



Session F6

A Systematic Approach Towards UPLC® Method Development

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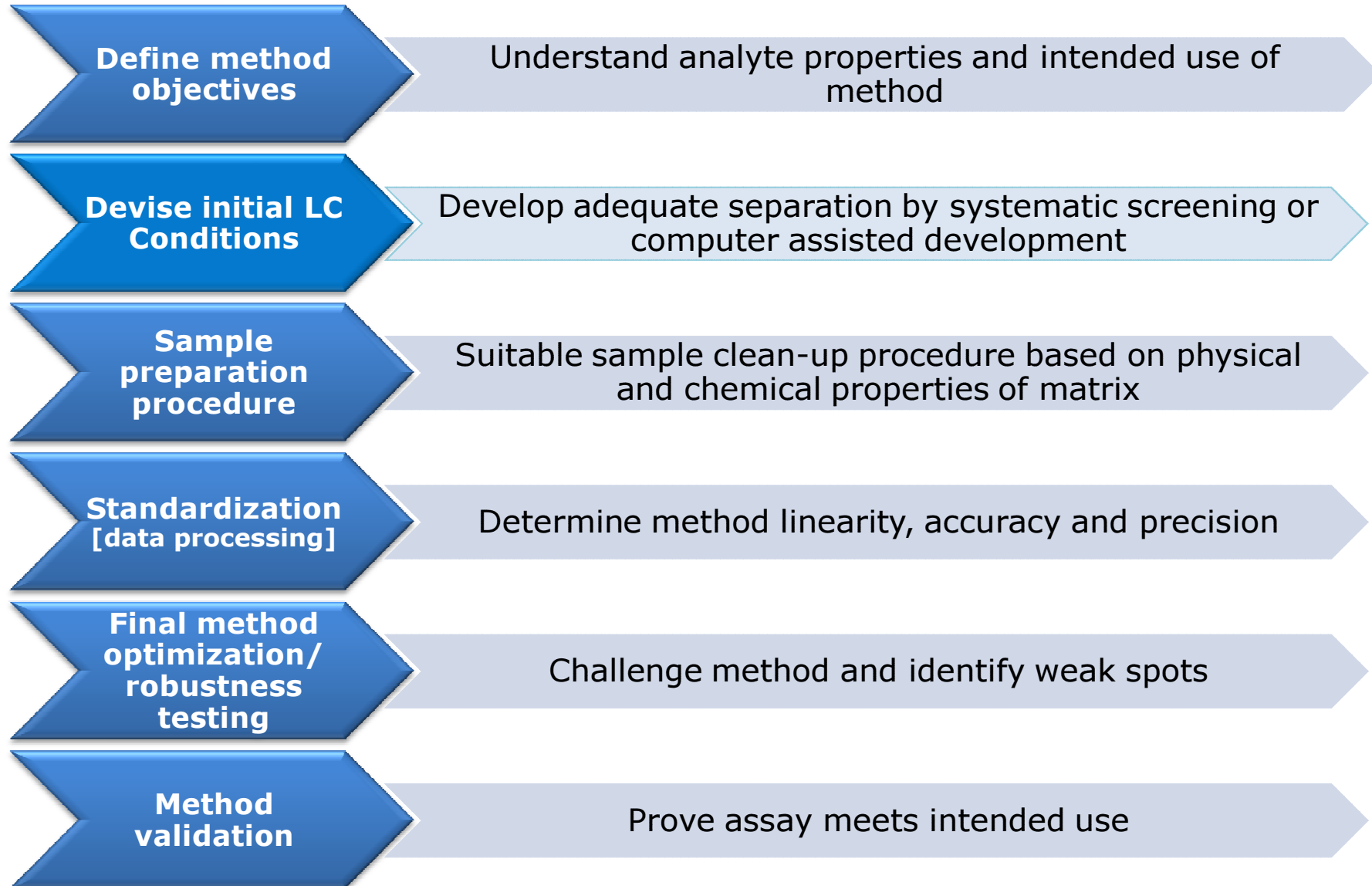
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**Waters Finland
Kutomotie 16
00380 Helsinki**

- Chromatographic methods are developed for different applications constantly, like throughout the drug development process
 - Samples vary in complexity
 - Redundancy exists across an organization
- Method development is costly and time consuming
 - Desire to streamline processes to bring products to market faster
 - Faster chromatographic methods will improve profitability

Critical Components of Method Development



- **Introduction**
 - Approaches Toward Method Development
 - UPLC Technology
 - Success Criteria
- **Controlling Selectivity and Retention**
 - Stationary Phase and Particle Substrate Design
 - Organic Modifier
 - Mobile Phase pH
- **Method Development Strategy**
 - Systematic Screening Protocol
 - Quality by Design [Q_bD] Approach
- **Implementing the Approach**
 - Case Study
- **Conclusion**

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Approaches Toward Method Development: Deriving Initial LC Conditions

- Match LC conditions to the chemical properties of the analyte[s]
 - Educated guess based on past experience [speculation]
 - Usually supplemented with a literature search
 - Ask a colleague

- Stepwise incremental approach
 - Next step experimental design based on results from previous experiment

- ★ Systematic screening protocol
 - Evaluate combinations of mobile phase pH, organic modifier and stationary phase
 - Select best combination of these parameters
 - Method optimization
 - Gradient slope/Temperature

UPLC Technology enables faster method development



ACQUITY UPLC H-Class

Develop methods in a single work day!

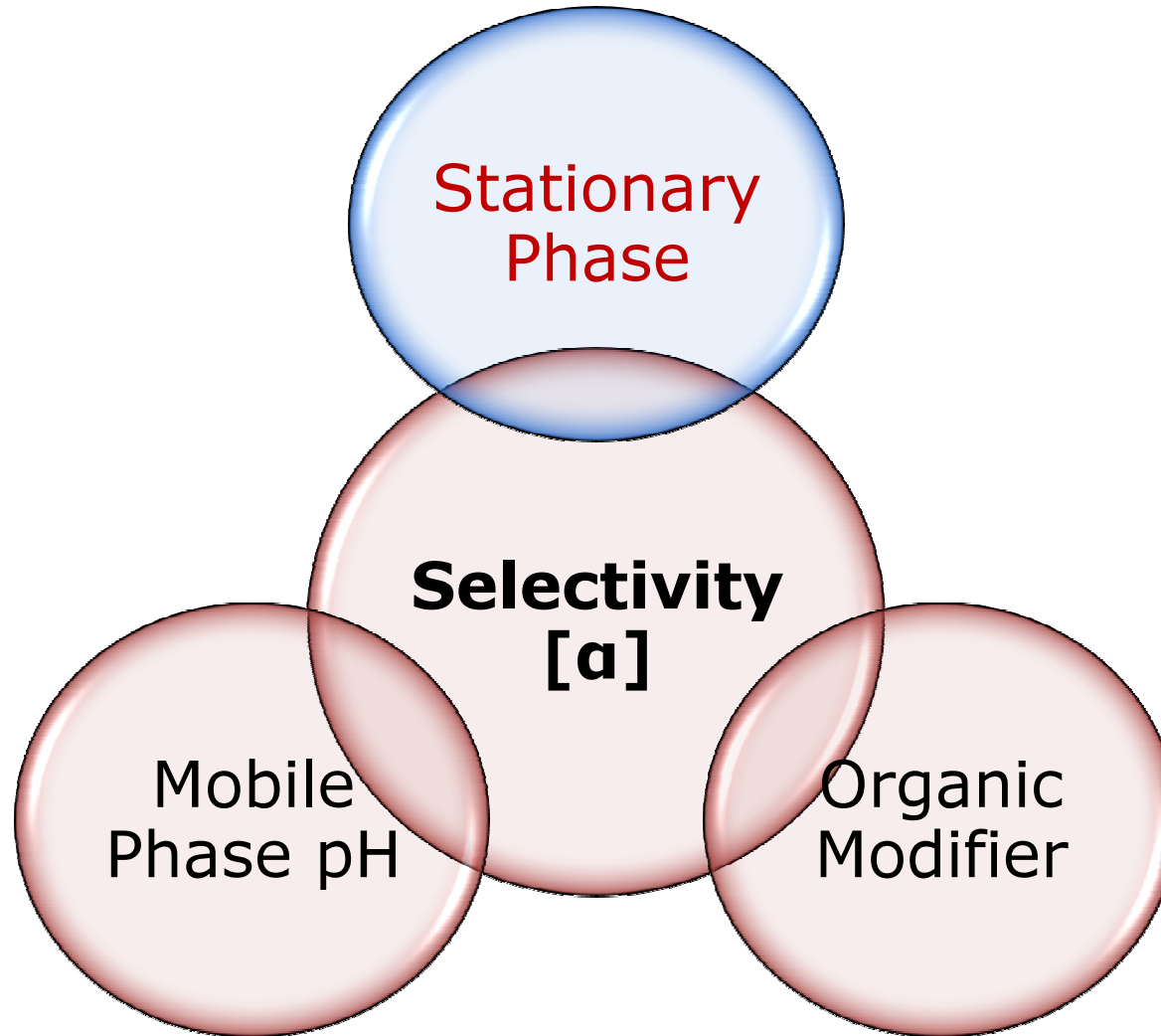
- Systematic screening protocol involving pH, organic modifier and column chemistry
- High resolution sub 2 μm column technology creates high resolution separations, faster
- Automated column and mobile phase selection
- Quaternary solvent mixing [ACQUITY UPLC H-Class]

Before You Start: Information Gathering

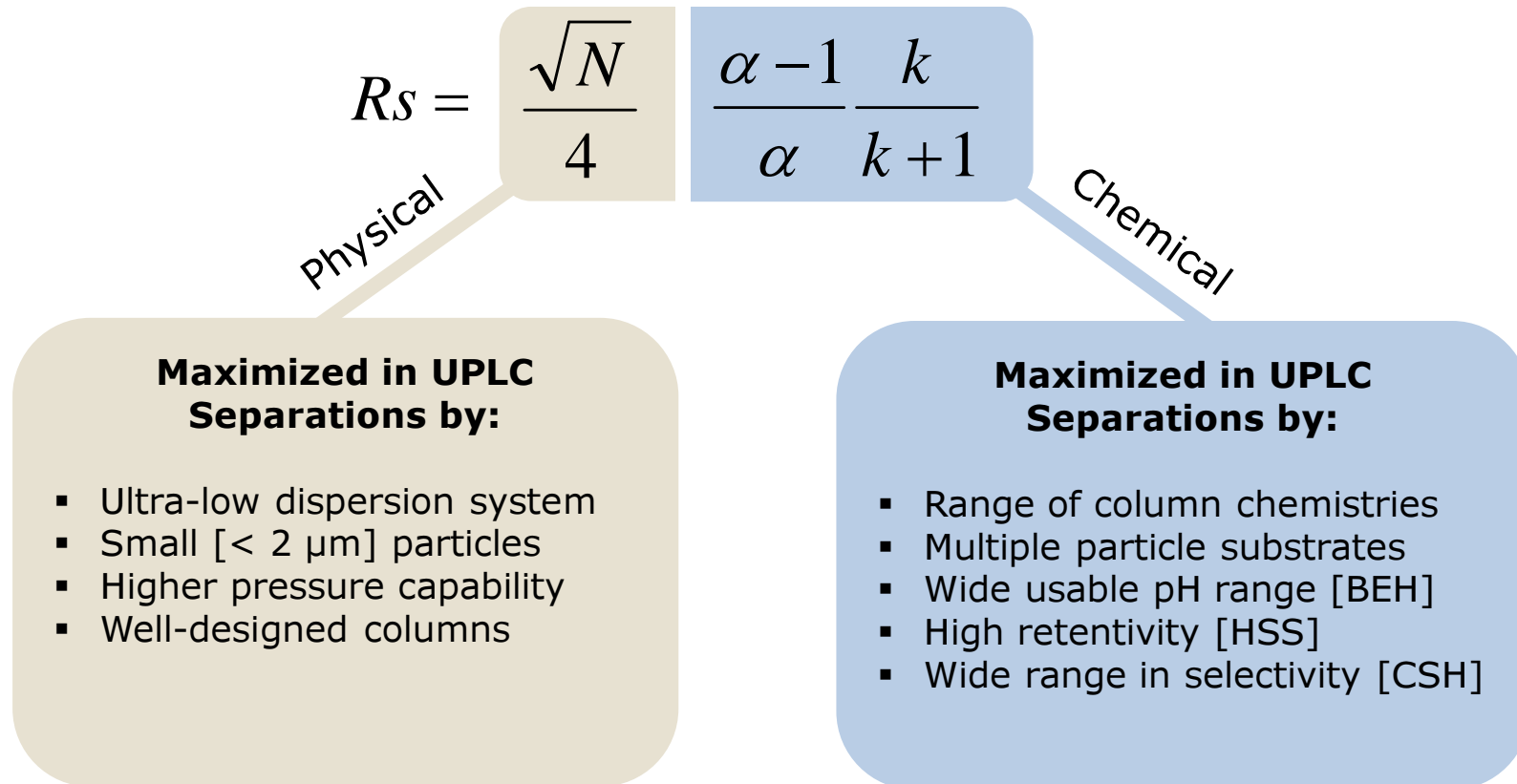
- Chemical properties [functional groups]
 - Ionizable species, polarity, pKa, molecular weight
- Sample solubility
- Number of compounds present
 - How many components are you trying to separate?
- Sample matrix
- Detection technique [UV, ELS, RI, FL, MS etc.]
 - Based on available equipment or sensitivity requirements of assay
- Criteria for success
 - Concentration range and quantitative requirements
 - System suitability

- **Introduction**
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Chemical Factors that Impact Selectivity



Improving Resolution with Complementary Selectivity



Impact on Resolution % Improvement

Double N	20 – 40%
Double k	15 – 20%
Double α	> 400%

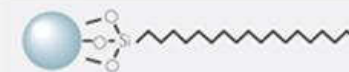
Stationary Phase Selectivity: Bonded-Phase [Ligand] and Particle Substrate

- Silanol activity and surface charge
 - Influences secondary interactions [ion-exchange], peak shape and sample loadability
- Hydrophobicity
 - Longer alkyl chain lengths will provide increased retention
 - Shorter, ionizable ligands will increase polarity
- Hydrolytic stability
 - Column lifetime will be impacted by the number of attachment points to the particle surface
- Ligand density
 - Influences retention and sample loadability

The Widest UPLC Column Offering

- **Five particle substrates**
 - 130Å, 200Å and 300Å BEH [Ethylene Bridged Hybrid], HSS [High Strength Silica] and **CSH [Charged Surface Hybrid]**
 - All are available in HPLC and UPLC particle sizes
- **Wide and growing selection of column chemistries**
 - BEH 130Å C₁₈, C₈, Shield RP₁₈, Phenyl, HILIC and Amide
 - BEH 300Å C₁₈ and C₄
 - BEH 200Å SEC
 - HSS C₁₈, T3, C₁₈ SB (and soon Cyano and PFP)
 - **CSH C₁₈, Fluoro-Phenyl and Phenyl-Hexyl**
- **Proven application-based solutions**
 - AAA, OST, PST, PrST and Glycan
- **Transferability between HPLC and UPLC**
 - XBridge HPLC and ACQUITY UPLC BEH columns
 - HSS HPLC and ACQUITY UPLC HSS columns
 - **XSelect HPLC and ACQUITY CSH columns**
- **VanGuard Pre-columns**
- **eCord Technology**

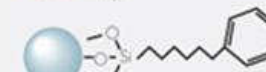
BEH C₁₈



BEH C₈



BEH Phenyl



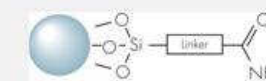
BEH Shield RP18



BEH HILIC



BEH Amide



HSS T3



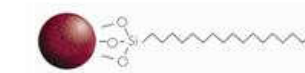
HSS C₁₈



HSS C₁₈ SB



CSH C₁₈



CSH Fluoro-Phenyl



CSH Phenyl-Hexyl



Industry Trends: Current State of Reversed-Phase Separations

- Advances in stationary phase design
 - Hybrid particle technology
 - Extended usable pH range [1-12]
 - Exceptional peak shape and efficiency
 - Rugged and reliable column life
 - Sub 2 μm particle technology
 - Improvements in resolution, sensitivity and speed of analysis
 - Pellicular [core-shell] particles

- Instrument platform of choice
 - UltraPerformance LC with UV and mass spectrometry [UPLC/MS/[MS]]
 - Requires volatile mobile phases
 - Excludes typical UV-based buffers [i.e., phosphate buffers]
 - Preference towards low ionic strength additives [i.e., formic acid, acetic acid, ammonium hydroxide]
 - Avoid preparation of buffers if possible

Defining the Problem: Low Ionic Strength Mobile Phases

- Poor mass loading of charged cationic [basic] solutes in low pH mobile phases due to limited sample capacity
 - High tailing factors
 - Poor signal intensity

- Slow equilibration at low pH
 - Drifting retention times with repeat injections

- Elution [retention] time shift after exposure to a higher pH mobile phase*¹
 - Irreproducible assay performance when performing method screening
 - Low/high pH switching with un-buffered mobile phases

