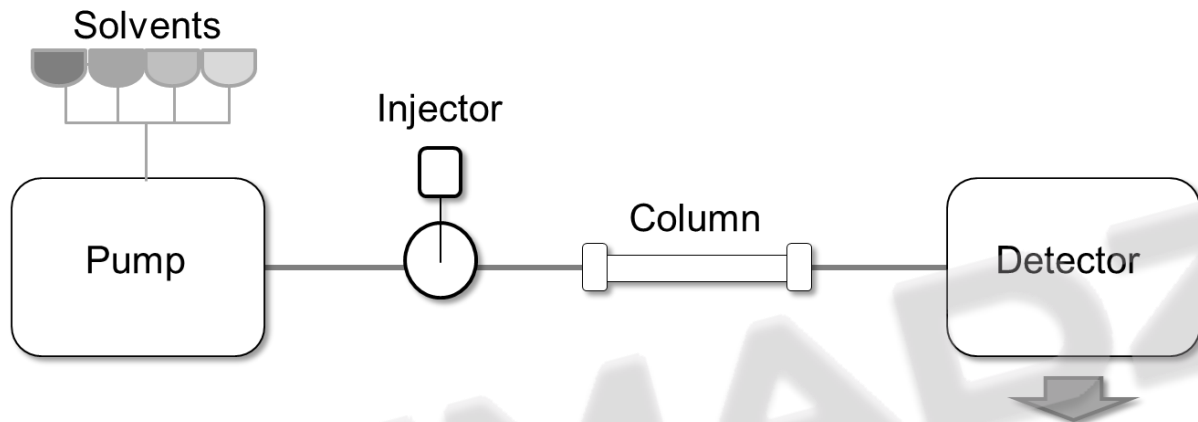
Structure Formula

- Naturally occurring cannabinoids of primary interest for potency determination

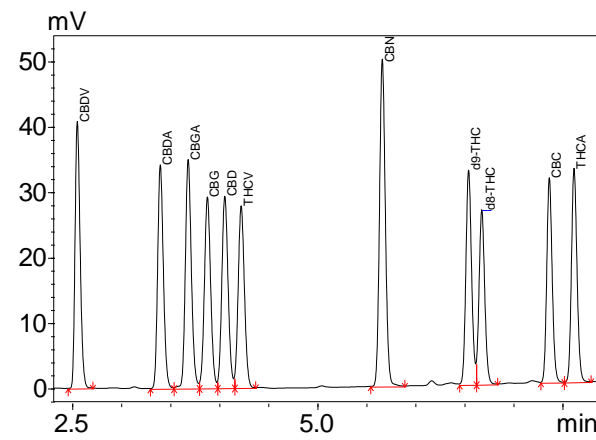
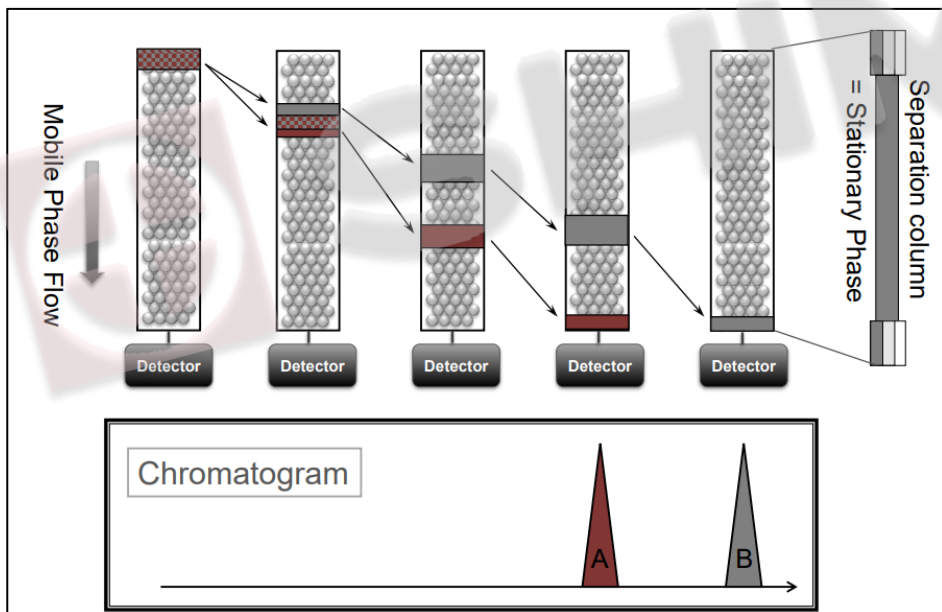


Principle of Liquid Chromatography

HPLC: High Performance Liquid Chromatography



Chromatogram



The i-Series Cannabis Analyzer

- **A complete solution for quantitative determination of cannabinoid content**

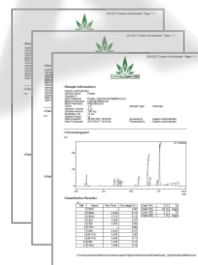
- The analyzer is designed for potency determination by quantitation of the active constituents. It includes all required hardware, software, consumables and procedures, so the analyst is running samples in the shortest possible time.



Columns and guard columns



Software and Method Package



App notes, sample preparation procedures, report templates



Prominence-i compact HPLC instrument with integrated UV detector

- Three proprietary instrument methods to meet your analytical objectives. No need to spend valuable lab time developing methods – just run samples.



Included Methods

- **High Throughput HPLC Method Package**

- Designed for analysis of the 10 most commonly requested cannabinoids in under 8 minutes. (Does not include THCV)



- **High Sensitivity HPLC Method Package**

- Adds THCV to the target analyte list, with an instrument cycle time of under 10 minutes. The short analysis time produces the sharpest chromatographic peaks for the best overall sensitivity.



- **High Resolution HPLC Method Package**

- Full baseline resolution for all 11 compounds and an analysis time under 30 minutes. This method is preferred for research purposes, or when additional compounds must be added to the analysis in response to new state regulatory requirements.

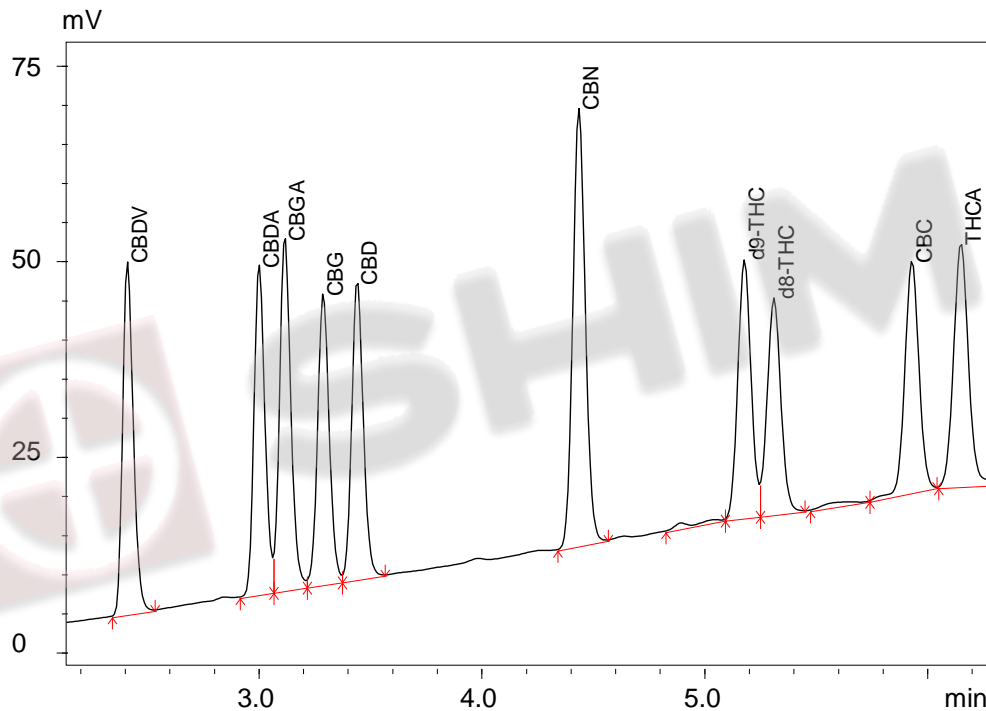


→ **All methods were exhaustively tested for ruggedness, repeatability and quantitative accuracy !**

High Throughput

- **High Throughput HPLC Method Package**

- Designed for analysis of the 10 most commonly requested cannabinoids in under 8 minutes. (Does not include THCv)