*¹ Marchand, D.H., et al., J. Chromatogr. A **2003**, 1015, 53-64

Explanation of CSH Technology

Step 1



Unbonded BEH Particle

Start with the rugged, ultra-efficient, ethylene bridged hybrid (BEH) particle

Step 2



Apply Controlled Surface Charge

Add reproducible low-level charge to particle surface

Step 3

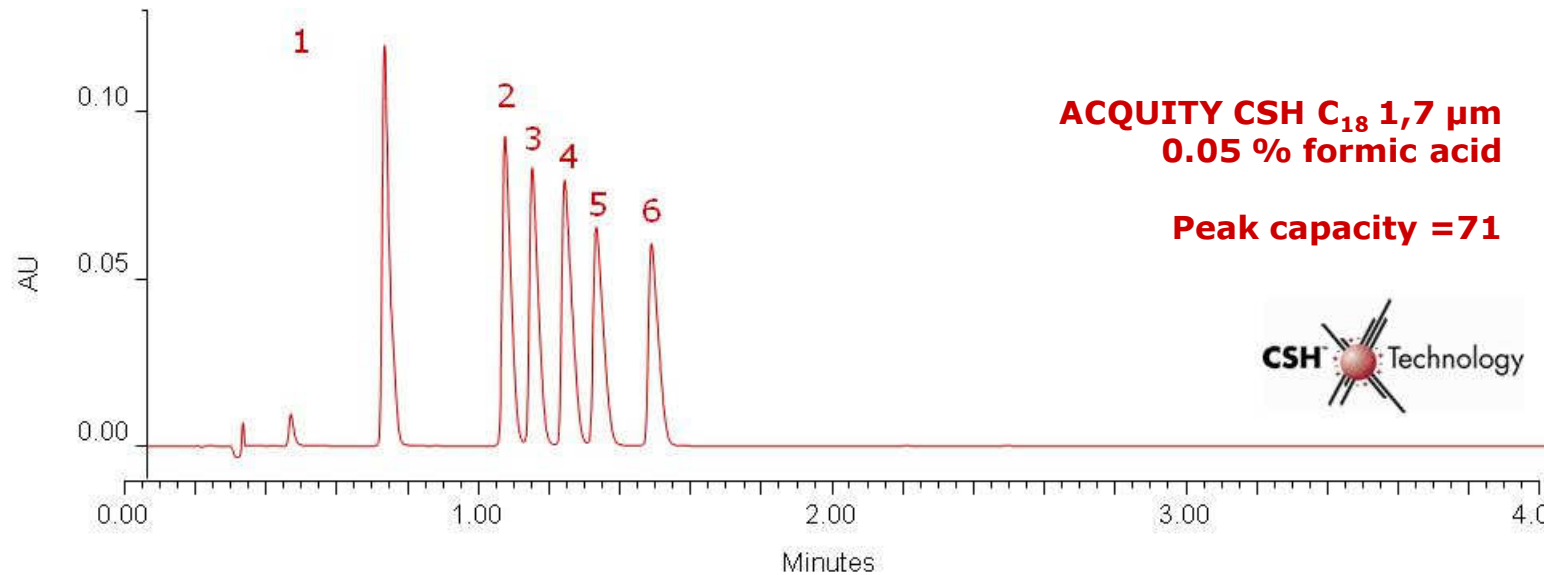


Bond and End Cap

Functionalize with appropriate bonded phase chemistry

CSH Technology: Controlled Surface Charge Yields High Performance

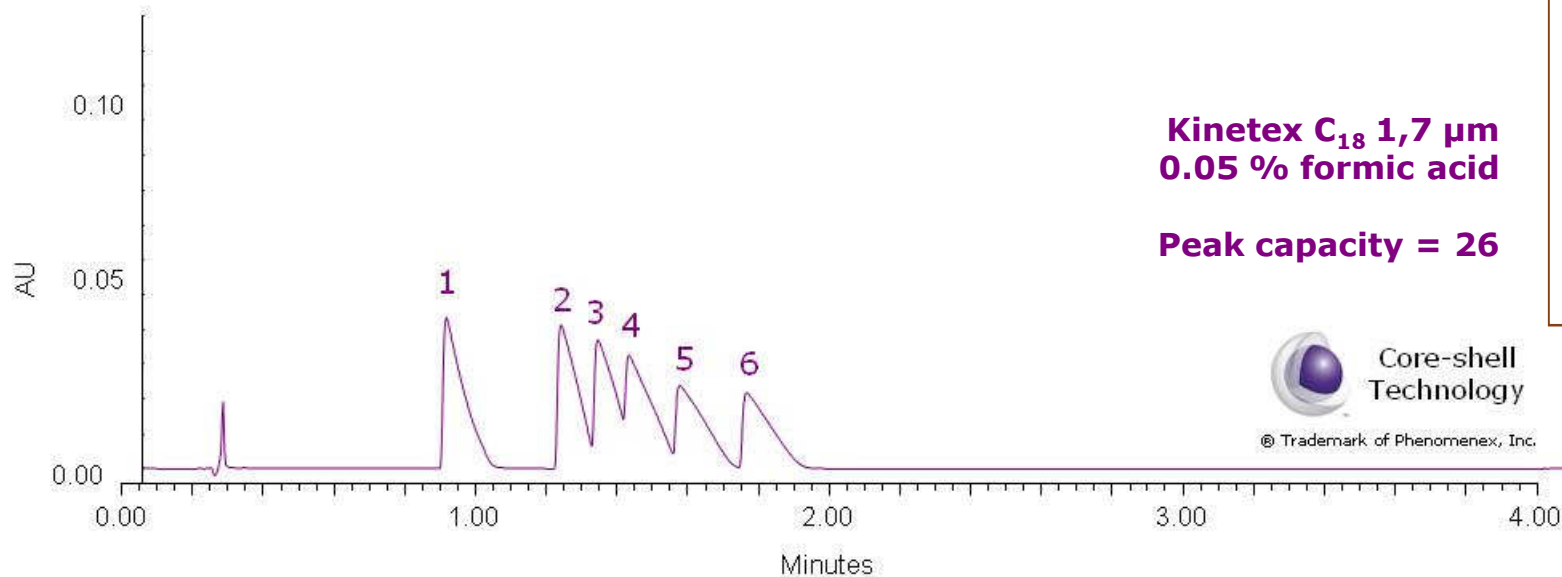
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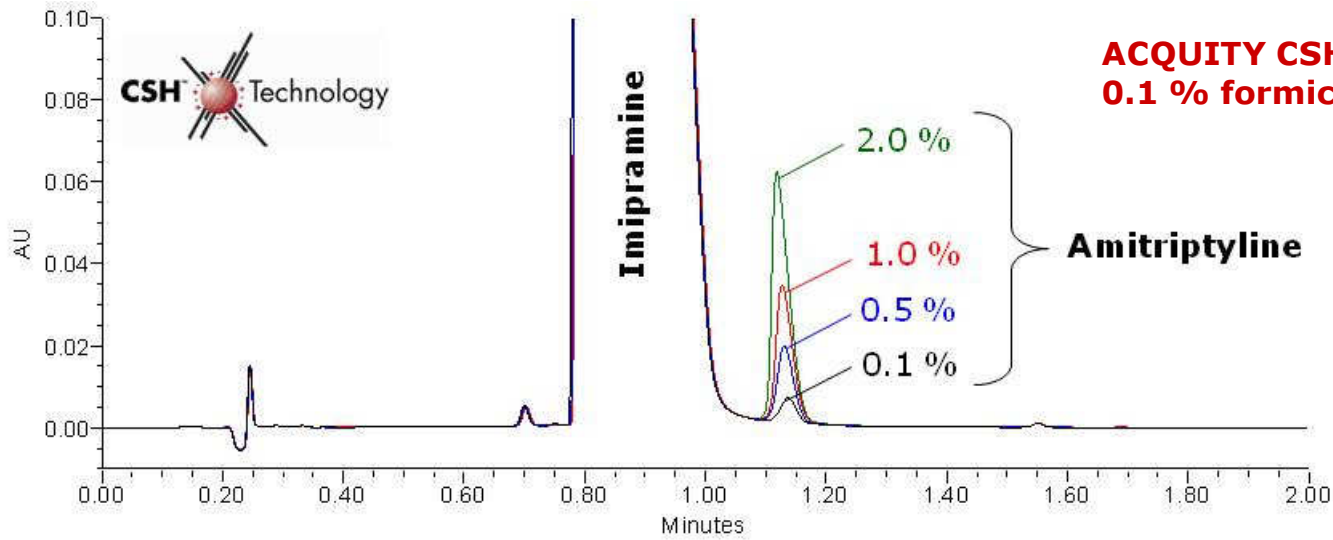
Columns: 2,1 x 50 mm
MP A: water
MP B: acetonitrile
MP C: 2% formic acid
Gradient: 25-35%B in 2 min, 35 – 95%B from 2 - 3 min; [2.5% C held constant]
Inj. Vol.: 5 μl
Sample Conc.: 10 μg/ml
Detection UV @ 254 nm
Sampling rate: 20 Hz
Filter: 0.1 sec
System: ACQUITY UPLC H-Class with ACQUITY UPLC PDA and SQD

Tricyclic antidepressants:

1. Doxepin
2. Desipramine
3. Imipramine
4. Nortriptyline
5. Amitriptyline
6. Trimipramine



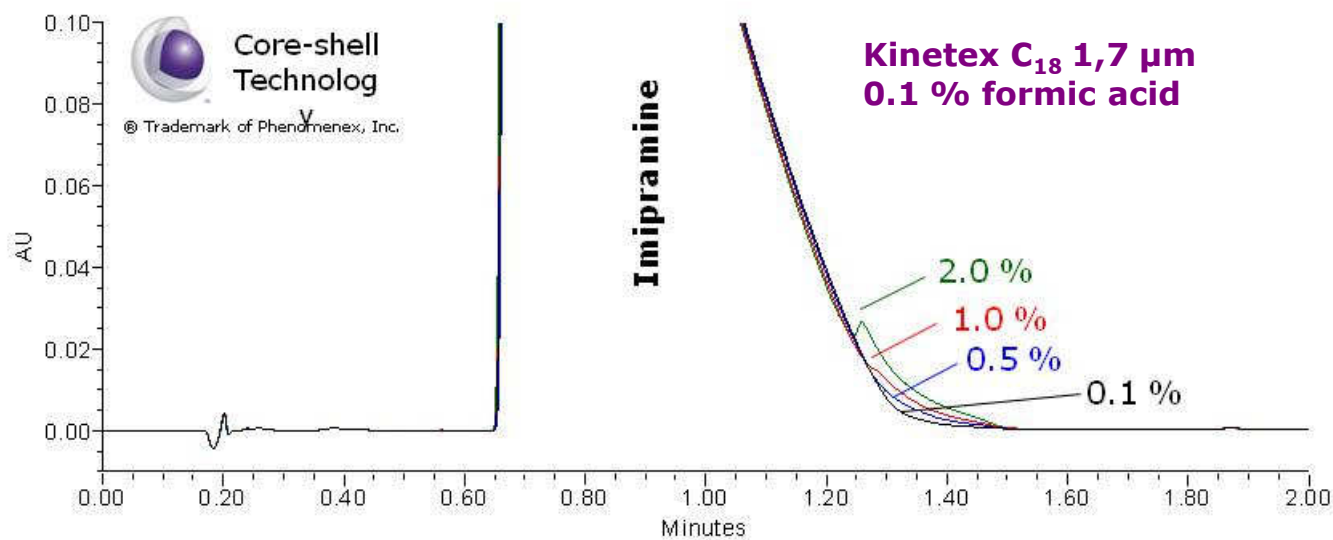
CSH Technology: Influence of Sample Loading on Trace Impurity Detection



Observations

CSH Technology enables superior peak shape and efficiency in low ionic strength mobile phases

Improved sensitivity for trace level impurity analysis



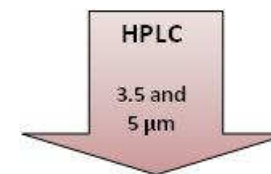
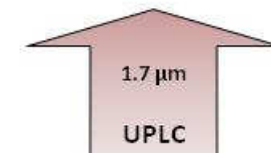
**Imipramine concentration held constant at 0.5 mg/ml;
0.1% formic acid mobile phase**

CSH [Charged Surface Hybrid] Chemistries of UPLC Technology

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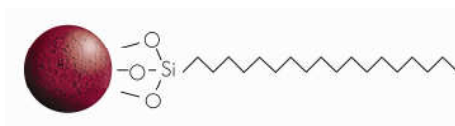
- **CSH C₁₈**
 - Trifunctionally bonded C₁₈
 - Wide pH range for maximum selectivity [pH 1 - 11]
 - Superior peak shape and efficiency in buffered and low ionic strength mobile phases
- **CSH Phenyl-Hexyl**
 - Trifunctionally bonded C₆-Phenyl
 - Wide pH range [1 - 11]
 - Complementary selectivity for aromatic species
- **CSH Fluoro-Phenyl**
 - Trifunctionally bonded, non-encapped, pentafluorophenyl [pH 1 - 8]
 - Unique selectivity compared to alkyl columns
 - Stable and reproducible manufacturing process

Acquity
UPLC®



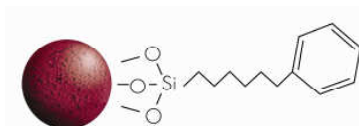
XSELECT™
Columns

ACQUITY UPLC Column Selection: Systematic Screening



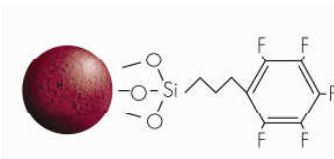
■ CSH C₁₈

- Wide pH range for maximum selectivity [pH 1 - 11]
- Superior peak shape and efficiency in buffered and low ionic strength mobile phases



■ CSH Phenyl-Hexyl

- Trifunctionally bonded C₆-Phenyl [pH 1 - 11]
- Complementary selectivity for aromatic species



■ CSH Fluoro-Phenyl

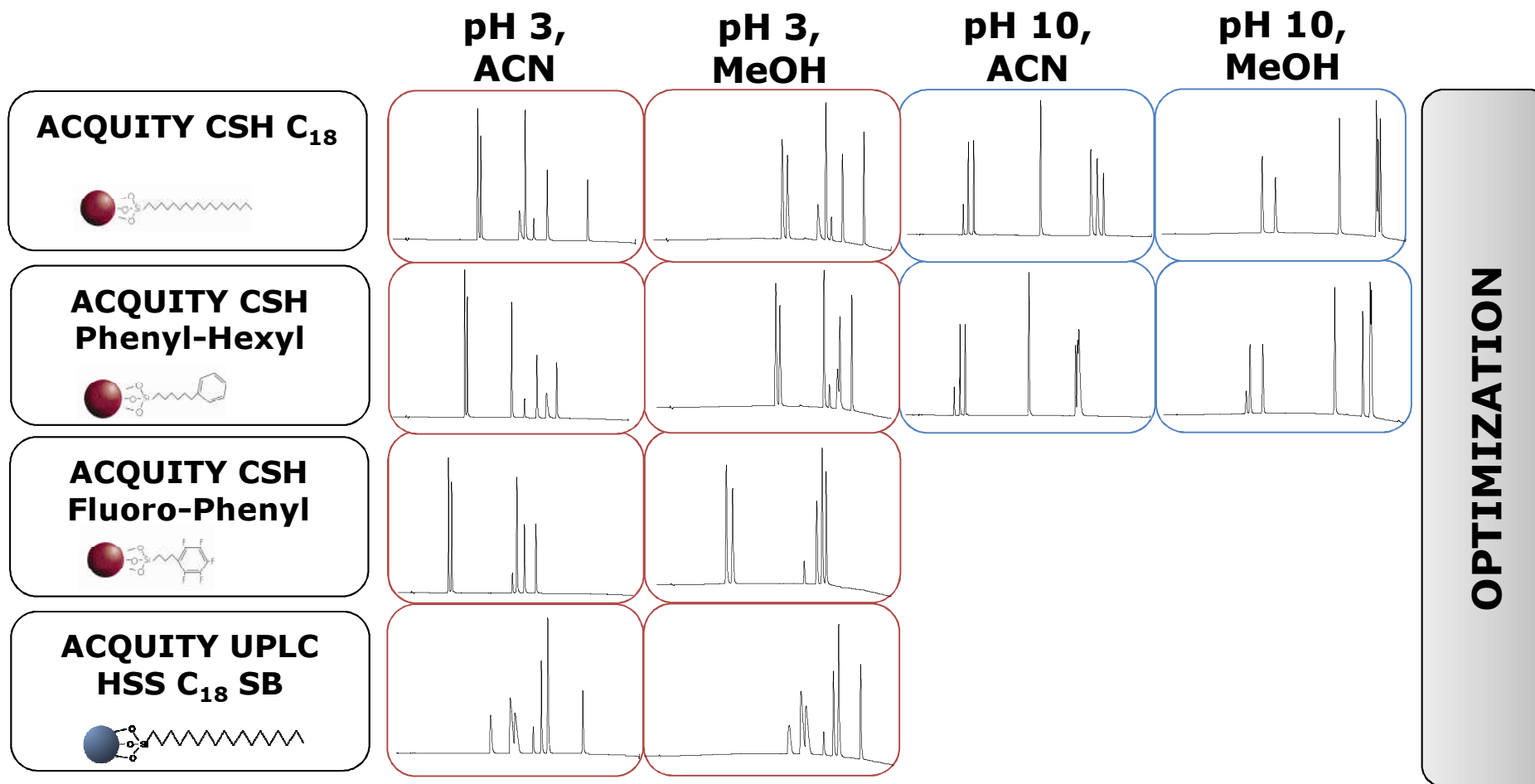
- Trifunctionally bonded pentafluorophenyl, non-endcapped [pH 1 - 8]
- Unique selectivity compared to alkyl columns



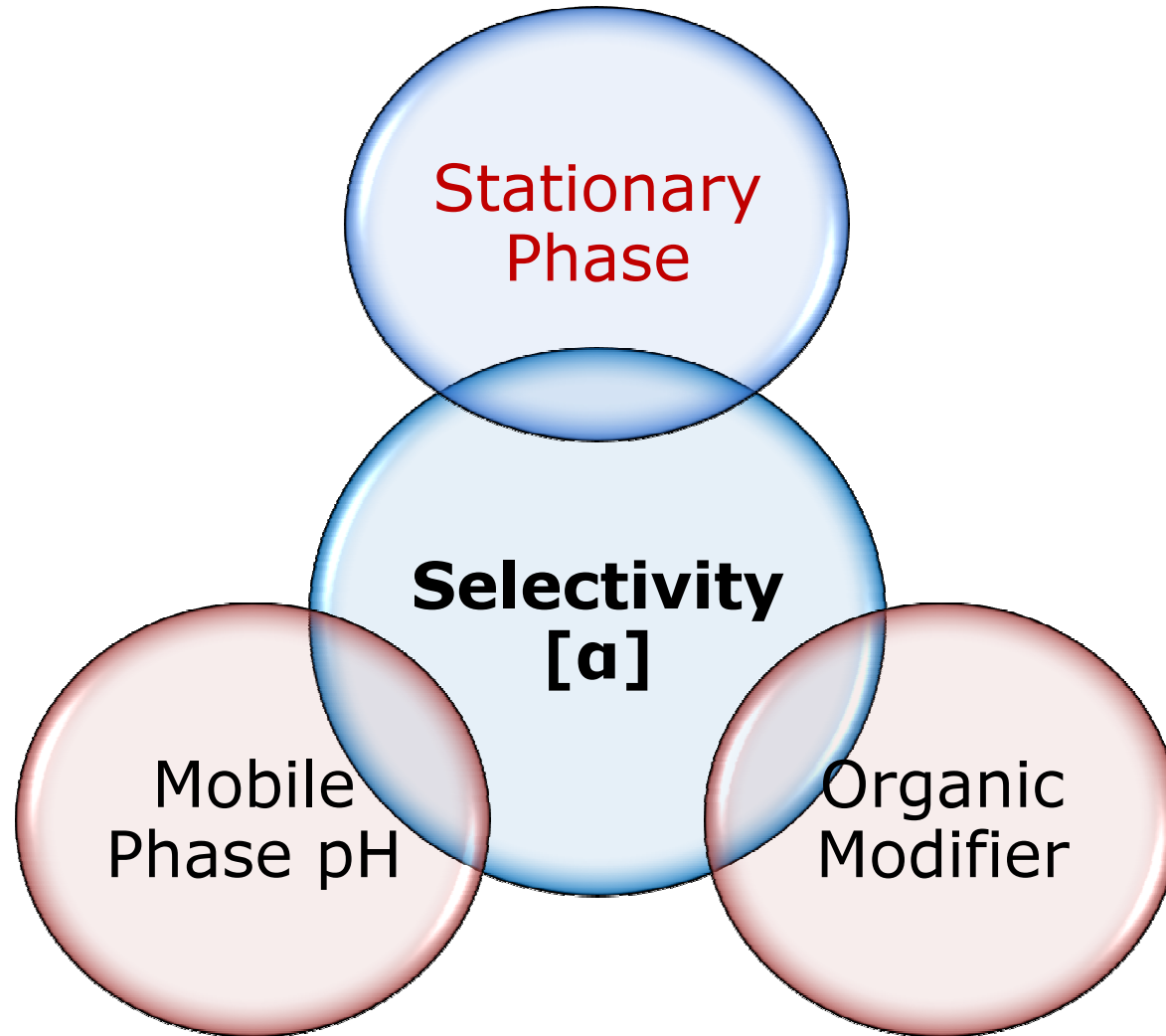
■ HSS C₁₈ SB [Selectivity for Bases]