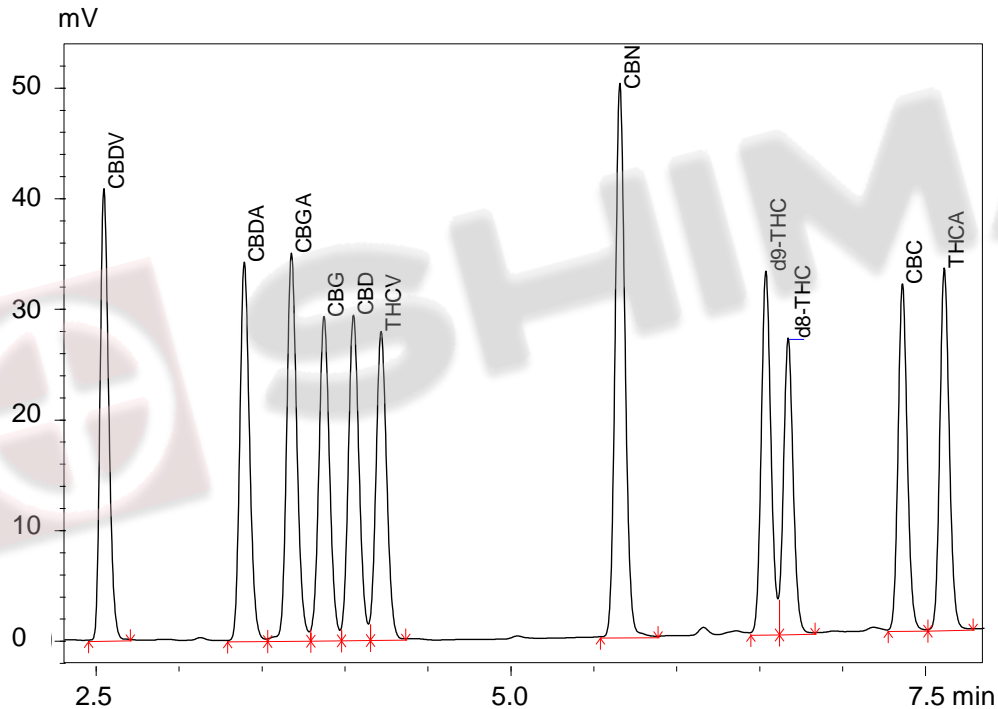


- When high sample throughput is paramount, 60 samples per 8 hr day
- Quantitative for 10 cannabinoids

High Sensitivity

- **High Sensitivity HPLC Method Package**

- Adds THCV to the target analyte list, with an instrument cycle time of under 10 minutes. The short analysis time produces the sharpest chromatographic peaks for the best overall sensitivity.

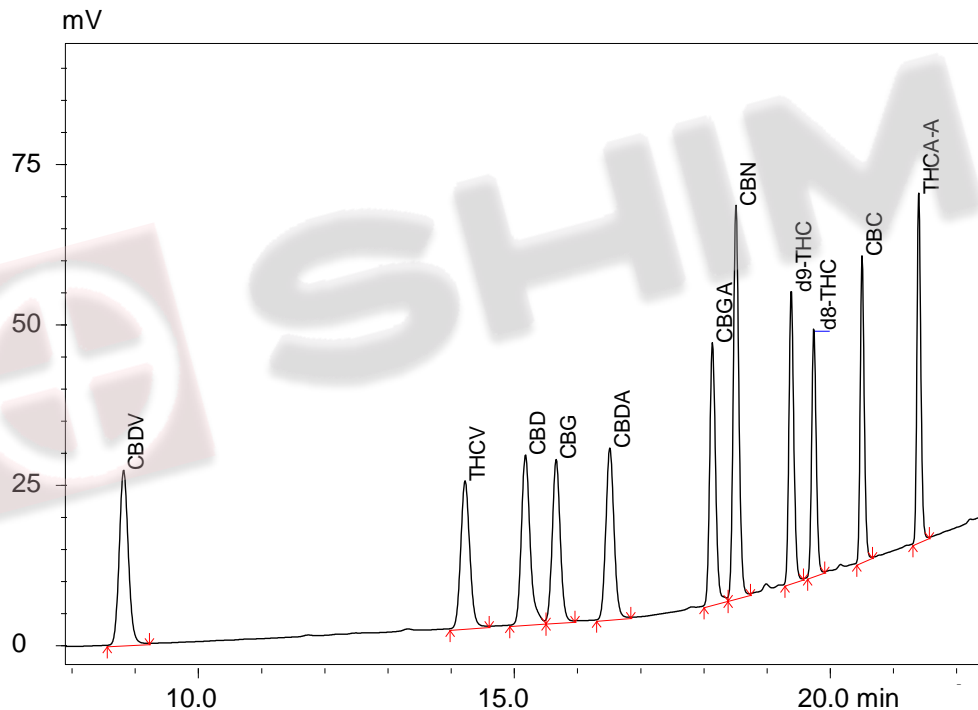


- When THCV is important, yet with good sample throughput, 48 samples per 8 hr day
- Quantitative for 11 cannabinoids

High Resolution

- **High Resolution HPLC Method Package**

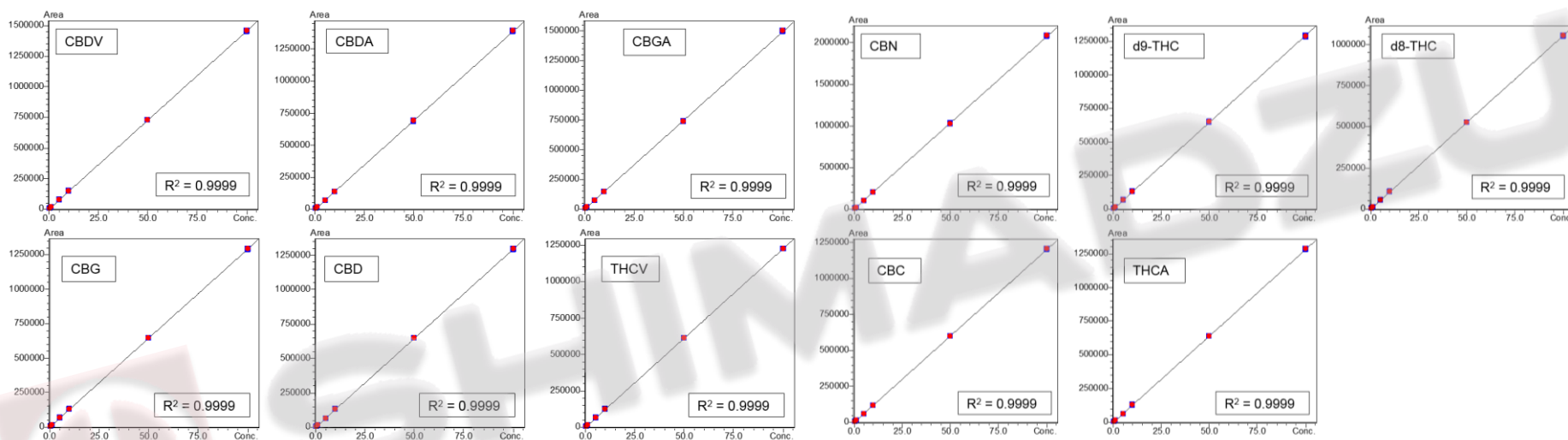
- Full baseline resolution for all 11 compounds and an analysis time under 30 minutes. Preferred method for research purposes, or when additional compounds must be added in response to new regulatory requirements.



- For baseline separation of 11+ components, including THCV.
Minimum Resolution = 2.0
- 16 samples per 8 hr day
- Add cannabinoid targets as regulations change

Quantitative Accuracy

- **Standard Calibration Curves (High Sensitivity Method)**
 - $R^2 > 0.999$ for all targets



Level (ppm)

Accuracy % (Bias)

Range

Low 2

109.7 – 111.1

± 1.4

Med 20

101.2 – 103.4

± 2.2

High 70

99.2 – 100.2

± 1.0

Limits of Detection and Quantitation

- **Determined LOQs and LODs (High Sensitivity Method)**
 - LOQ \leq 0.64 mg/L for all targets
 - LOD \leq 0.21 mg/L for all targets

Target Compound List	Abbreviation	LOQ (mg/L)	LOD (mg/L)
Cannabichromene	CBC	0.60	0.20
Cannabidiol	CBD	0.59	0.20
Cannabidiolic acid	CBDA	0.55	0.18
Cannabidivarin	CBDV	0.43	0.14
Cannabigerol	CBG	0.59	0.20
Cannabigerolic acid	CBGA	0.53	0.18
Cannabinol	CBN	0.37	0.12
d8-Tetrahydrocannabinoid	d8-THC	0.64	0.21
d9-Tetrahydrocannabinoid	d9-THC	0.52	0.17
Tetrahydrocannabivarin	THCV	0.62	0.20

All values correspond to Dry WT % of less than 0.1%

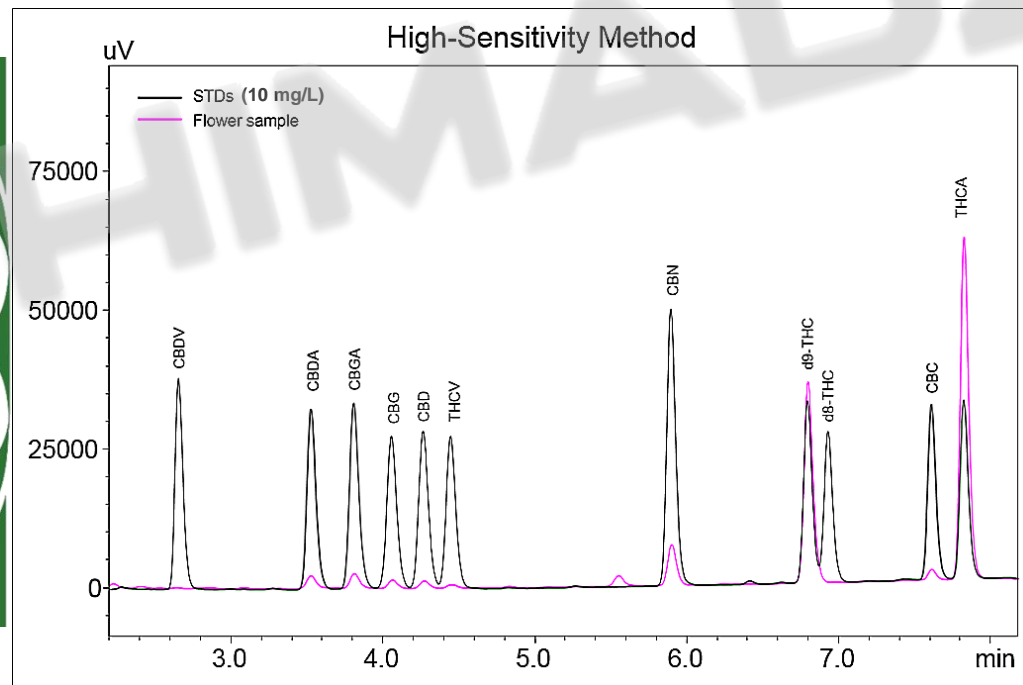
Analytical Results

- **Example of quantification from 200 mg cannabis flower**

→ $\text{DRY WT\%} = [\text{Target}] \times (\text{Dilution}) \times (\text{Extraction Vol./Dry wt mg}) \times 100$