- Low ligand density, trifunctionally bonded C₁₈ [pH 2 - 8]
- Non-endcapped C₁₈ designed for silanophilic interactions and alternate selectivity with exceptional peak shape for bases

Systematic Screening Protocol

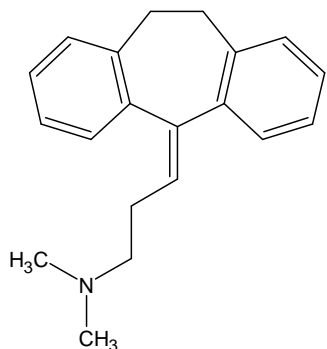


Chemical Factors that Impact Selectivity

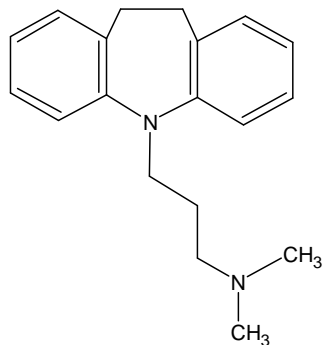


Demonstrating Selectivity: Chemical Structures

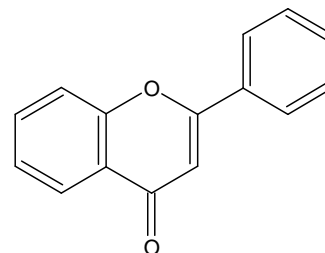
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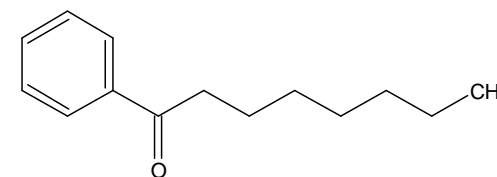
Amitriptyline [B]
m.w. 277.40



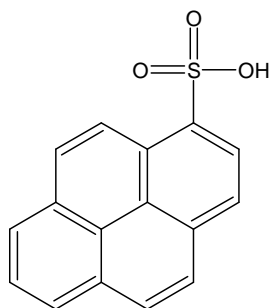
Imipramine [B]
m.w. 280.40



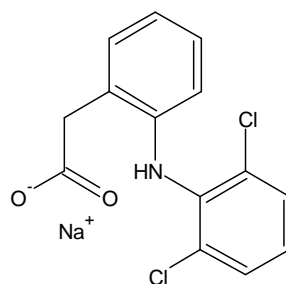
Flavone [N]
m.w. 222.24



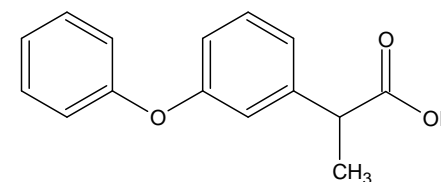
Octanophenone [N]
m.w. 204.31



1-pyrenesulfonic acid [A]
m.w. 304.3



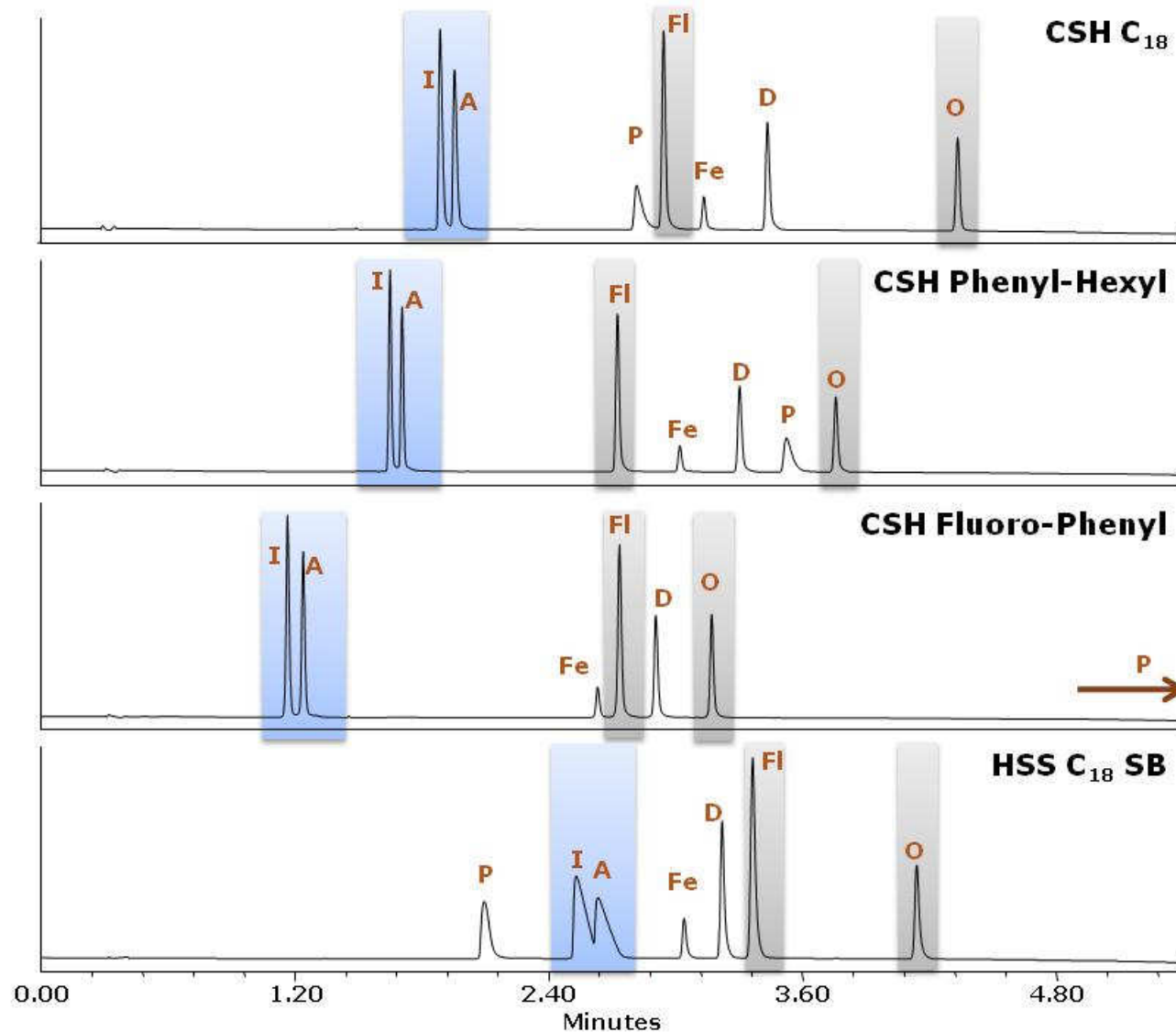
Diclofenac [A]
m.w. 318.13



Fenopropfen [A]
m.w. 242.27

Stationary Phase Selectivity: Basic and Neutral Compounds

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Acetonitrile, pH 3.0

Test Probes:

I: Imipramine [B]

A: Amitriptyline [B]

Fl: Flavone [N]

O: Octanophenone [N]

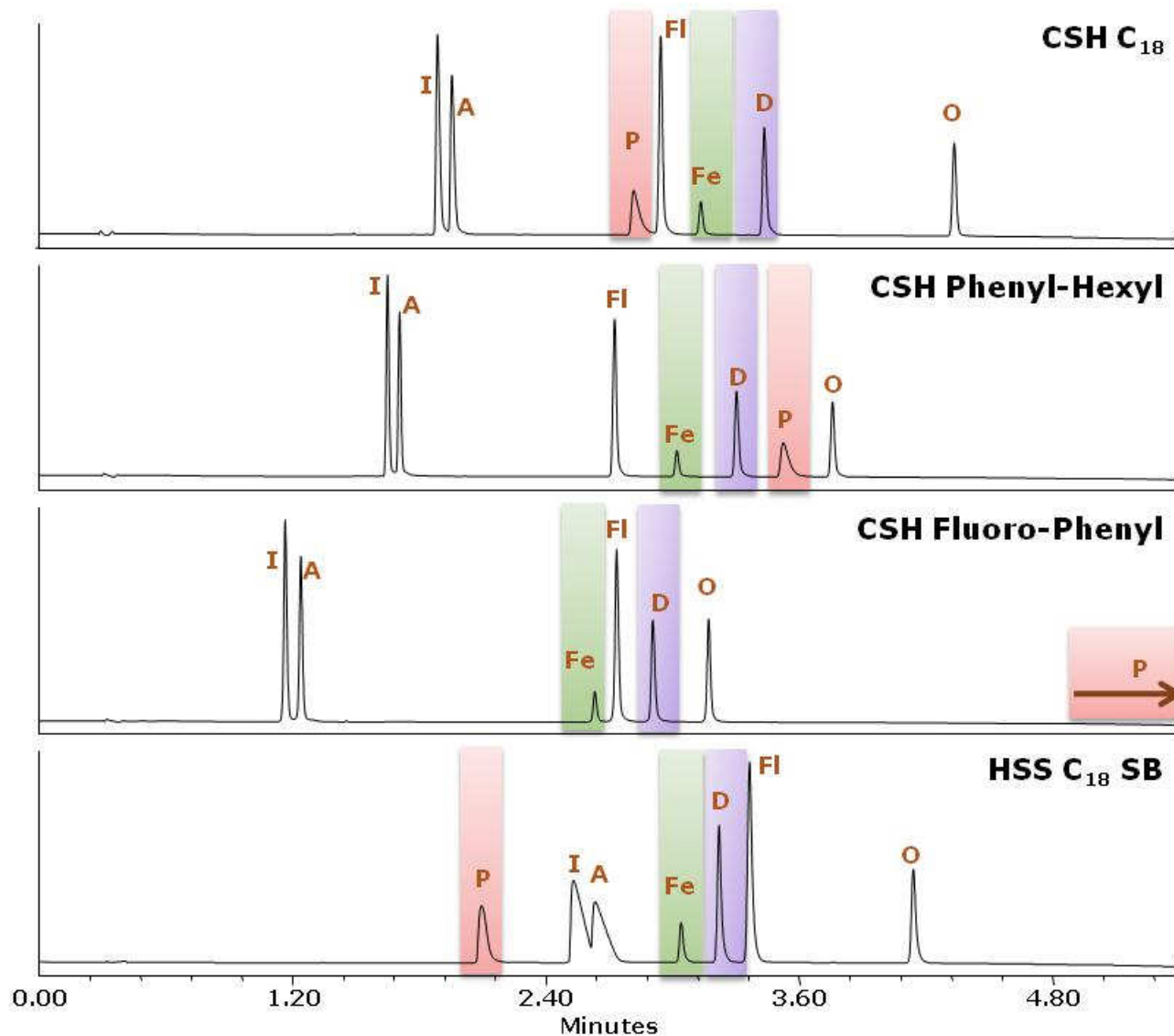
Observations

Similar selectivity α
between the bases

Large differences in
stationary phase
selectivity for overall
mixture

Stationary Phase Selectivity: Acidic Compounds

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**Acetonitrile,
pH 3.0**

Test Probes:

P: 1-pyrenesulfonic
acid [A]

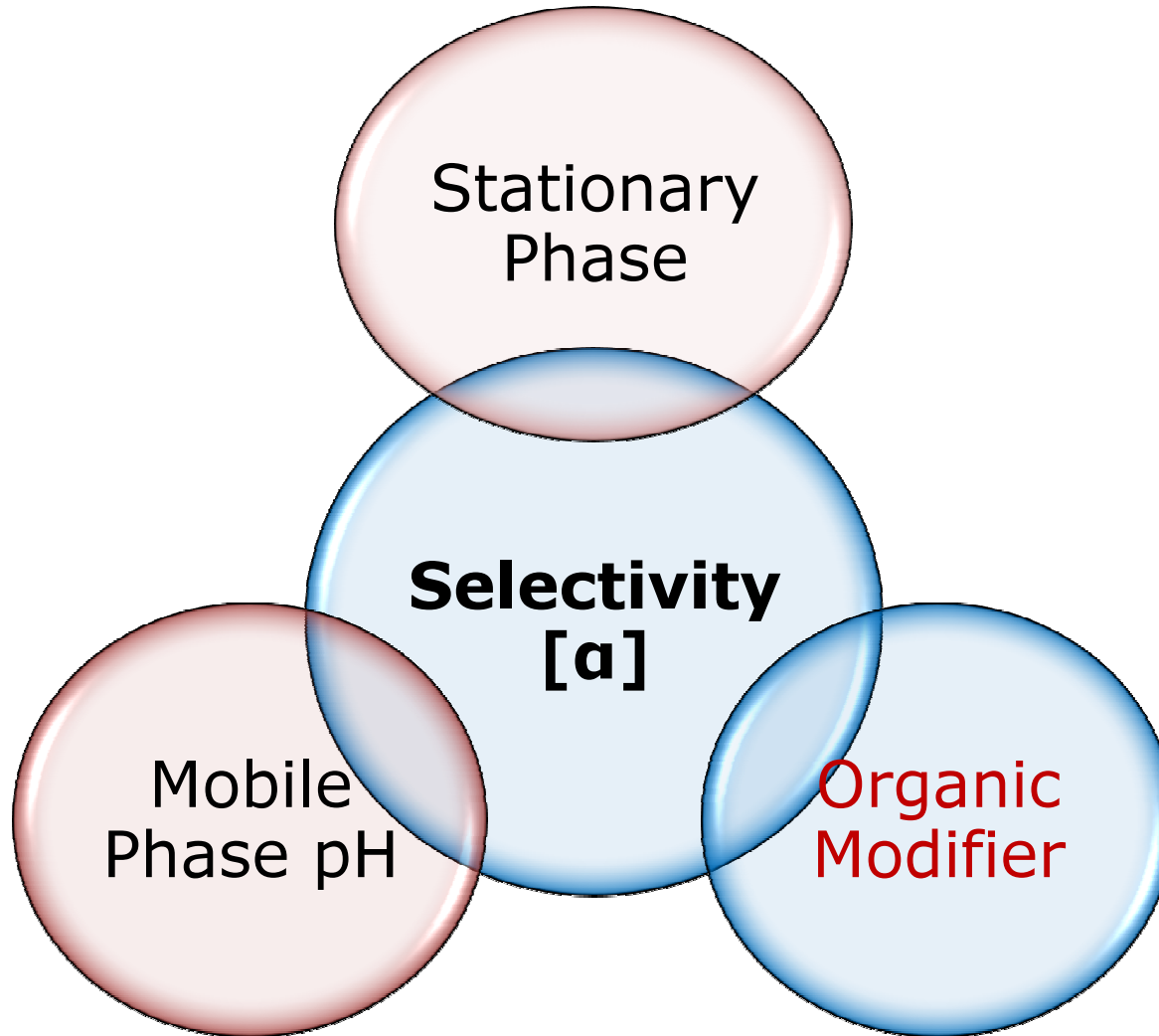
Fe: fenopropfen [A]

D: diclofenac [A]

Observations

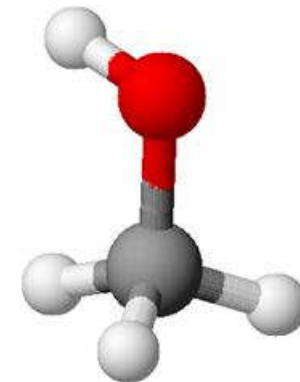
Large differences in
stationary phase
selectivity for acids

Chemical Factors that Impact Selectivity



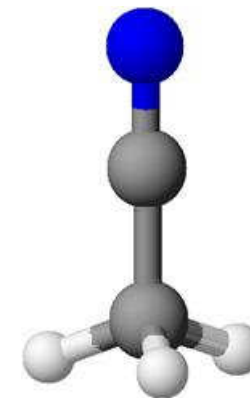
■ Methanol

- Protic solvent [hydrogen bond donor]
- Weak elution solvent [compared to acetonitrile]
- Higher viscosity than acetonitrile



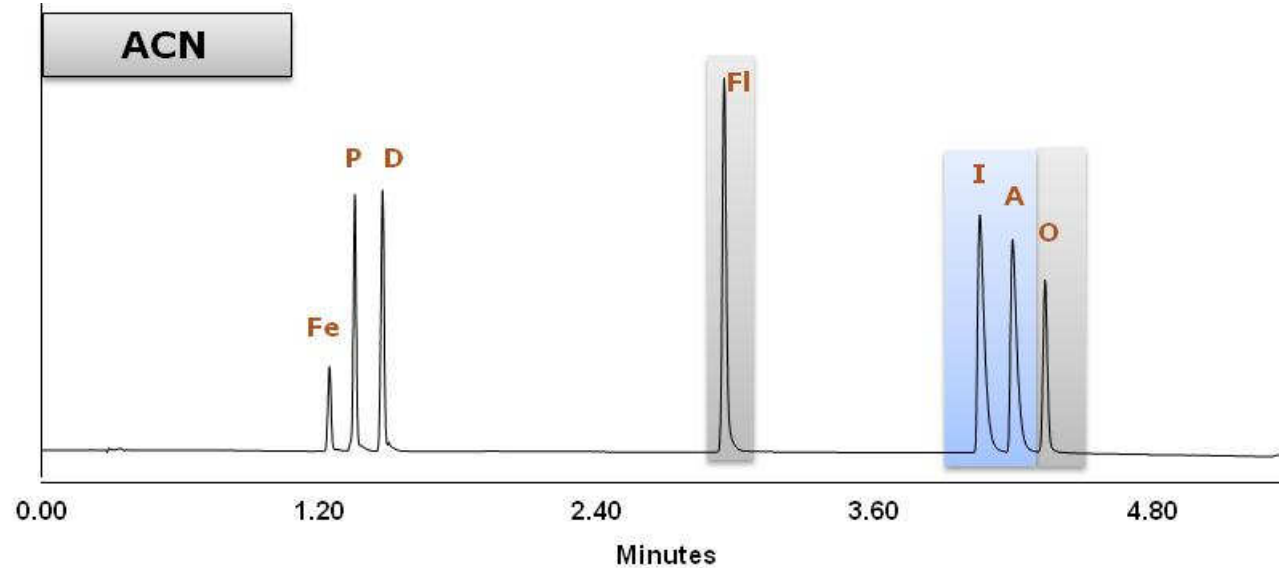
■ Acetonitrile

- Aprotic solvent [hydrogen bond acceptor]
- Strong elution solvent [compared to methanol]
- Low viscosity