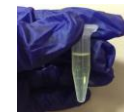
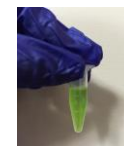
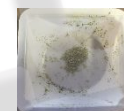
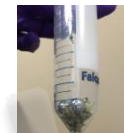
$\text{THCA WT\%} = 3.8 \% = [18.9 \text{ mg/L}] \times (20/1) \times (0.02 \text{ L}/200 \text{ mg}) \times 100$

$\text{d9-THC WT\%} = 2.0 \% = [10.2 \text{ mg/L}] \times (20/1) \times (0.02 \text{ L}/200 \text{ mg}) \times 100$



Sample Preparation of Flowers

- **Prepare extracts from flowers, leaves or stems**
 - Weigh 200 mg of flower or leaf cuttings into 50 mL centrifuge tube
 - Add two 9.5 mm steel balls into the tube
 - Shake at 1000 rpm for 1 min with a Grinder or similar
 - Add 20 mL of methanol to the tube
 - Shake at 1000 rpm for 1 min
 - Wait for 15 min
 - Mix using a vortex mixer for 1 min.
 - Transfer 1 mL of the mixture into a 1.5 mL micro-tube and centrifuge at 3000 rpm for 5 min
 - Transfer 100 μ L of supernatant to a new 1.5 mL micro-tube
 - Add 900 μ L of methanol
 - Filter the mixture through a 0.45 μ m syringe filter and transfer to a 1.5 mL sample vial



Sample Preparation of Oils

- **Prepare extracts from flowers, leaves or stems**
 - Add 400 μL isopropanol to a 2 mL glass vial
 - Add 10 μL hemp oil sample and completely dissolve
 - Agitate for 30 seconds
 - Add 400 μL methanol to the mixture
 - Agitate for 30 seconds
 - Filter the mixture through a 0.2 μm PTFE syringe filter into an HPLC vial
 - (Note: Total dilution factor x 81)



Workflow in the Software (1)

1. Setting samples in the Analysis Screen

Ready

Instrument1

Admin

Startup

Analysis

Monitor

Version Info.

Analysis

Standard + Unknown Unknown Only

Clear

Paste

Standards set up automatically according to the method

Unknowns to be analyzed

Vial	Sample Name	Sample ID	Sample Amount [mg]	Extraction Vol. [mL]	Dilution Factor
1	11 Standards mixture	0.5ppm			1
2	11 Standards mixture	1ppm			1
3	11 Standards mixture	5ppm			1
4	11 Standards mixture	10ppm			1
5	11 Standards mixture	50ppm			1
6	11 Standards mixture	100ppm			1
10	Flower	001	200	20	20
11	Oil	001	200	20	20

Standard Count : 6

Unknown Count : 2

Add Start

Workflow in the Software (2)

2. Monitoring of progress on the Analysis Screen

The screenshot displays the software's monitoring interface, divided into several sections:

- Left Sidebar:** Contains navigation options: 'Running' (selected), 'Instrument1', 'Admin', 'Startup', 'Analysis', and 'Monitor'. A 'Version Info.' button is at the bottom.
- Monitor Section:** Features four 8x5 grids of numbered circles (1-54) representing sample positions. A legend below indicates status colors: Selected (blue outline), Ready (green), Running (cyan), Completed (purple), Duplicated (orange), and Interrupted (red). Grid 1 shows a 'Selected' status at position 26. Grid 2 shows 'Ready' status for positions 1-10. Grids 3 and 4 are currently empty.
- Task Monitor Section:** A window titled 'Task Monitor' with a 'Delete All' button. It lists five analysis tasks:
 - Analysis 02/22/2017 - 001: 09:19 AM to 09:39 AM
 - Analysis 02/22/2017 - 002: 09:19 AM to 09:39 AM
 - Analysis 02/22/2017 - 003: 09:20 AM to 09:40 AM (indicated as 'Running' with a blue triangle)
 - Analysis 02/22/2017 - 004: 09:20 AM to 09:40 AM
 - Analysis 02/22/2017 - 005: 09:20 AM to 09:40 AM
 Each task entry includes a progress bar, a 'Detail' button, and a trash icon.
- Bottom Right:** Contains 'Start' (green) and 'Stop' (red) buttons.

Workflow in the Software (3)

3. Quick access to quantitative reports

The screenshot displays the 'Monitor' section of the software. On the left is a navigation sidebar with options: Running, Instrument1, Admin, Startup, Analysis, Monitor (selected), and Version Info. The main monitor area shows a grid of 54 sample positions (9 rows by 6 columns) with status indicators. A legend below the grid defines the colors: Selected (blue circle), Ready (green circle), Running (cyan circle), Completed (purple circle), Duplicated (orange circle), and Interrupted (red circle). In the grid, samples 1-11 are marked as 'Completed' (purple), and sample 12 is 'Running' (cyan). A red arrow points from the 'Completed' status to a 'Task Monitor' window on the right.

The 'Task Monitor' window displays the following information:

- <Sample Information>**
 - System Administrator: Flower
 - Sample Name: Flower_HighSensitivityMethod.tcd
 - Sample ID: HighSensitivity.tcd
 - Data Filename: ReportTest.tcd
 - Method Filename: ReportTest.tcd
 - Batch Filename: ReportTest.tcd
 - Vial #: 1-2
 - Injection Volume: 200 µL
 - Sample Amount: 200 mg
 - Extraction Vol.: 20 mL
 - Dilution Factor: X6
 - Date Acquired: 2017/02/17 19:05:00
 - Date Processed: 2017/02/17 19:39:46
 - Sample Type: Unknown
 - Acquired by: System Administrator
 - Processed by: System Administrator
- <Chromatogram>**
 - UV 220nm
 - Chromatogram showing peaks for CBDA, CBGA, CBG, CBN, d9-THC, d8-THC, and THCA.
- <Quantitative Results>**

ID#	Name	Ret. Time	Dry weight %	Total THC	Total CBD
1	CBDV	0.00	0.00	5.27	%
2	CBDA	3.430	0.13	52.74	mg/g
3	CBGA	3.723	0.13	0.17	%
4	CBG	3.904	0.07	1.75	mg/g
5	CBD	4.063	0.06		
6	THCV	0.00	0.00		
7	CBN	5.643	0.27		
8	d9-THC	6.509	2.00		
9	d8-THC	6.648	0.11		
10	CBC	7.335	0.11		
11	THCA	7.584	3.74		

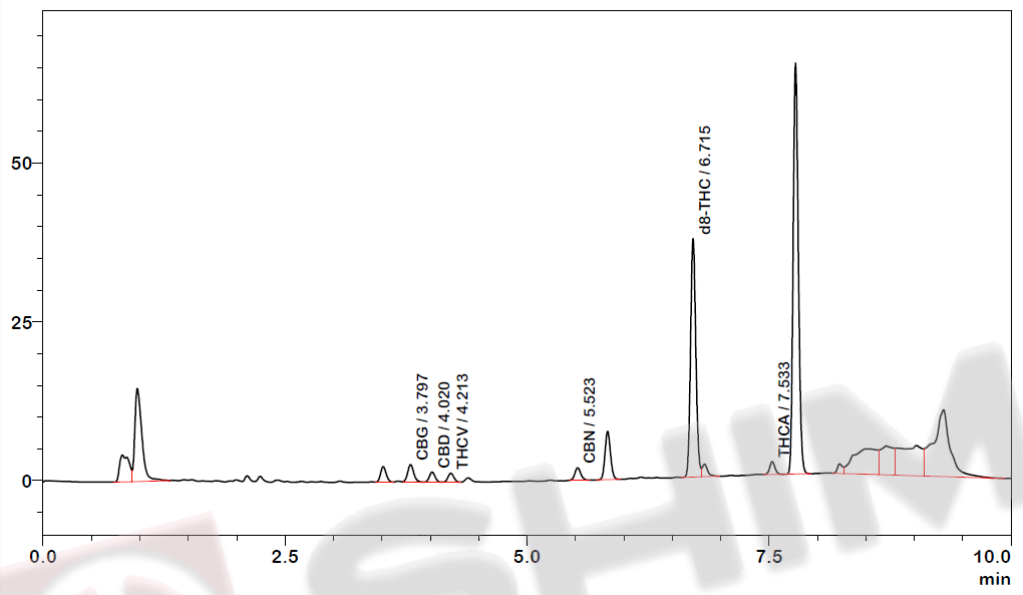
Double-click on completed run to show report

ID#	Name	Ret. Time	Dry weight %
1	CBDV	0.00	0.00
2	CBDA	3.430	0.13
3	CBGA	3.723	0.13
4	CBG	3.904	0.07
5	CBD	4.063	0.06
6	THCV	0.00	0.00
7	CBN	5.643	0.27
8	d9-THC	6.509	2.00
9	d8-THC	6.648	0.11
10	CBC	7.335	0.11
11	THCA	7.584	3.74