Solvent Selectivity: Basic and Neutral Compounds

ACQUITY CSH C₁₈, pH 10



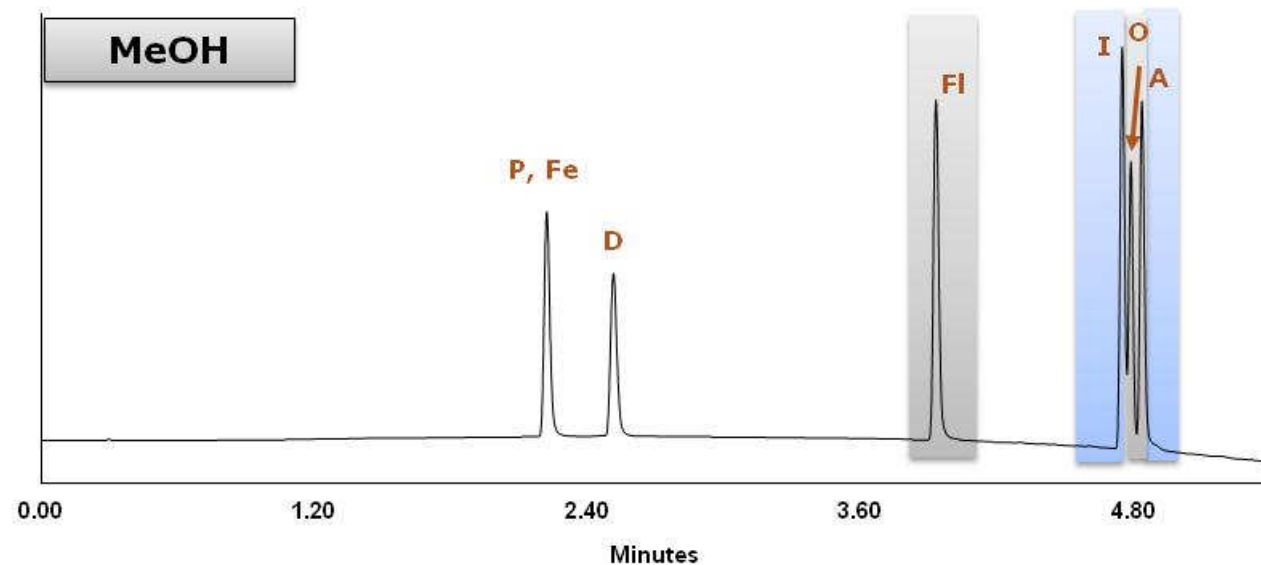
Test Probes:

I: Imipramine [B]

A: Amitriptyline [B]

FI: Flavone [N]

O: Octanophenone [N]



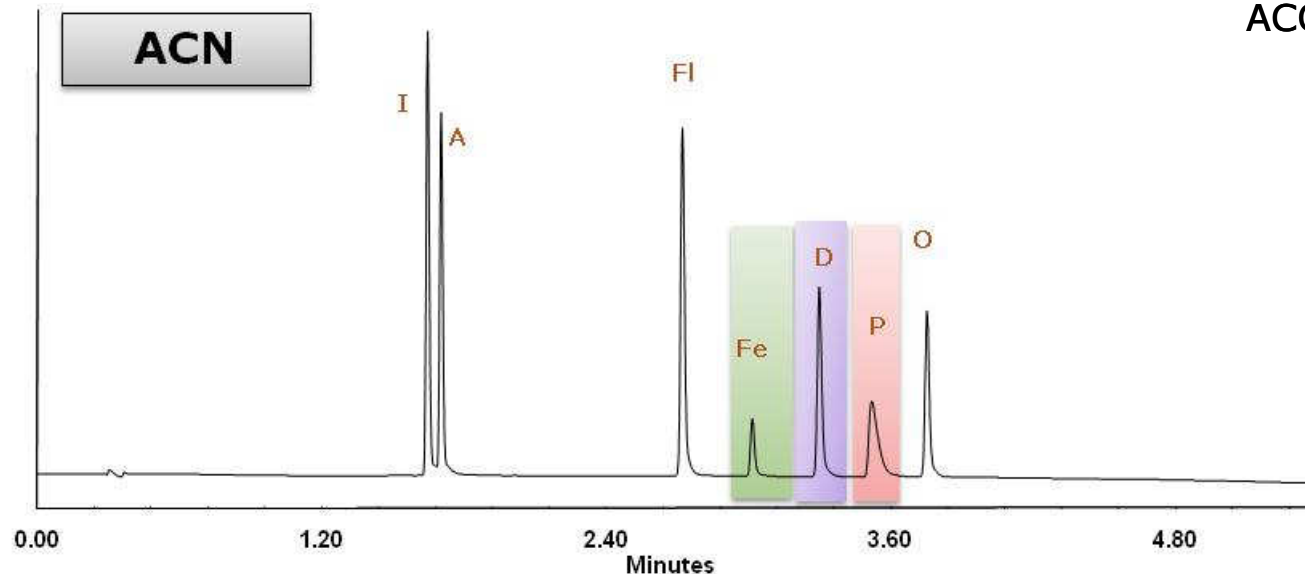
Observations

Methanol is a weaker elution solvent than acetonitrile resulting in greater retention for all analytes

Selectivity difference between bases relative to neutral test probes

Solvent Selectivity: Acidic Compounds

ACQUITY CSH Phenyl-Hexyl, pH 3



Test Probes:

P: 1-pyrenesulfonic acid [A]

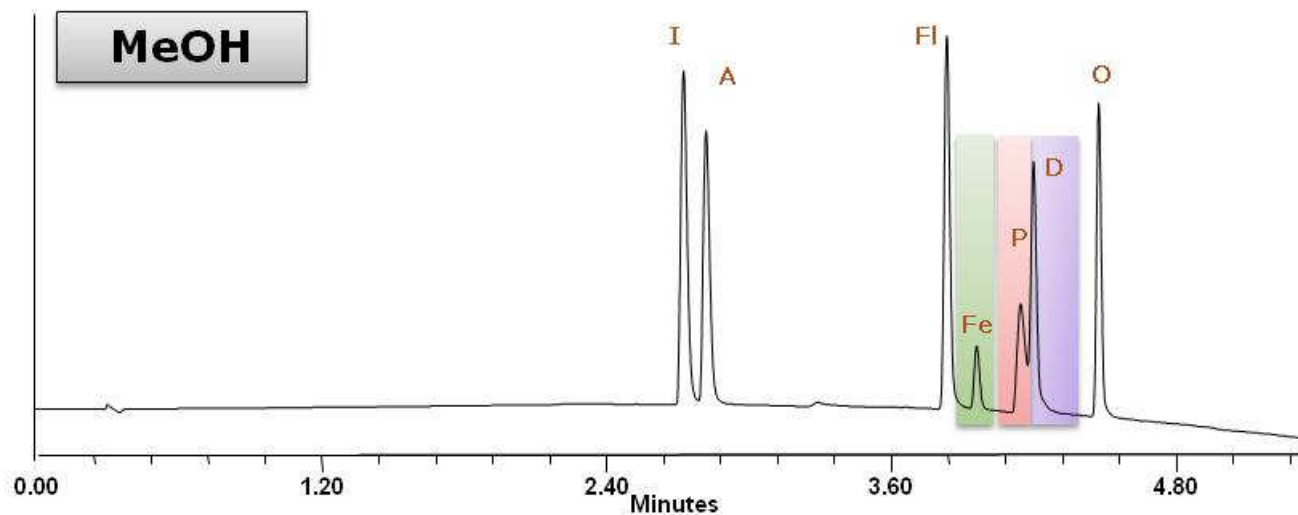
Fe: fenopufen [A]

D: diclofenac [A]

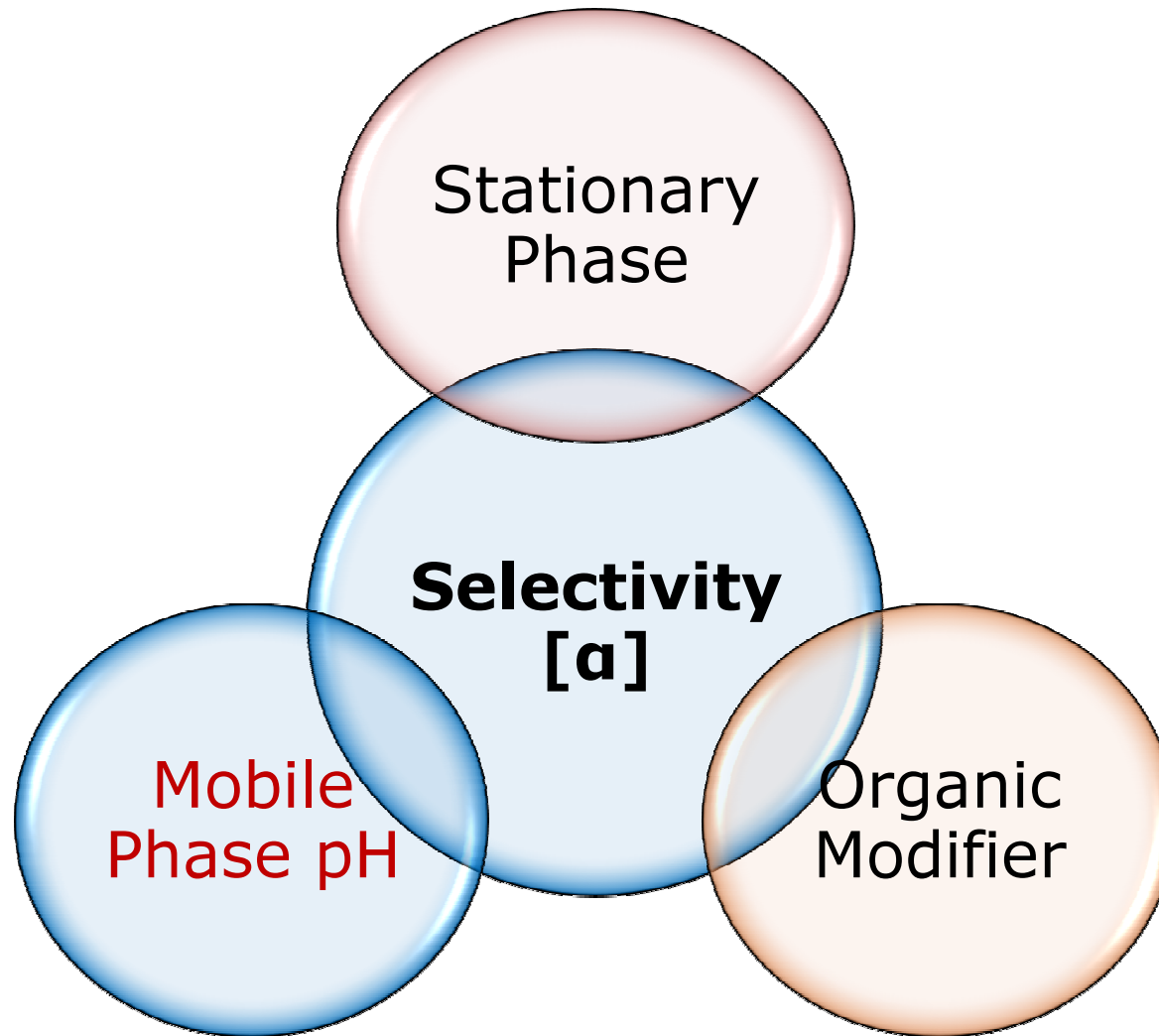
Observations

Methanol is a weaker elution solvent than acetonitrile resulting in greater retention for all analytes

Elution order change for acidic test probes



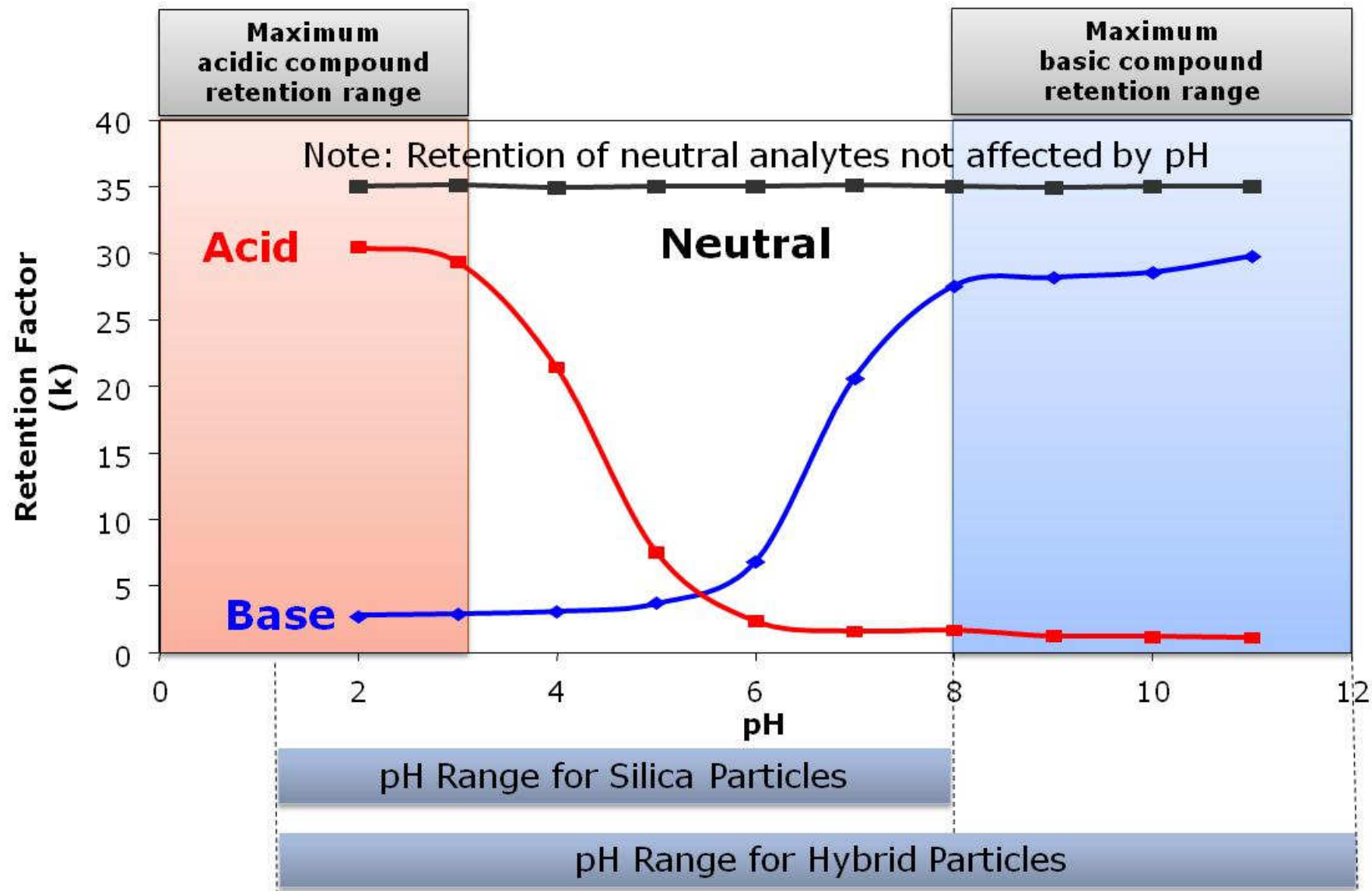
Chemical Factors that Impact Selectivity



Impact of Mobile Phase pH on Retention and Selectivity

- Impacts analytes with ionizable functional groups
 - Amines
 - Carboxylic acids
 - Phenols
- Some compounds contain more than one ionizable group
- Strong selectivity changes can be observed with changes in mobile phase pH

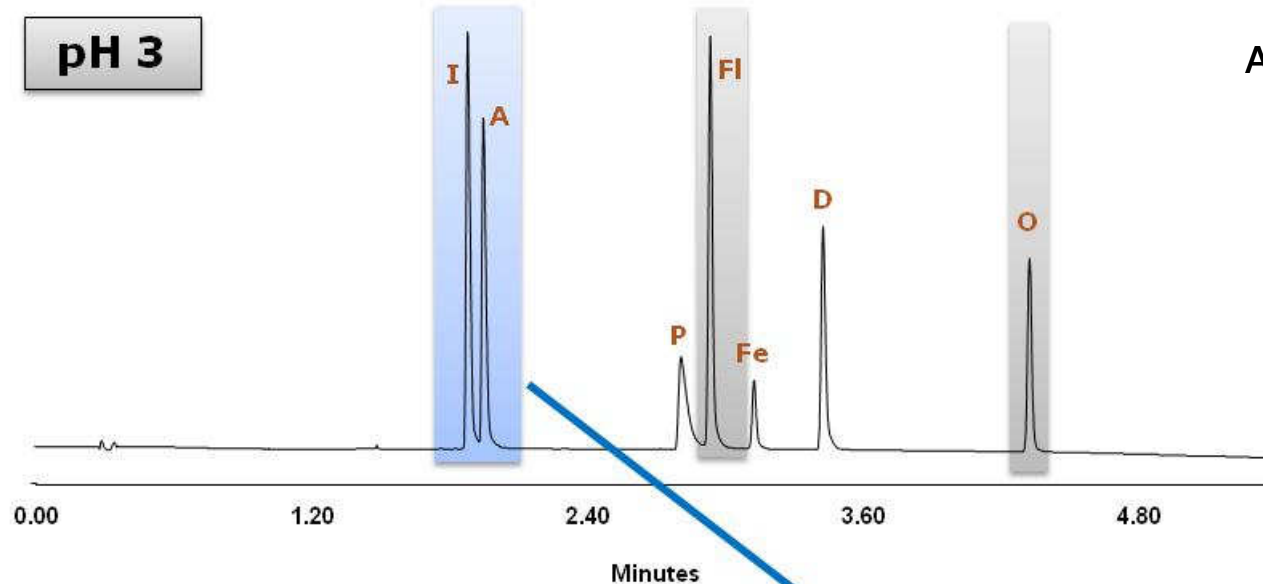
Reversed-Phase Retention Map: The Impact of pH on Ionizable Compounds