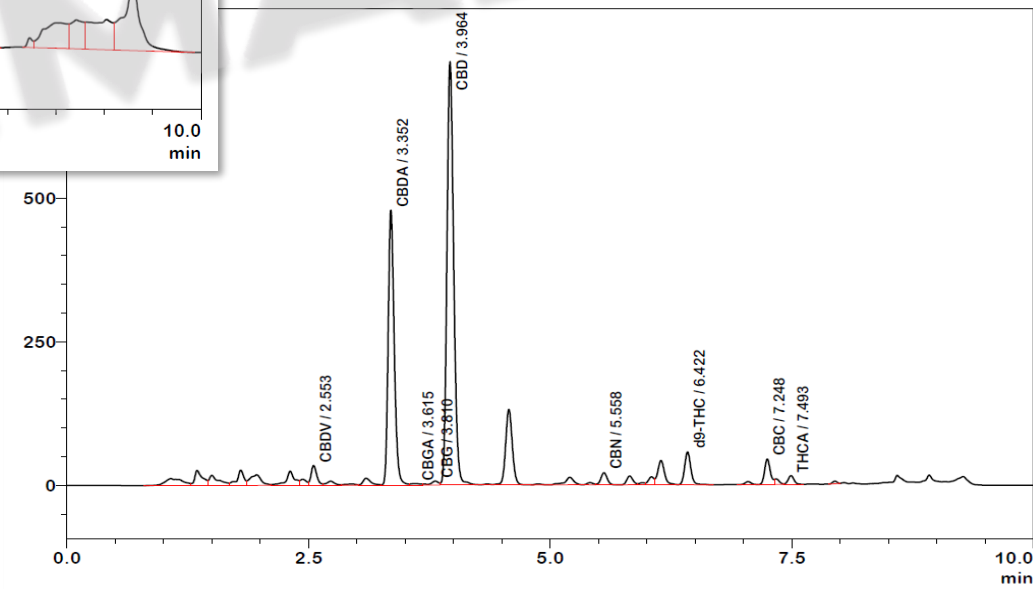
Total THC	5.27	%
Total THC	52.74	mg/g
Total CBD	0.17	%
Total CBD	1.75	mg/g

Examples of Real Samples

Flower Sample



Hemp Oil Sample



Supporting Documentation

Application News

No. HPLC-018

High Performance Liquid Chromatography

The Determination of CBD and General Cannabinoid Content in Hemp Oils Using HPLC with UV Detection

Introduction
Medical marijuana generally possesses high levels of the therapeutic cannabinoid, CBD, and lower levels (generally less than 0.3%) of the psychotropic tetrahydrocannabinol, Δ⁹-THC. Pain mitigation and reduced severity of nausea and seizures are just a few of the therapeutic benefits reported by medical cannabis patients. Little has been done to better understand the chemistry of benefits from CBD. To complicate matters, there is evidence that a combination of CBD, a host of other minor cannabinoids and a complex array of terpenoids may be the most beneficial – called the “entourage effect.” CBD-rich oil has become increasingly popular and is administered via sublingual drops, gel capsules or as a topical ointment.

The main source of CBD-rich oil is industrial hemp. Hemp is considered a rustic plant as it is frost resistant, adapts to poor soil, reproduces easily, and does not require chemical fertilizers/pesticides/herbicides/fungicides to thrive. A hemp crop tends to resist mildew and requires less water than cotton. Hemp textiles are considered softer than cotton.

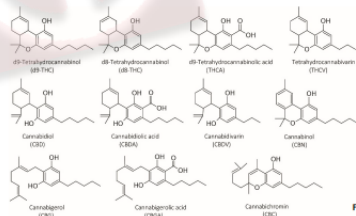


Figure 1: Cannabinoids found in hemp and marijuana

Application News

No. HPLC-020

High Performance Liquid Chromatography

The Potency Determination of 16 Cannabinoids by UHPLC with Diode-Array Detection

Introduction
Since the legalization of cannabis in several US states and, recently, Canada, the quantitative determination of cannabinoids in cannabis products has been of great interest. There are more than 100 cannabinoids that can be found in the plant or extracts¹. Tetrahydrocannabinol (THC) and cannabidiol (CBD) are two of the highest priority in potency testing along with their acidic forms. The acidic forms, Tetrahydrocannabinolic acid (THCA) and cannabidiolic acid (CBDA), are primarily found in the plant, subsequently converting to THC and CBD through decarboxylation from exposure to heat and light².

Traditional HPLC is the gold standard for cannabinoids analysis, including the acidic forms, providing nearly complete separation of the cannabinoids and robust quantitation. Several methods have been developed for optimal results of resolution, sensitivity, and throughput. To assist in optimizing for high throughput while maintaining sensitivity and resolution, this application note proves a 4.5-minute isocratic method using a UHPLC system.

Experimental
Potency analysis was performed using a Shimadzu Nexera® (LC-2040C 3D) UHPLC with a photodiode array detector. The method conditions are shown in Table 1. Historically, 276 nm is ideal for acidic cannabinoids, but non-acidic cannabinoids give weak responses. Consistent with previous literature, a wavelength of 228 nm was chosen as an acceptable compromise³. Experimentation with the PDA supported this finding (Figure 1).

Table 1. Instrument Method Parameters	
Liquid Chromatography	Nexera (LC-2040C 3D)
Mobile Phase A	Water: 5 mM Ammonium Formate, 0.1% Formic Acid
Mobile Phase B	Acetonitrile, 0.1% Formic Acid
UPLC Composition	Isocratic, 25/75
Column	Shimadzu NakedSil CBX II, 1.8 μm, 3.0 × 100mm (Z20-91525-75) Shimadzu NakedSil CBX II Guard, 1.8 μm (Z20-91525-76)
Oven Temperature	30°C
Flow rate	1.0 mL/min
Wavelength Monitored	228 nm

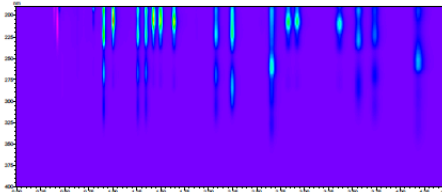


Figure 1: PDA contour plot showing wavelengths 190 to 400 nm

Application News

High Performance Liquid Chromatography, I-series, Cannabinoids

The Determination of Cannabinoids Content within Gummy Based Confectionary

Angela Jain¹
1 Shimadzu UK Ltd, United Kingdom

User Benefits

- Extraction of Cannabinoids from Confectionary Products
- Accurate analysis of Cannabinoids
- Extraction of spiked samples and Commercially available products

Introduction

The growth of the Cannabinoid industry has led to a wide variety of nutraceutical products being commercially available. The leading products within this range are confectionary in nature, typically gummy based sweets. This application looks at the extraction method from the confectionary, utilising Shimadzu's High Sensitivity Cannabinoid method for the analytical analysis.

The extraction method is tested using standard spiking addition techniques using non cannabinoid containing gummy confectionary, as well as using commercially available CBD gummy products. This combination of testing procedures ensured a robust and accurate extraction method was developed for a variety of gummy based confectionary products. The standard spiking tested out the methods precision, accuracy across differing spiking levels and specificity.

Investigations were also carried out to ensure the method is suitable for gummy based confectionary that is suitable for vegans, which does not contain gelatin.

Analytical Conditions

Analysis of all samples was carried out using Shimadzu's High Sensitivity method (HPLC-018), using a Nexera I-series (2060) with photodiode array detector. Reference standards were prepared from individual cannabinoid standards across a concentration range of 0.5 to 90.9 ppm for 11 specified cannabinoids.

Spiking Solutions

Spiking solution (A) for CBD only was prepared at approximately 100 mg/L using an isolate of CBD with known purity in Methanol.

Spiking solution (B) for all cannabinoids was prepared using a known volume of each cannabinoid directly into the sample.

Extraction Sample Preparation

The extraction method used a control gummy confectionary, commercially available without cannabinoids present. These gummies were cut into smaller pieces and thoroughly mixed to form a representative sample.

Spiking with specific quantities of spiking solution A or B was carried out and allowed to absorb into the gummy before proceeding with the extraction path shown in Figure 1.

Figure 1. Extraction Flow Path

Gummy confectionary 2.0 g ± 0.2 g

- Spike if required and leave to absorb
- Add 20 mL water
- Warm in water bath set at 50° C with occasional agitation
- Ensure all Gummy is dissolved
- Add 20 mL acetonitrile
- Agitate for 5 minutes (manually or mechanically)
- Add 8.0 g ± 0.25 g Magnesium Sulfate²
- Agitate for 5 minutes (manually or mechanically)
- Allow sample to settle into distinct layers

- Filter an aliquot (top layer) using 0.2 μm PTFE syringe filter (A)
- Sample A used for minor cannabinoid content
- Dilute by a factor of 4 with methanol for CBD content HPLC Analysis

² - Magnesium Sulfate (MgSO₄) Extra Pure Dried

Extraction Testing

Precision samples were prepared by spiking with spiking solution A for a final concentration of 20 ppm. Six replicates were prepared. Additional precision samples were prepared using spiking solution (B), these were carried out with no final dilution, duplicate samples were prepared.

Commercially available products are sold in a wide range of concentrations and gummy sizes, therefore Accuracy testing was carried out at 5ppm, 20 ppm and 40 ppm, each was carried out in duplicate. Specification testing was carried out to test for peaks present within the gummy/solvent itself that may interfere with cannabinoid determination. Therefore, the following specification criteria were tested:

- Clear only gummy / no spike
- Red only gummy / no spike
- Orange only gummy / no spike
- Yellow only gummy / no spike
- Green only gummy / no spike
- All coloured gummy / no spike
- Random coloured gummy / no spike
- No gummy present / spike
- No gummy present

Disclaimer



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