Mobile Phase pH Selectivity: Basic and Neutral Compounds

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pH 3

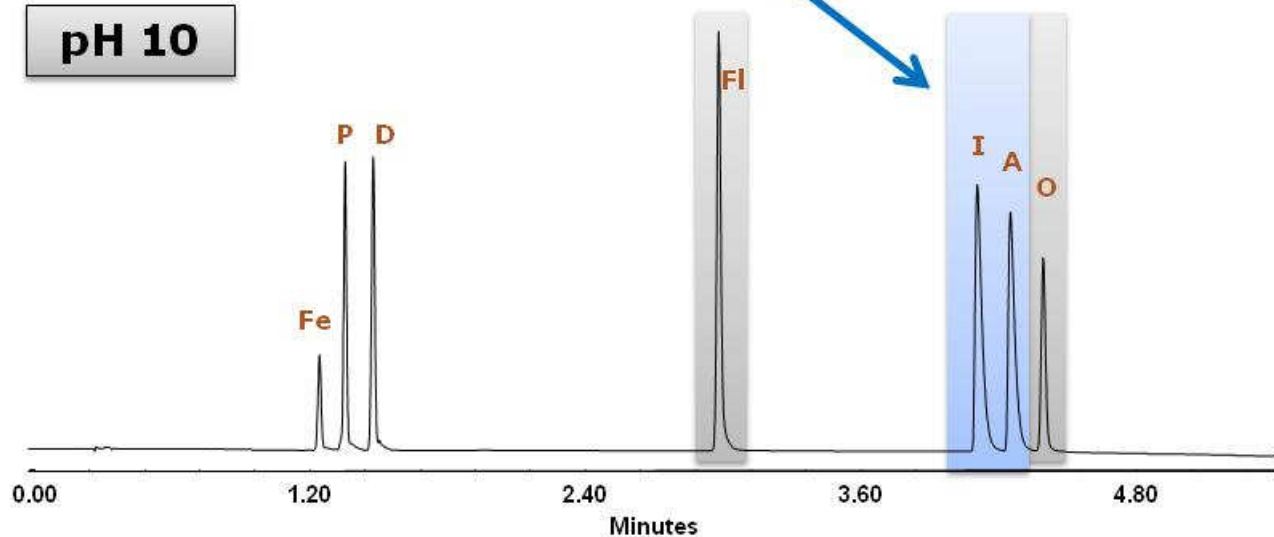


ACQUITY CSH C₁₈, acetonitrile

Test Probes:

- I: Imipramine [B]
- A: Amitriptyline [B]
- FI: Flavone [N]
- O: Octanophenone [N]

pH 10

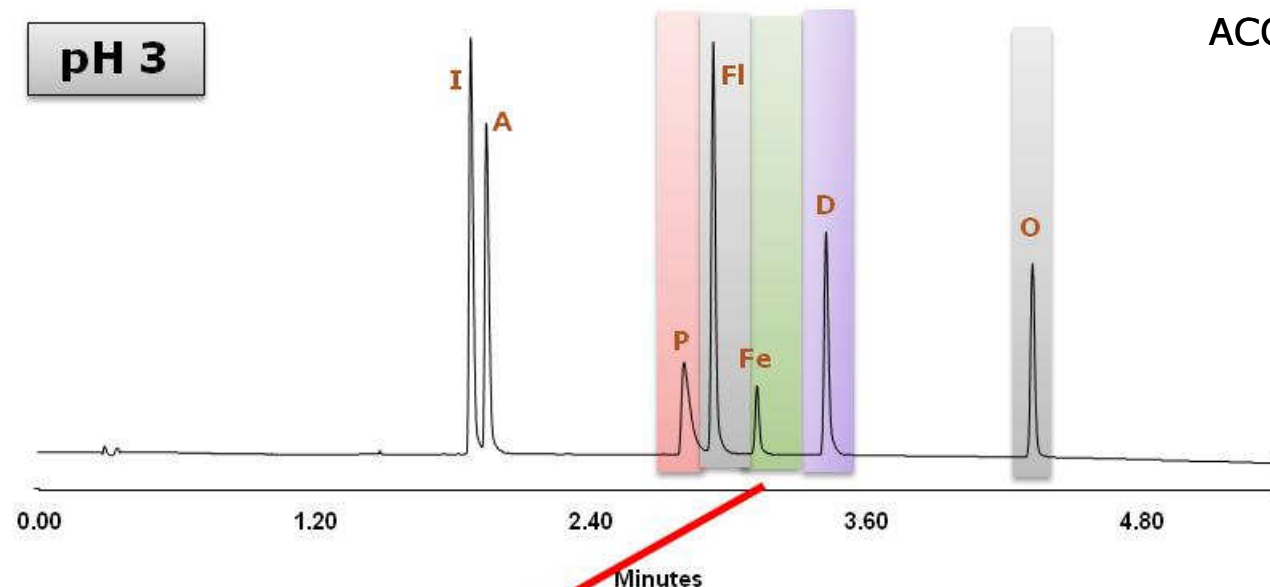


Observations

At high pH, bases are in their neutral [unionized] form, resulting in greater retention

Neutral compounds are unaffected by pH

Mobile Phase pH Selectivity: Acidic and Neutral Compounds



Test Probes:

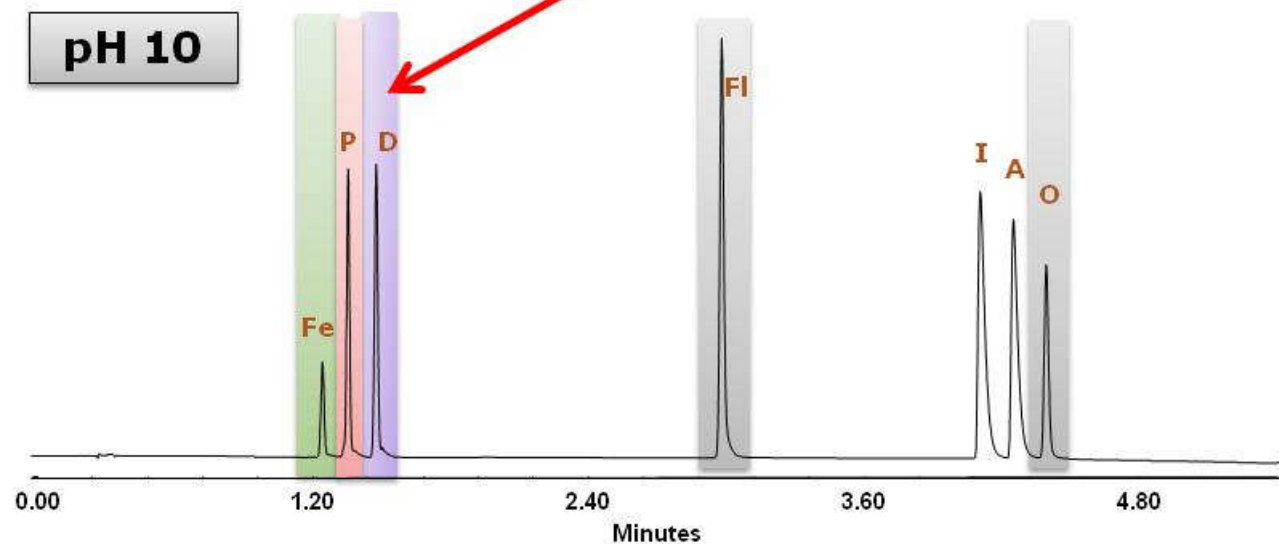
P: 1-pyrenesulfonic acid [A]

Fe: fenoprofen [A]

D: diclofenac [A]

FI: Flavone [N]

O: Octanophenone [N]



Observations

At low pH, acids are in their neutral [unionized] form, resulting in greater retention

Elution order change for acidic compounds

Neutral compounds are unaffected by pH

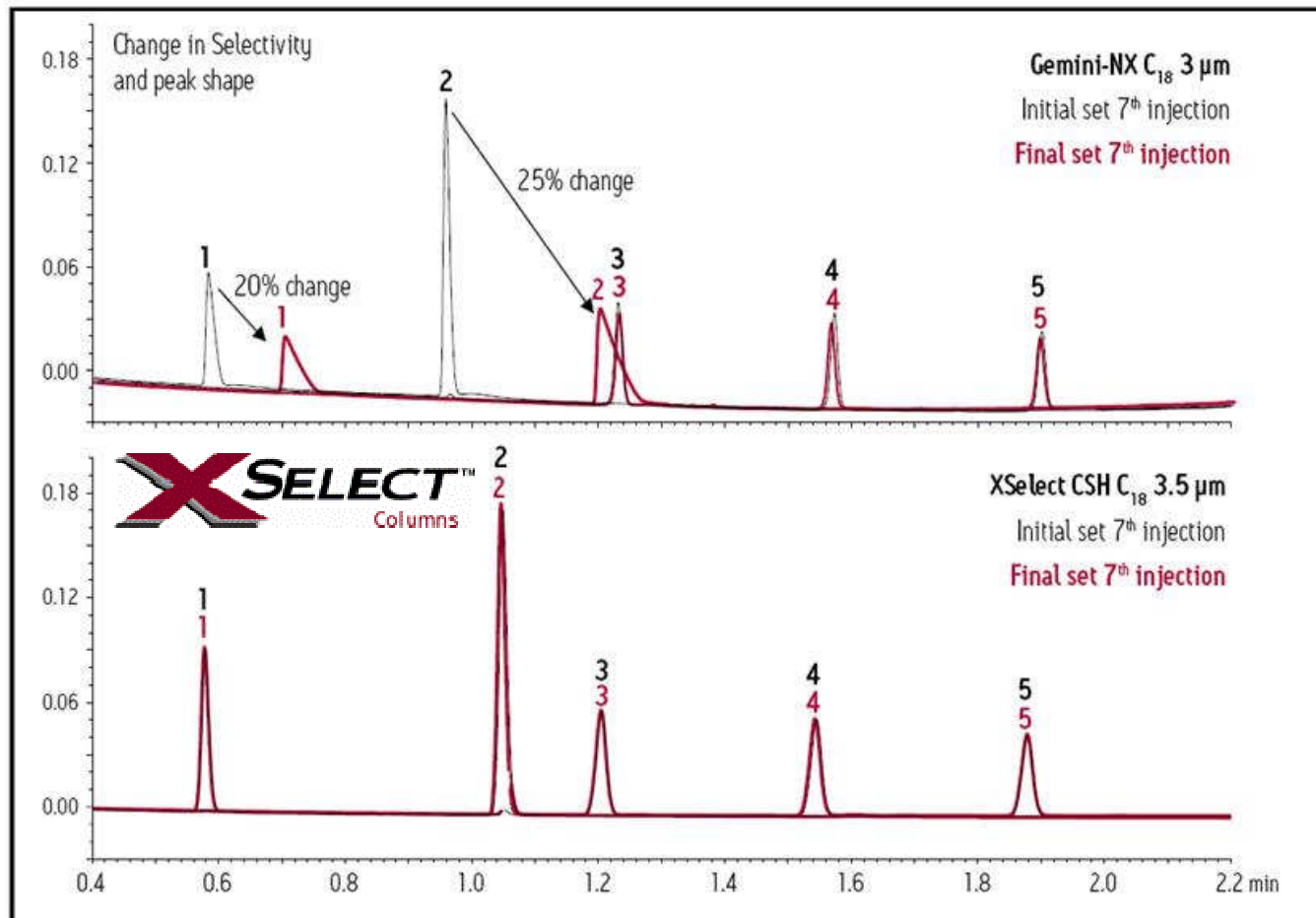
Implementing Mobile Phase pH Switching: Monitoring Column Performance

- Our systematic screening protocol evaluates high and low pH mobile phases.
 - Screen multiple columns and organic modifiers at pH 3 and pH 10
 - Stationary phase must be re-equilibrated when exposed to a new set of conditions
- With low ionic strength mobile phases [i.e., formic acid, ammonium hydroxide], column performance [retention and selectivity] can change *1
 - Slow surface equilibration at low pH
 - Inconsistent selectivity can impact open access systems and method transfer

*1 Marchand, D.H., et al., J. Chromatogr. A **2003**, 1015, 53-64

Implementing Mobile Phase pH Switching: Monitoring Column Performance

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Separations on Gemini-NX C18 (top) and XSelect CSH C18 (bottom) columns (both 2,1 x 50 mm) before and after exposure to a pH 10 mobile phase. Gradient: A: 0.1% formic acid in water; B: acetonitrile; 5 to 95% B linear in 2.5 minutes. Temperature: 30 °C. Injection volume: 2 μL. Detection: 260 nm. Flow rate: 0.8 ml/min. Analytes: (1) metoprolol; (2) amitriptyline; (3) dimethylphthalate; (4) diethylphthalate; (5) dipropylphthalate. System: ACQUITY UPLC.

Observations

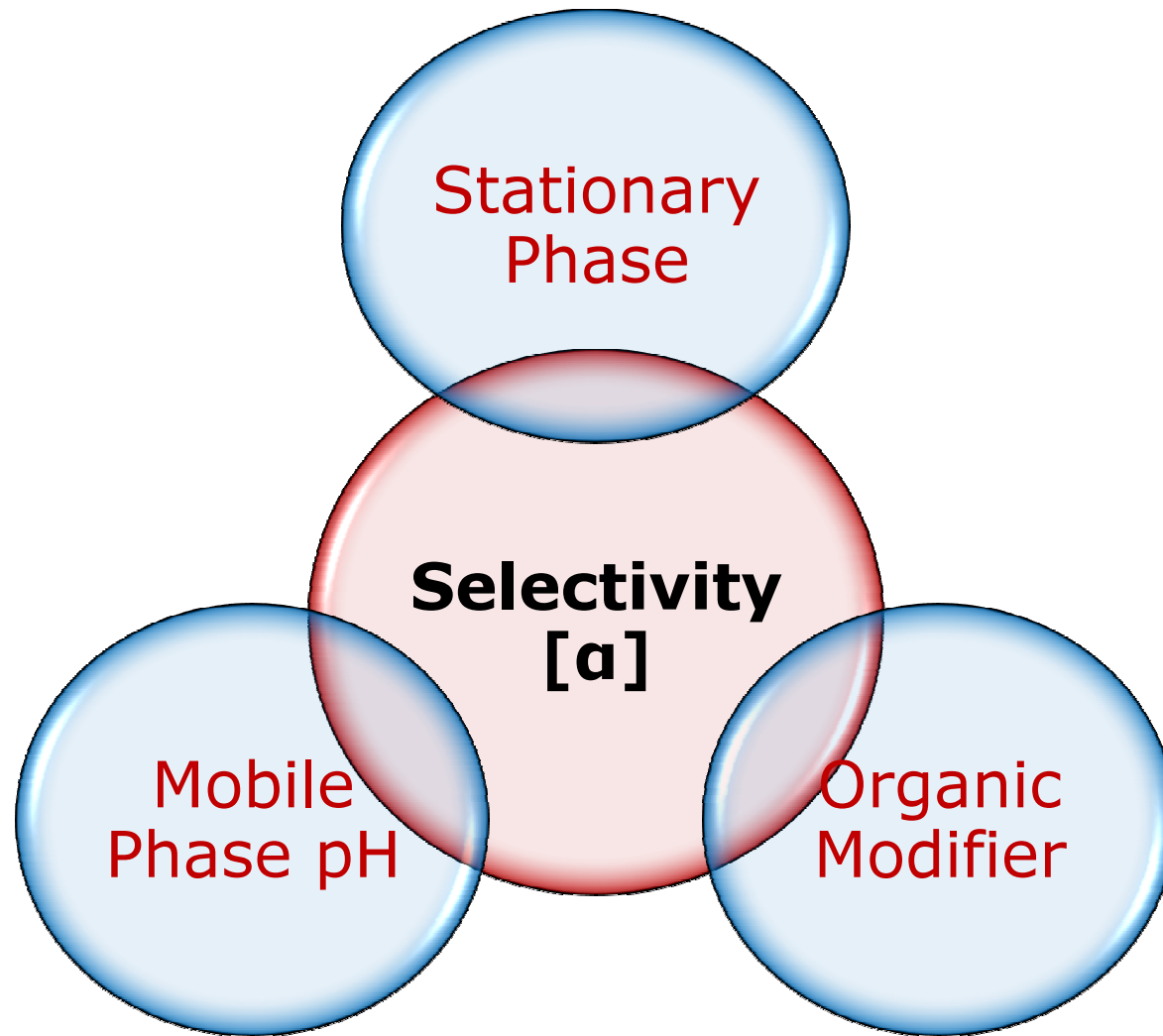
Gemini-NX C₁₈ shows a 20 – 25 % change in retention at low pH, after exposure to high pH mobile phases.

No significant retention or selectivity shift was observed on the XSelect column.

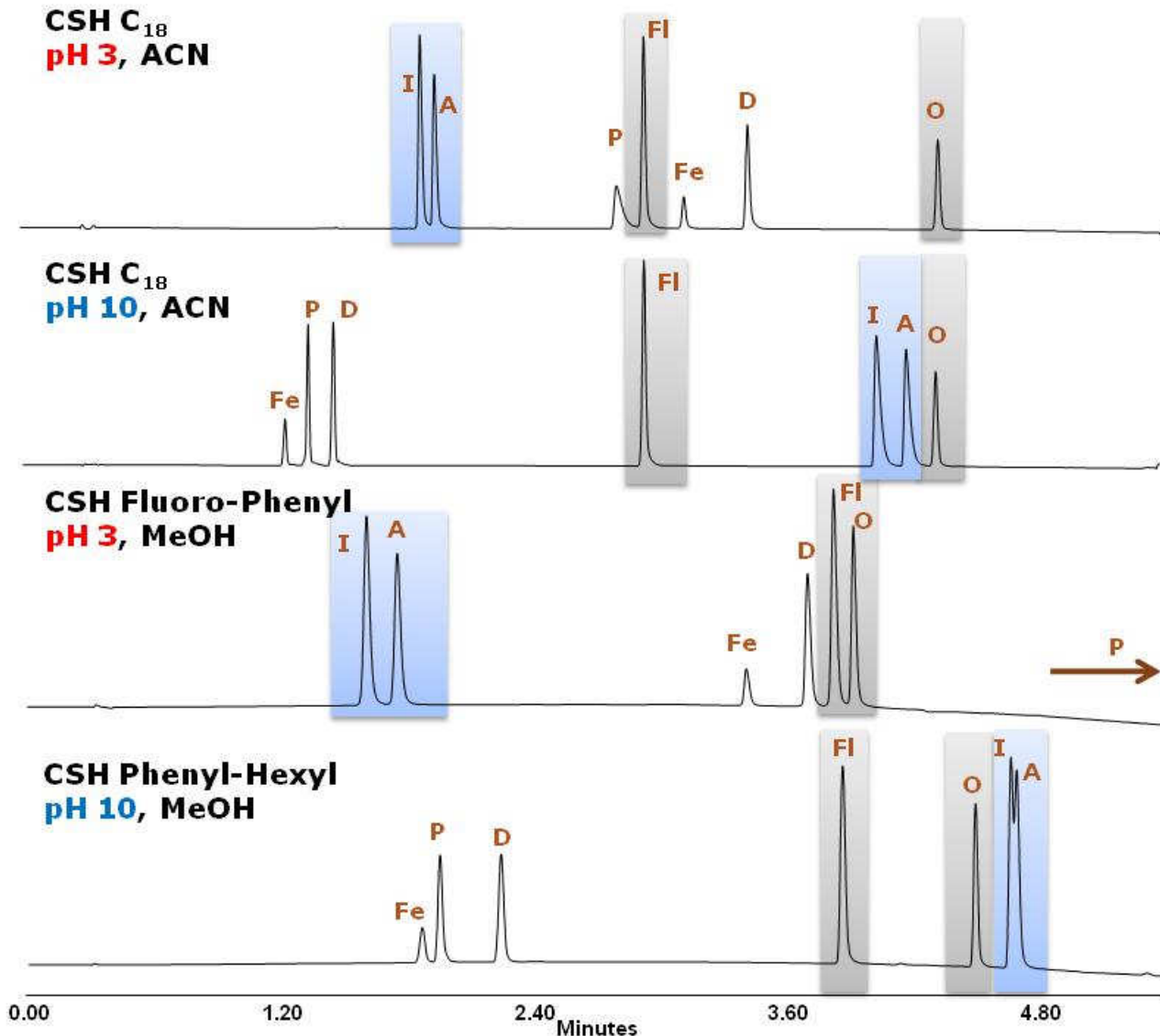


Gemini is a trademark of Phenomenex, Inc.

Chemical Factors that Impact Selectivity



Maximizing Selectivity Differences: Combining Stationary Phase, Organic Modifier and Mobile Phase pH



BASIC AND NEUTRAL TEST PROBES

Test Probes:

I: Imipramine [B]

A: Amitriptyline [B]

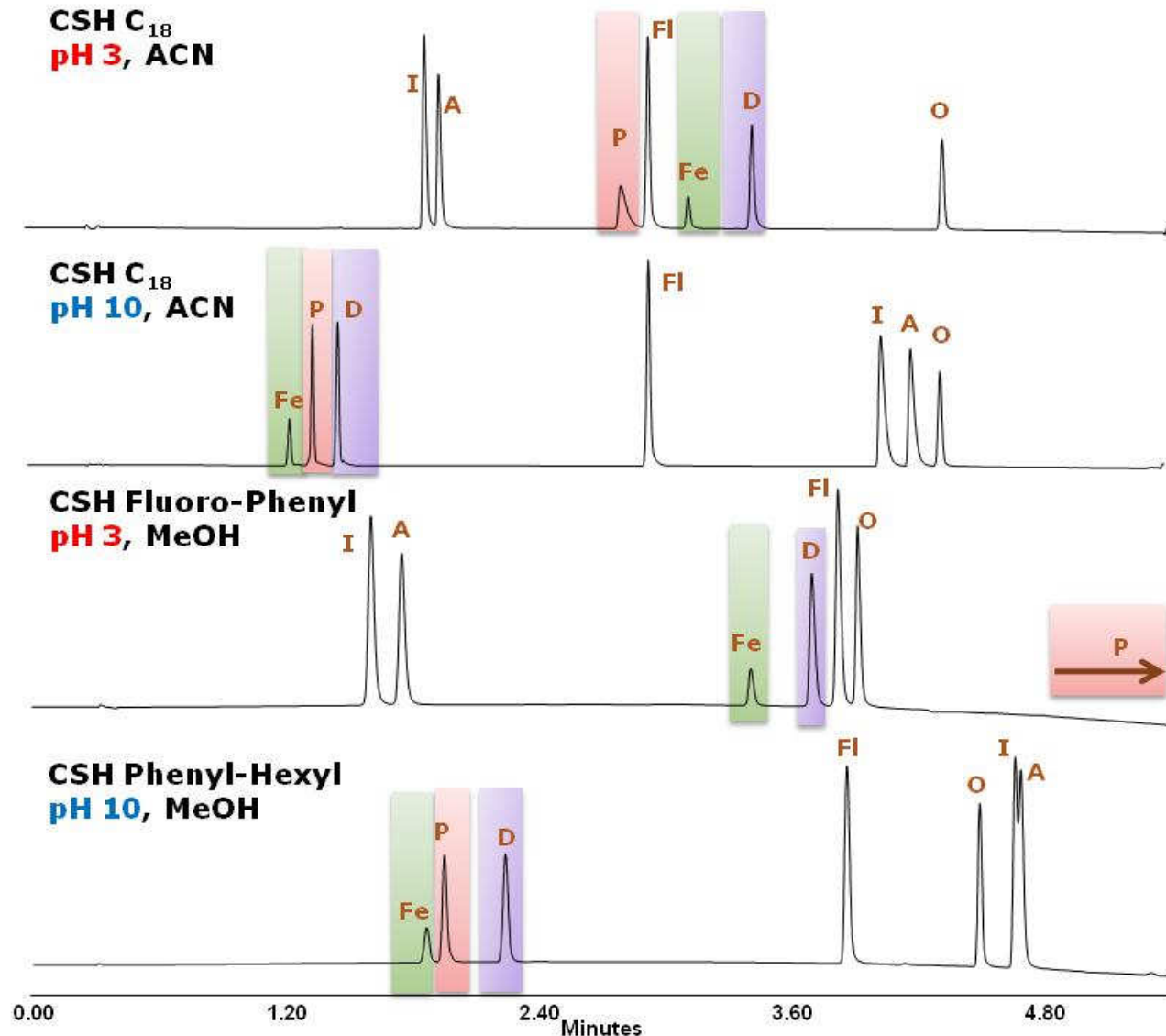
FI: Flavone [N]

O: Octanophenone [N]

Observations

Large differences in selectivity are observed when evaluating combinations of stationary phase, organic modifier and mobile phase pH

Maximizing Selectivity Differences: Combining Stationary Phase, Organic Modifier and Mobile Phase pH



ACIDIC TEST PROBES

Test Probes:

- P: 1-pyrenesulfonic acid [A]
- Fe: fenopufen [A]
- D: diclofenac [A]

Observations

Large differences in selectivity are observed when evaluating combinations of stationary phase, organic modifier and mobile phase pH

- Analytes in their un-ionized [neutral] form yield greater retention
- Methanol is a weaker elution solvent than acetonitrile, and therefore exhibits increased retention of all components, as well as selectivity differences, compared to acetonitrile
- Large differences in selectivity are observed when a change in mobile phase pH alters the charge state of the analyte
- Large selectivity differences are observed between the stationary phases at any given condition
- The most significant selectivity differences occur when comparing combinations of stationary phase, organic modifier and mobile phase pH

- Manipulation of parameters for method development [as described previously] is applicable to both HPLC and UPLC separations
 - Column selectivity [ACQUITY CSH or XSelect CSH HPLC columns]
 - Acetonitrile and methanol
 - pH 3 and pH 10 mobile phases
- Hybrid particle technology enables the exploration of pH extremes in method development
 - Stability from pH 1 – 11
- CSH Technology columns facilitate:
 - Selectivity differences independent of the mobile phase conditions employed
 - The use of low ionic strength mobile phases with high sample capacity
 - Reliable performance when switching between mobile phase pH's
- Evaluation of data from the complete systematic screening protocol is essential to fully understand the analytes chromatographic behavior

Why UPLC Technology for method development?

Develop Methods Faster with UPLC Technology: Maintaining Separation Power

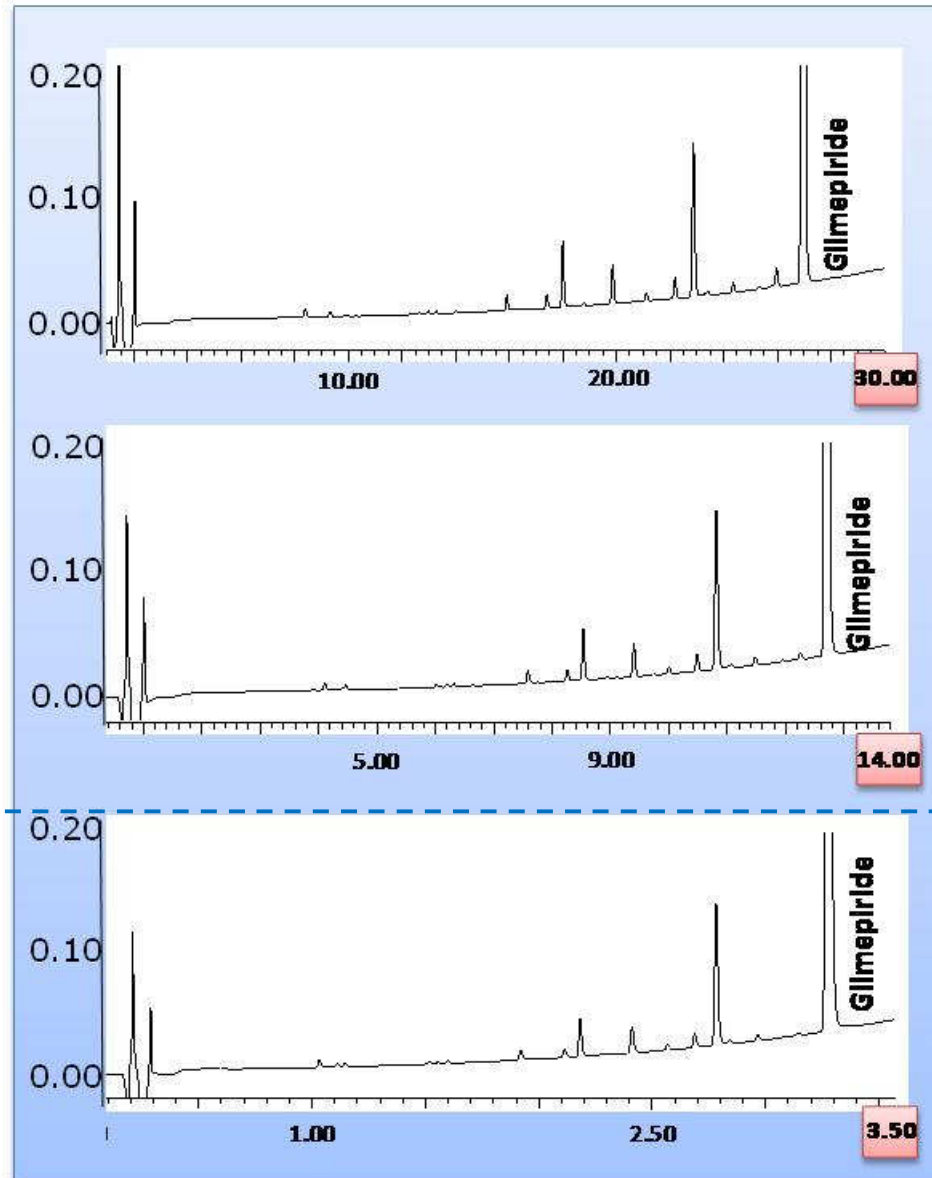
Waters
THE SCIENCE OF WHAT'S POSSIBLE.™

HSS
HIGH STRENGTH SILICA

HPLC
3,5 and
5 μm

1,8 μm
UPLC

Acquity
UPLC®



HPLC

5 μm – 150 mm
 $L/d_p = 30\ 000$

3,5 μm – 100 mm
 $L/d_p = 28\ 571$

UPLC

1,8 μm – 50 mm
 $L/d_p = 27\ 778$

Develop Methods Faster with UPLC Technology: Time Savings

UPLC Gradient Conditions:

Column: 2,1 x 50 mm, 1,7/1,8 μm

Flow Rate: 0,5 ml/min

Gradient: Time [min]	Profile		
	%A	%B	%C
0.0	90	5	5
5.0	5	90	5
5.1	90	5	5
5.5	90	5	5

Peak Capacity [P_c] = 150

HPLC Gradient Conditions:

Column: 4,6 x 100 mm, 3,5 μm

Flow Rate: 1,17 ml/min

Gradient: Time [min]	Profile		
	%A	%B	%C
0.0	90	5	5
20.6	5	90	5
21.0	90	5	5
22.6	90	5	5

Peak Capacity [P_c] = 150

$$P_c = 1 + \frac{t_g}{w}$$

**UPLC can achieve the same separation
4X faster than 3.5 μm HPLC**

Develop Methods Faster with UPLC Technology: Time Savings

UPLC Method Development Protocol:

2.1 x 50 mm, 1.7/1.8 µm, 0.5 mL/min

<u>pH 3/ acetonitrile</u>	<u>Time</u>
Flow ramp	3 min
Column conditioning [2 blank gradients]	11 min
Sample injection [2 replicates]	11 min
<u>pH 3/ methanol</u>	<u>Time</u>
Flow ramp	3 min
Column conditioning [2 blank gradients]	11 min
Sample injection [2 replicates]	11 min
Column purge	10 min
<u>pH 10/ acetonitrile</u>	<u>Time</u>
Flow ramp	3 min
Column conditioning [2 blank gradients]	11 min
Sample injection [2 replicates]	11 min
<u>pH 10/ methanol</u>	<u>Time</u>
Flow ramp	3 min
Column conditioning [2 blank gradients]	11 min
Sample injection [2 replicates]	11 min
Column purge	10 min
SCREENING TIME	120 min
2 hours/ column [low/high pH switching]	
x 2 column	
1 hour/ column [low pH only]	
x 2 column	

TOTAL SCREENING TIME: 6 HOURS

HPLC Method Development Protocol:

4.6 x 100 mm, 3.5 µm, 1.17 mL/min

<u>pH 3/ acetonitrile</u>	<u>Time</u>
Flow ramp	3 min
Column conditioning [2 blank gradients]	45.6 min
Sample injection [2 replicates]	45.6 min
<u>pH 3/ methanol</u>	<u>Time</u>
Flow ramp	3 min
Column conditioning [2 blank gradients]	45.6 min
Sample injection [2 replicates]	45.6 min
Column purge	41 min
<u>pH 10/ acetonitrile</u>	<u>Time</u>
Flow ramp	3 min
Column conditioning [2 blank gradients]	45.6 min
Sample injection [2 replicates]	45.6 min
<u>pH 10/ methanol</u>	<u>Time</u>
Flow ramp	3 min
Column conditioning [2 blank gradients]	45.6 min
Sample injection [2 replicates]	45.6 min
Column purge	41 min
SCREENING TIME	459 min
7.65 hours/ column [low/high pH switching]	
x 2 column	
3.82 hour/ column [low pH only]	
x 2 column	

TOTAL SCREENING TIME: 23 HOURS

**Develop methods 4X faster
with UPLC**

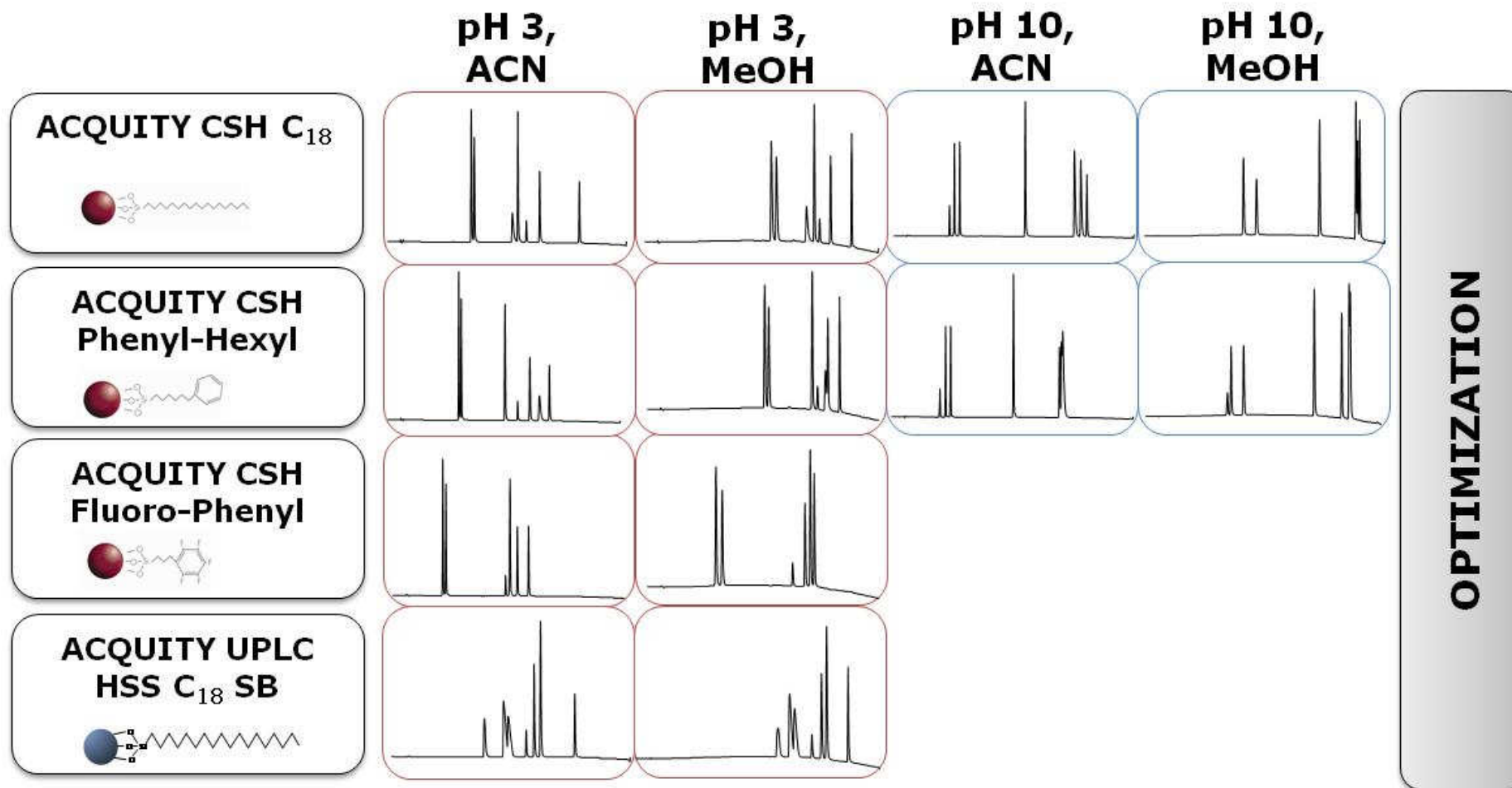
- **Introduction**
 - Approaches Toward Method Development
 - UPLC Technology
 - Success Criteria
- **Controlling Selectivity and Retention**
 - Stationary Phase and Particle Substrate Design
 - Organic Modifier
 - Mobile Phase pH
- **Method Development Strategy**
 - Systematic Screening Protocol
 - Quality by Design [Q_bD] Approach
- **Implementing the Approach**
 - Case Study
- **Conclusion**



Systematic Screening Protocol

- ACQUITY UPLC H-Class Quaternary Solvent manager [QSM] with solvent select valve
 - Mix up to 4 solvents
 - Optional solvent select valve enables an additional 5 solvent lines
- ACQUITY UPLC H-Class Column Manager
 - Flexible modules to select between 2 and 6 columns
 - Utilize 2,1 x 50 mm, 1,7/1,8 μm columns
- Fast, 5 minute gradient from 5 – 90 % organic at 0,5 ml/min

Systematic Screening Protocol



Method Development: Quality by Design [Q_bD] Approach

- Systematic Screening Protocol
 - Good first pass, rapid method development
 - Choice of best combination of parameters [i.e., stationary phase, organic modifier, mobile phase pH] is subjective
 - Optimum separation conditions may be outside of screening approach [i.e., pH 5 with a mixture of acetonitrile and methanol]

- Quality by Design with Design of Experiments [DOE] Approach
 - Start with systematic screening protocol
 - Define separation objectives
 - Gain knowledge about the product or process
 - Create sufficient scientific understanding to establish a design space, specifications and controls
 - Defines robust operating space

Method Development: Quality by Design [Q_bD] Approach

Design of Experiments [DOE] Approach

- Fusion AE Method Development Software
 - Aligned with FDA and EMA Q_bD initiatives
 - Applies DOE approach to method development using simple templates
 - Facilitates data interpretation
 - Incorporates robustness modeling into the chromatographic development process
 - Automates sample and method set creation in Empower



[1] Define Experimental Region

Select study variables

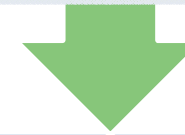
Define study ranges



[2] Develop Knowledge Space

Conduct experimental design

Analyze results and study variable effects



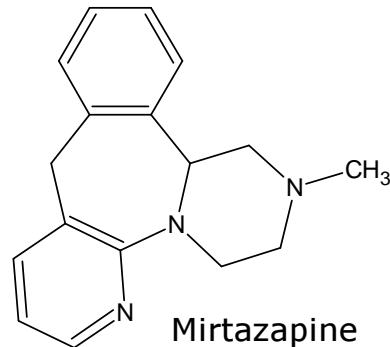
[3] Establish Design Space

Define optimum conditions

Define robust operating space

- **Introduction**
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 - UPLC Technology
 - Success Criteria
- **Controlling Selectivity and Retention**
 - Stationary Phase and Particle Substrate Design
 - Organic Modifier
 - Mobile Phase pH
- **Method Development Strategy**
 - Systematic Screening Protocol
 - Quality by Design [Q_bD] Approach
- **Implementing the Approach**
 - Case Study
- **Conclusion**

Implementing the Approach: Mirtazapine and Impurities



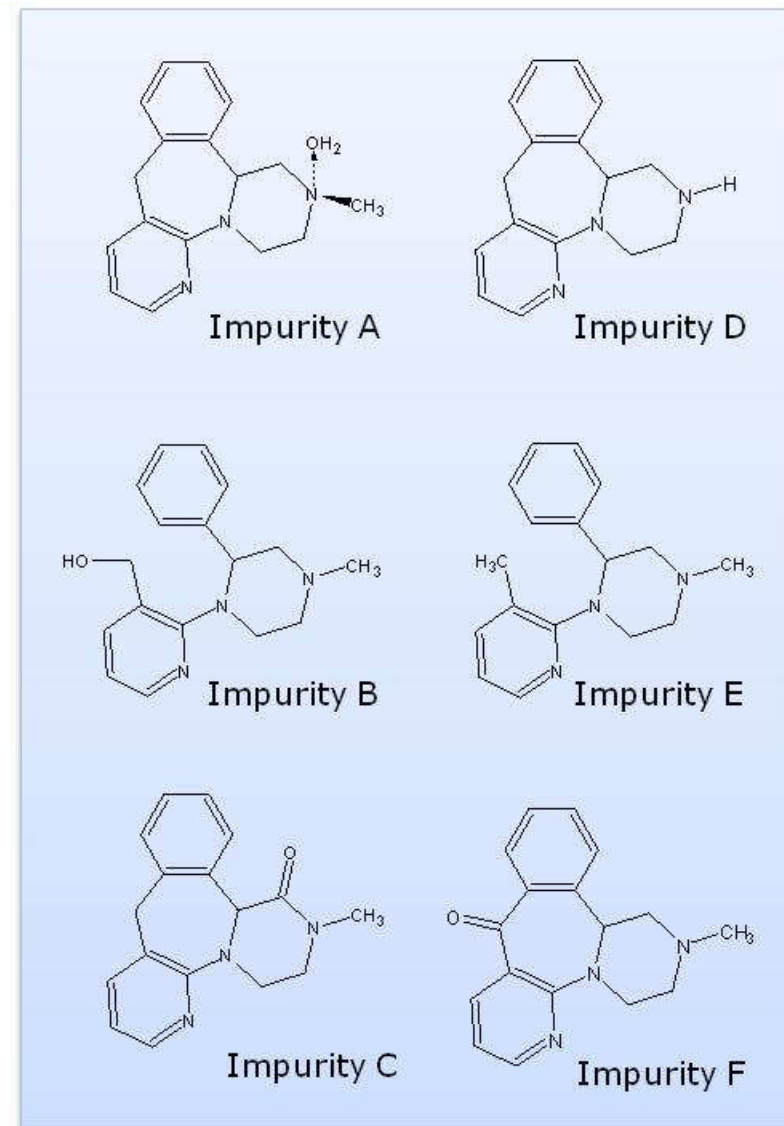
Mirtazapine [m.w. 265.35]

Used primarily for the treatment of clinical depression

USP mirtazapine resolution mix
RS 1.0 mg/ml in 50:50
ACN:H₂O

Method development and optimization

- Empower 2 CDS with Fusion AE method development software
 - Uses statistically significant combination of different parameters [*software will not run every combination of every parameter*]
 - 4 column chemistries
 - 2 organic modifiers
 - 2 mobile phase pH's
 - Gradient times: 2, 3.1, 4.3, 5.4, 6.5 min



Fusion AE Experimental Design

Replicates built into design:
 1.a.1.a, 12.a.1.a; 2.a.1.a, 13.a.1.a
 3.a.1.a, 14.a.1.a; 4.a.1.a, 15.a.1.a
 18.a.1.a, 19.a.1.a; 22.a.1.a, 30.a.1.a
 23.a.1.a, 28.a.1.a; 24.a.1.a, 32.a.1.a
 26.a.1.a, 33.a.1.a; 34.a.1.a, 37.a.1.a

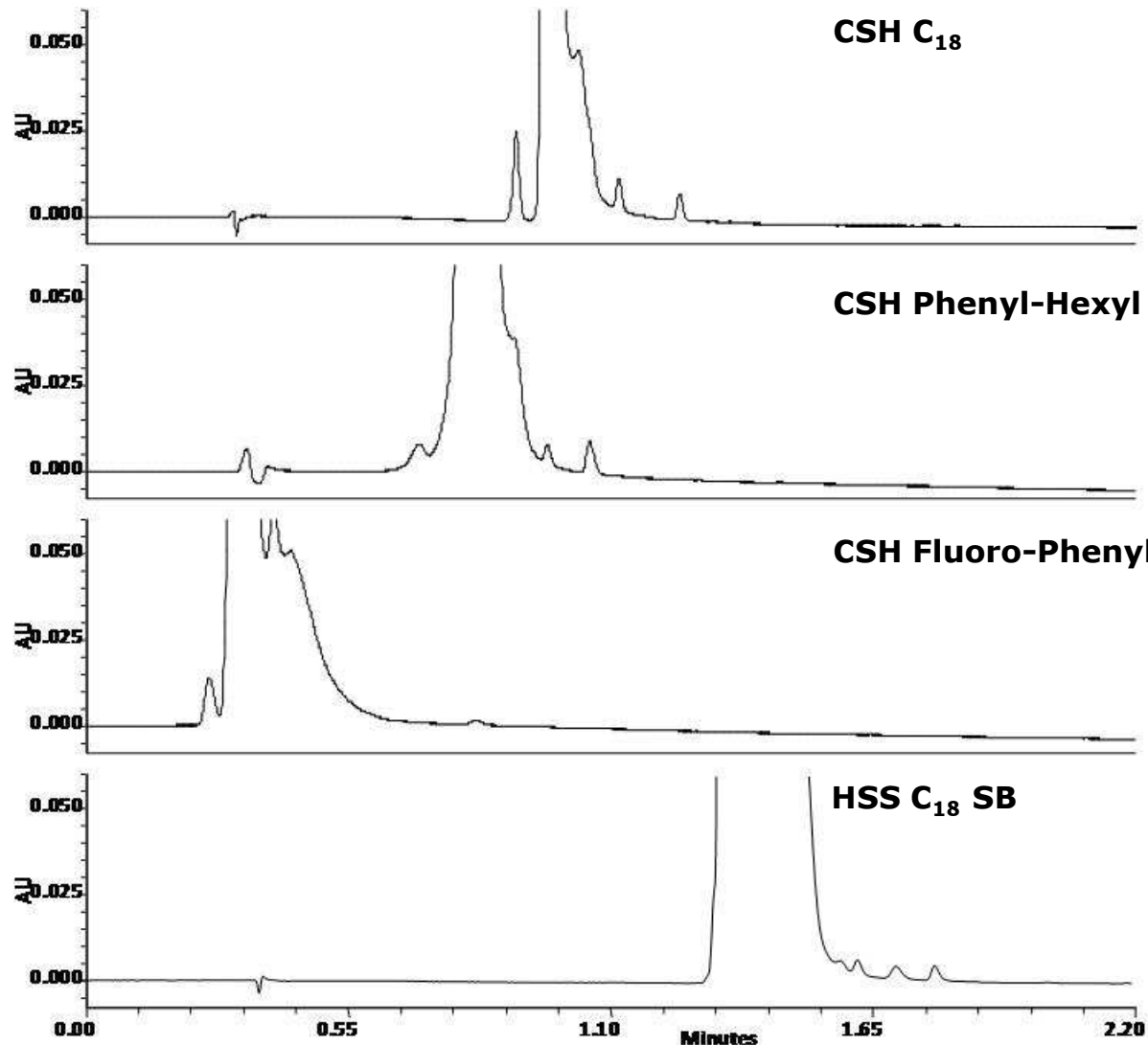
Run No.	Gradient Time (min)	Organic Solvent Type (*)	pH (*)	Column Type (*)
Wash - 1	0.1	Acetonitrile	2.6	CSH HL C18
Wash - 2	0.1	Acetonitrile	2.6	CSH Phenyl-Hexyl
Wash - 3	0.1	Acetonitrile	2.6	CSH Fluoro-Phenyl
Wash - 4	0.1	Acetonitrile	2.6	HSS C18 SB
1.a.1.a	4.3	Acetonitrile	2.6	CSH HL C18
2.a.1.a	4.3	Acetonitrile	2.6	HSS C18 SB
3.a.1.a	4.3	Acetonitrile	2.6	CSH Fluoro-Phenyl
4.a.1.a	4.3	Acetonitrile	2.6	CSH Phenyl-Hexyl
5.a.1.a	5.4	Acetonitrile	2.6	HSS C18 SB
6.a.1.a	6.5	Acetonitrile	2.6	CSH Phenyl-Hexyl
7.a.1.a	2	Acetonitrile	2.6	CSH HL C18
8.a.1.a	2	Acetonitrile	2.6	CSH HL C18
9.a.1.a	6.5	Acetonitrile	2.6	CSH Phenyl-Hexyl
10.a.1.a	6.5	Acetonitrile	2.6	CSH Fluoro-Phenyl
11.a.1.a	2	Acetonitrile	2.6	CSH Fluoro-Phenyl
12.a.1.a	4.3	Acetonitrile	2.6	CSH Fluoro-Phenyl
13.a.1.a	4.3	Acetonitrile	2.6	HSS C18 SB
14.a.1.a	4.3	Acetonitrile	2.6	CSH HL C18
15.a.1.a	4.3	Acetonitrile	2.6	CSH HL C18
Wash - 5	0.1	Acetonitrile	2.6	CSH Phenyl-Hexyl
Wash - 6	0.1	Acetonitrile	2.6	CSH HL C18
16.a.1.a	2	Acetonitrile	10.6	CSH Phenyl-Hexyl
17.a.1.a	4.3	Acetonitrile	10.6	CSH HL C18
18.a.1.a	6.5	Acetonitrile	10.6	CSH Phenyl-Hexyl
19.a.1.a	6.5	Acetonitrile	10.6	CSH Phenyl-Hexyl
Wash - 7	0.1	Methanol	2.6	CSH HL C18
Wash - 8	0.1	Methanol	2.6	CSH Phenyl-Hexyl
Wash - 9	0.1	Methanol	2.6	CSH Fluoro-Phenyl
Wash - 10	0.1	Methanol	2.6	HSS C18 SB

Fusion AE software automatically constructs a set of experiments by selecting the most efficient statistical experimental design

Instrument methods, method sets and sample sets are automatically created in Empower 2 to carry out the experiment

20.a.1.a	3.1	Methanol	2.6	CSH Phenyl-Hexyl
				HSS C18 SB
				CSH Fluoro-Phenyl
				CSH Phenyl-Hexyl
				HSS C18 SB
				CSH Phenyl-Hexyl
				CSH HL C18
				CSH HL C18
				CSH Phenyl-Hexyl
				CSH Fluoro-Phenyl
				CSH Fluoro-Phenyl
				HSS C18 SB
				CSH HL C18
				CSH HL C18
				CSH Phenyl-Hexyl
				CSH HL C18
35.a.1.a	2	Methanol	10.6	CSH Phenyl-Hexyl
36.a.1.a	2	Methanol	10.6	CSH HL C18
37.a.1.a	6.5	Methanol	10.6	CSH HL C18
38.a.1.a	5.4	Methanol	10.6	CSH Phenyl-Hexyl
Wash - 13	0.1	Methanol	10.6	CSH HL C18
Wash - 14	0.1	Methanol	10.6	CSH Phenyl-Hexyl
Wash - 15	0.1	Methanol	2.6	CSH Fluoro-Phenyl
Wash - 16	0.1	Methanol	2.6	HSS C18 SB

Stationary Phase Selectivity: Mirtazapine and Impurities



Acetonitrile, Low pH

T_g = 4.3 min 5-95% B

Observations:

Selectivity differences observed between the different columns

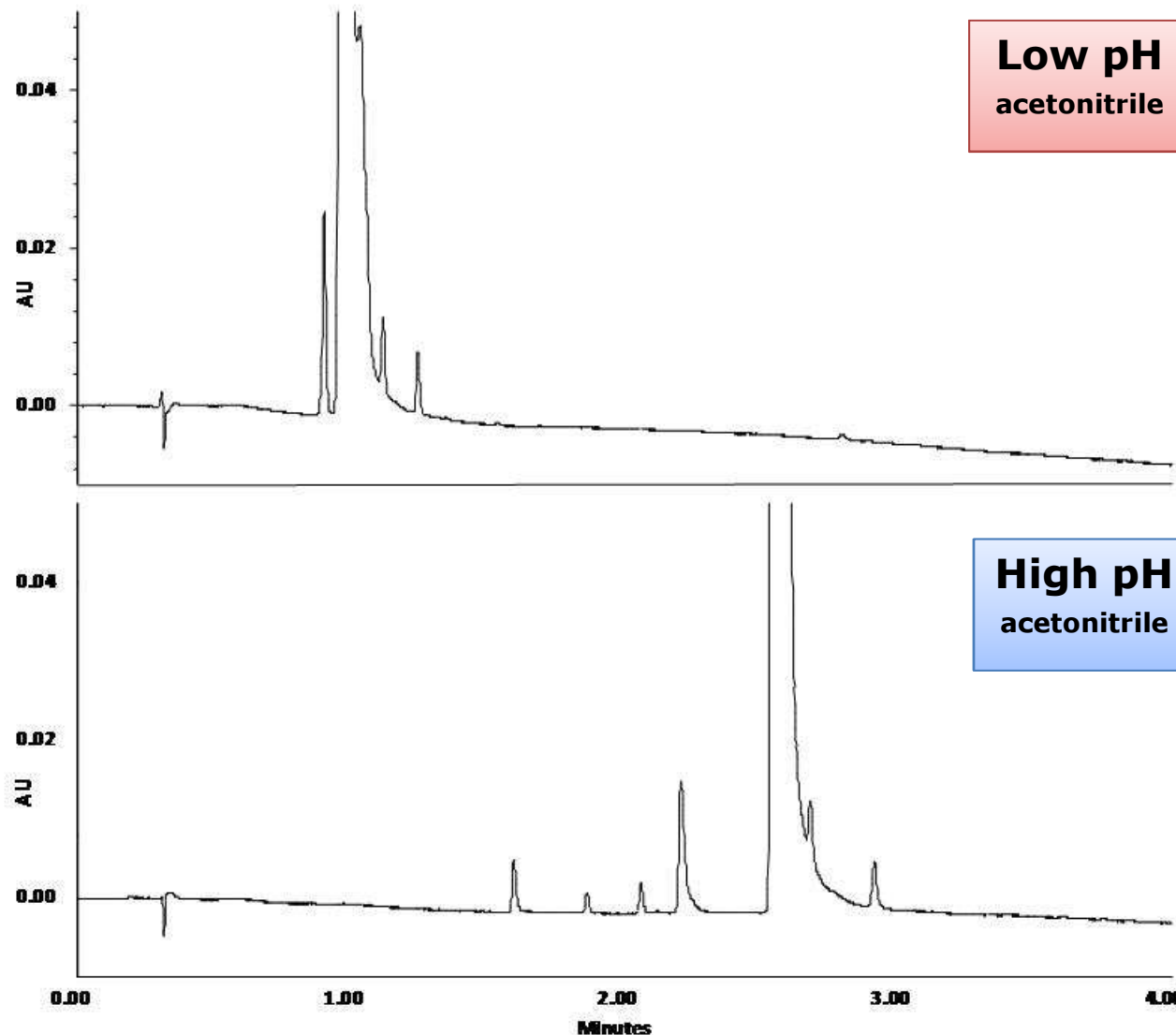
Inadequate retention on Fluoro-Phenyl

Poor resolution of mirtazapine and 6 impurities on all columns

Action:

Investigate mobile phase pH

Mobile Phase pH Selectivity: Mirtazapine and Impurities



Low pH
acetonitrile

ACQUITY CSH C₁₈
T_g = 4.3 min 5-95% B

Observations:

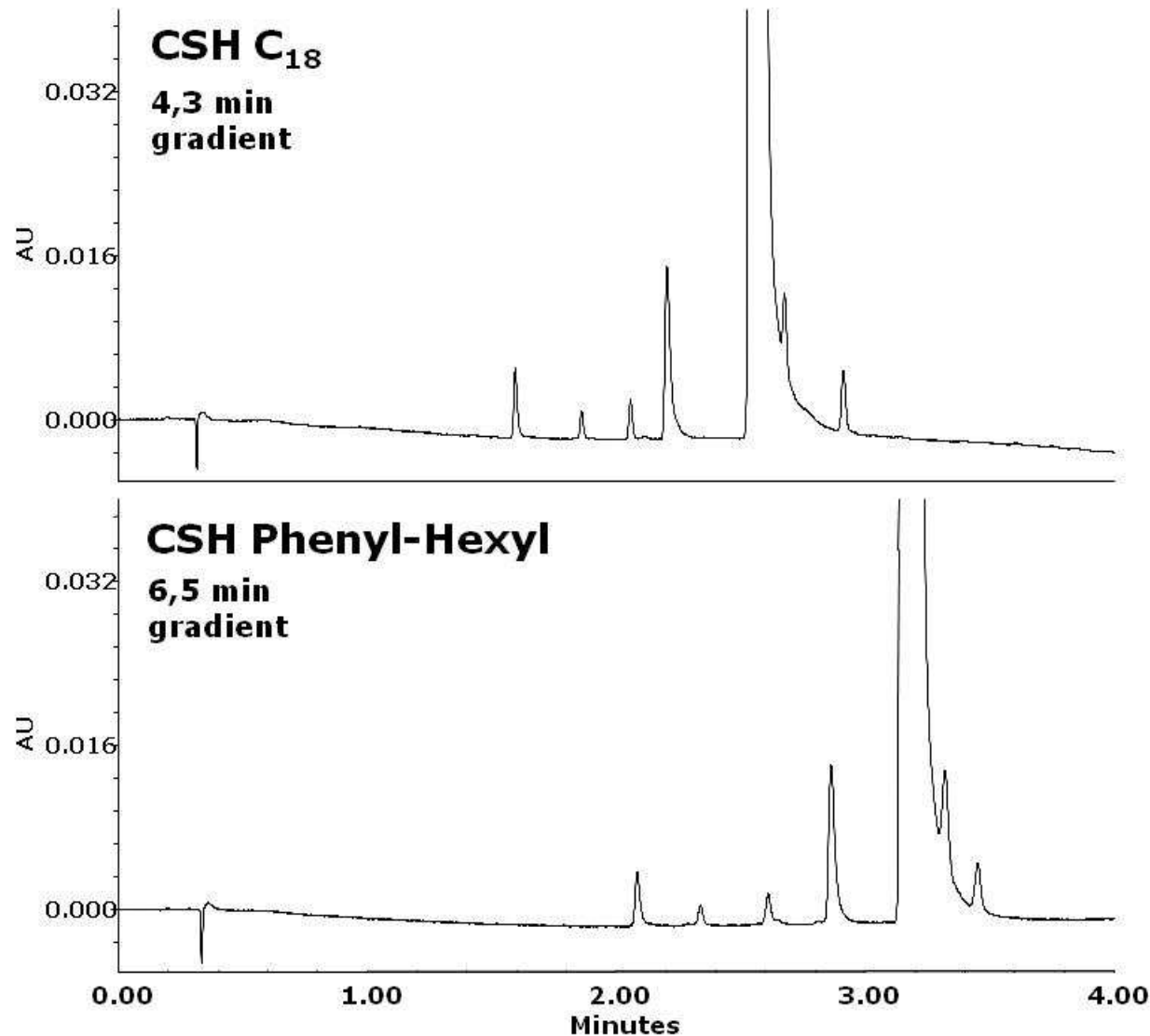
Better retention at high pH due to the neutral charge state of the analytes

Improved resolution of impurities from mirtazapine at high pH

Action:

Compare stationary phase selectivity at high pH

Stationary Phase Selectivity, High pH: Mirtazapine and Impurities



Acetonitrile, High pH
5-95% B

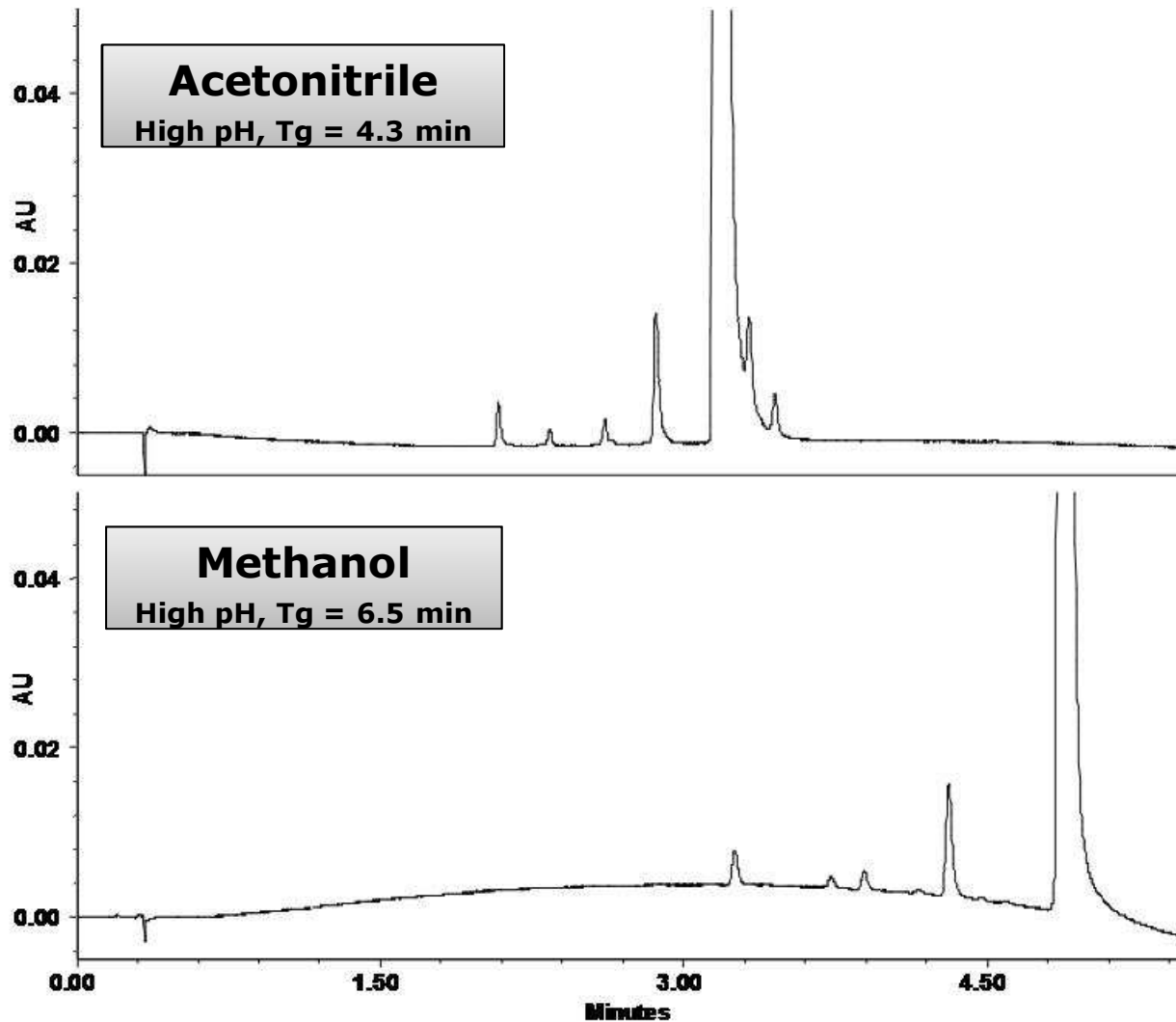
Observations:

Similar selectivity and resolution on both columns at high pH

Action:

Investigate organic modifier

Solvent Selectivity, High pH: Mirtazapine and Impurities



ACQUITY CSH C18

5-95% B

Observations:

Methanol is a weaker elution solvent resulting in greater retention

Better resolution of impurities from mirtazapine with acetonitrile

Action:

Fusion AE software
Automated Method
Optimizer

Data Analysis from Screening Protocol: Automated Method Optimizer

Optimize Responses - Study Variable Settings

Variable Name	Units	Type	Lower Bound	Upper Bound
Pump Flow Rate	mL/min	Continuous	0.200	
Gradient Time	min	Continuous	2.00	
Final % Organic	%	Continuous	60.00	
Oven Temperature	°C	Continuous	30.0	

Study Variable Settings Validation Results
Your settings are valid.

Restore Defaults << Back

Optimize Responses - Response Variable Goals

Response Name	Goal	Lower Bound	Upper Bound	Relative Rank
<input checked="" type="checkbox"/> No. of Peaks	Maximize	5	11	1
<input checked="" type="checkbox"/> No. of Peaks >= 1.00 - USPResolution	Maximize	2	10	1
<input checked="" type="checkbox"/> No. of Peaks >= 1.50 - USPResolution	Maximize	2	9	1
<input checked="" type="checkbox"/> No. of Peaks >= 2.00 - USPResolution	Maximize	2	8	0.9
<input type="checkbox"/> Max Peak #1 - USPTailing				
<input checked="" type="checkbox"/> Max Peak #1 - Area	Minimize	836,952.9395074	10,783,722.6259	0.6
<input checked="" type="checkbox"/> No. of Peaks <= 1.50 - USPTailing	Maximize	1	8	1
<input checked="" type="checkbox"/> Last Peak - RetentionTime	Minimize	1.569234511	7.081078898	0.9
<input checked="" type="checkbox"/> No. of Peaks <= 0.10 - WidthAt4_4Pct	Maximize	3	10	1
<input checked="" type="checkbox"/> No. of Peaks <= 0.05 - WidthAt4_4Pct	Maximize	0	9	1

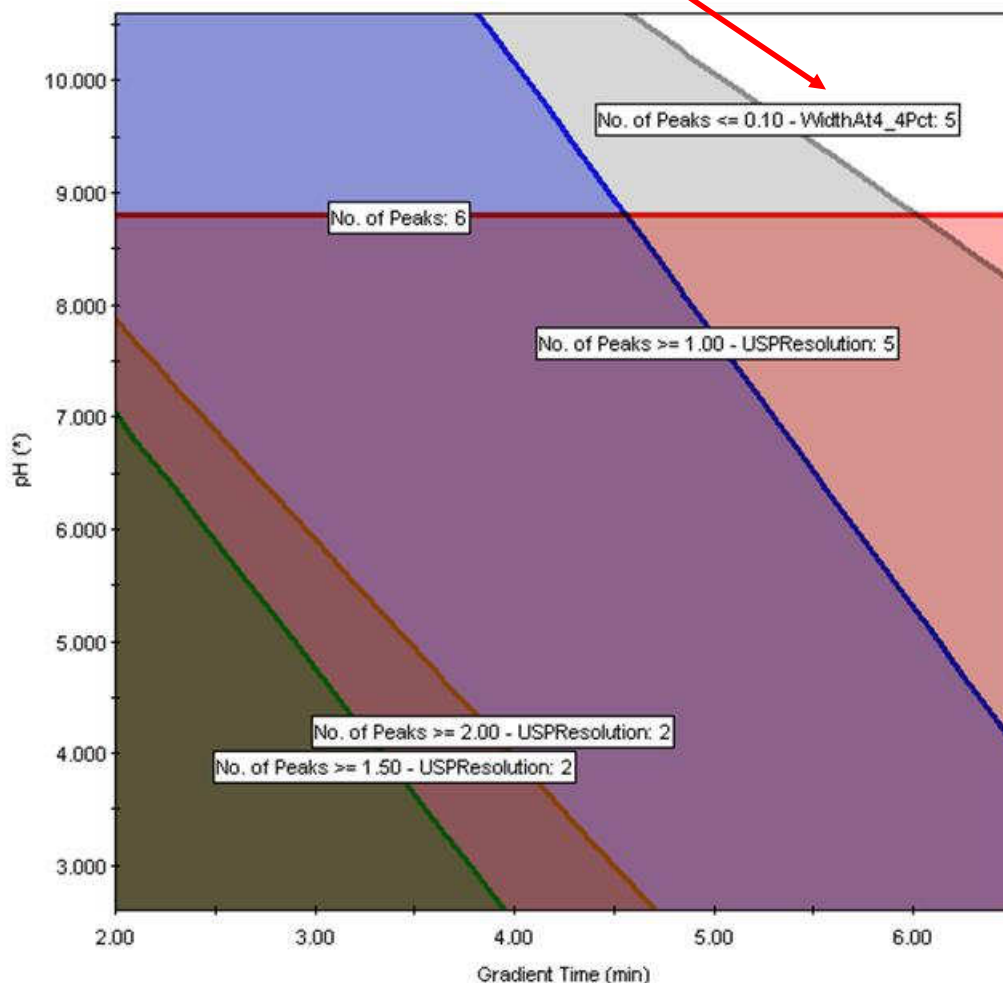
Confidence Limits for the Predicted Response(s) ± 2 Sigma

Study Variable Settings Validation Results
Your settings are valid.

Restore Defaults << Back Finish Cancel ?

Automized Method Optimization: Overlay Plot

White region represents operating region that meets specified success criteria



Optimized Results From Screening Protocol:

ACQUITY CSH C18

Acetonitrile, pH 10.6, $T_g = 5.13$ min

All 28 conditions included in the data analysis returned the same answer

Optimizer Answer #1: 28 of 28

Study Variable Data

Study Variable Name	Optimizer Answer Level Setting
Gradient Time	5.13
Organic Solvent Type	Acetonitrile
pH	10.600
Column Type	CSH HL C18

- Flow rate
 - Set window: 0,2 – 0,7 ml/min
- Gradient end point
 - Set window: 60 – 95 % acetonitrile
- Gradient time
 - Set window: 2 – 6,5 minutes
- Column temperature
 - Set window: 30 – 45 °C

Action:

Using column and mobile phase selections determined from screening protocol, Fusion AE determines an experimental design to optimize secondary effectors of selectivity

Determines interactions between variables including:

- Linear additive effects
- Simple interactions
- Complex interactions

Method Optimization Experimental Design

Run No.	Sample Set No.	Pump Flow Rate (mL/min)	Gradient Time (min)	Final % Organic (%)	Oven Temperature (°C)
Wash - 1	1	0.45	0.1	95	30
1.a.1.a	1	0.7	6.5	95	30
2.a.1.a	1	0.2	6.5	95	30
3.a.1.a	1	0.7	2	95	30
4.a.1.a	1	0.2	2	95	30
5.a.1.a	1	0.7	6.5	60	30
6.a.1.a	1				
7.a.1.a	1				
8.a.1.a	1				
9.a.1.a	1				
10.a.1.a	1				
11.a.1.a	1				
12.a.1.a	1				
13.a.1.a	1				
14.a.1.a	1				
15.a.1.a	1				
16.a.1.a	1				
17.a.1.a	1				
18.a.1.a	1				
19.a.1.a	1				
20.a.1.a	1				
21.a.1.a	1				
22.a.1.a	1				
23.a.1.a	1	0.575	5.4	89	37.5
24.a.1.a	1	0.325	5.4	69	37.5
25.a.1.a	1	0.575	3.1	69	37.5
26.a.1.a	1	0.575	5.4	86	37.5
27.a.1.a	1	0.325	3.1	86	37.5

28.a.1.a	1	0.7	6.5	60	45
29.a.1.a	1	0.2	6.5	60	45
30.a.1.a	1	0.7	2	60	45
31.a.1.a	1	0.2	2	60	45
32.a.1.a	1	0.45	4.3	78	45
33.a.1.a	1	0.7	6.5	95	45
34.a.1.a	1	0.2	6.5	95	45
35.a.1.a	1	0.2	4.3	78	45
36.a.1.a	1	0.45	4.3	78	45
				95	45
				95	45
				86	52.5
				69	52.5
				86	52.5
				60	60
				60	60
				60	60
				60	60
				95	60
				95	60
				78	60
				95	60
				95	60
				78	60
				95	60
				95	60
				95	60
				95	60
54.a.1.a	1	0.7	2	78	60
55.a.1.a	1	0.7	2	95	60
56.a.1.a	1	0.2	2	78	60
57.a.1.a	1	0.2	2	95	60
58.a.1.a	1	0.45	6.5	60	60
59.a.1.a	1	0.45	4.3	78	60
60.a.1.a	1	0.7	4.3	60	60
61.a.1.a	1	0.7	6.5	60	60
62.a.1.a	1	0.2	4.3	60	60
Wash - 2	1	0.45	0.1	60	60

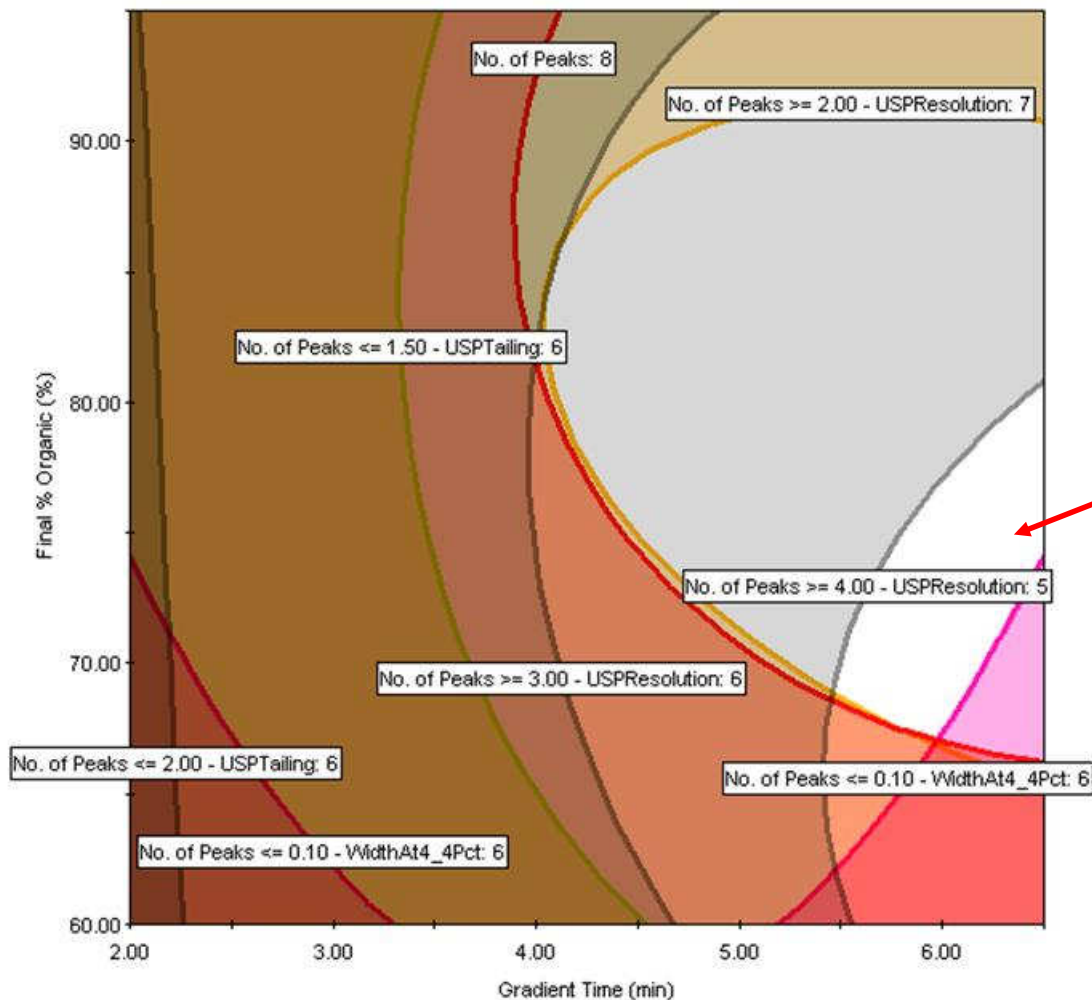
Fusion AE software automatically constructs a set of experiments by selecting the most efficient statistical experimental design

Instrument methods, method sets and sample sets are automatically created in Empower 2 to carry out the experiment

**Sample set run time: < 14 hours
Replicates highlighted**

Automized Method Optimization: Overlay Plot

Control space



The Final Result:

ACQUITY CSH C₁₈
2,1 x 50 mm, 1,7 μ m
 pH 10.6, T_g = 6.5 min, 5 – 78%
 acetonitrile Flow rate = 0.7
 ml/min, T = 45 °C

Optimizer Answer #4: 5 of 35

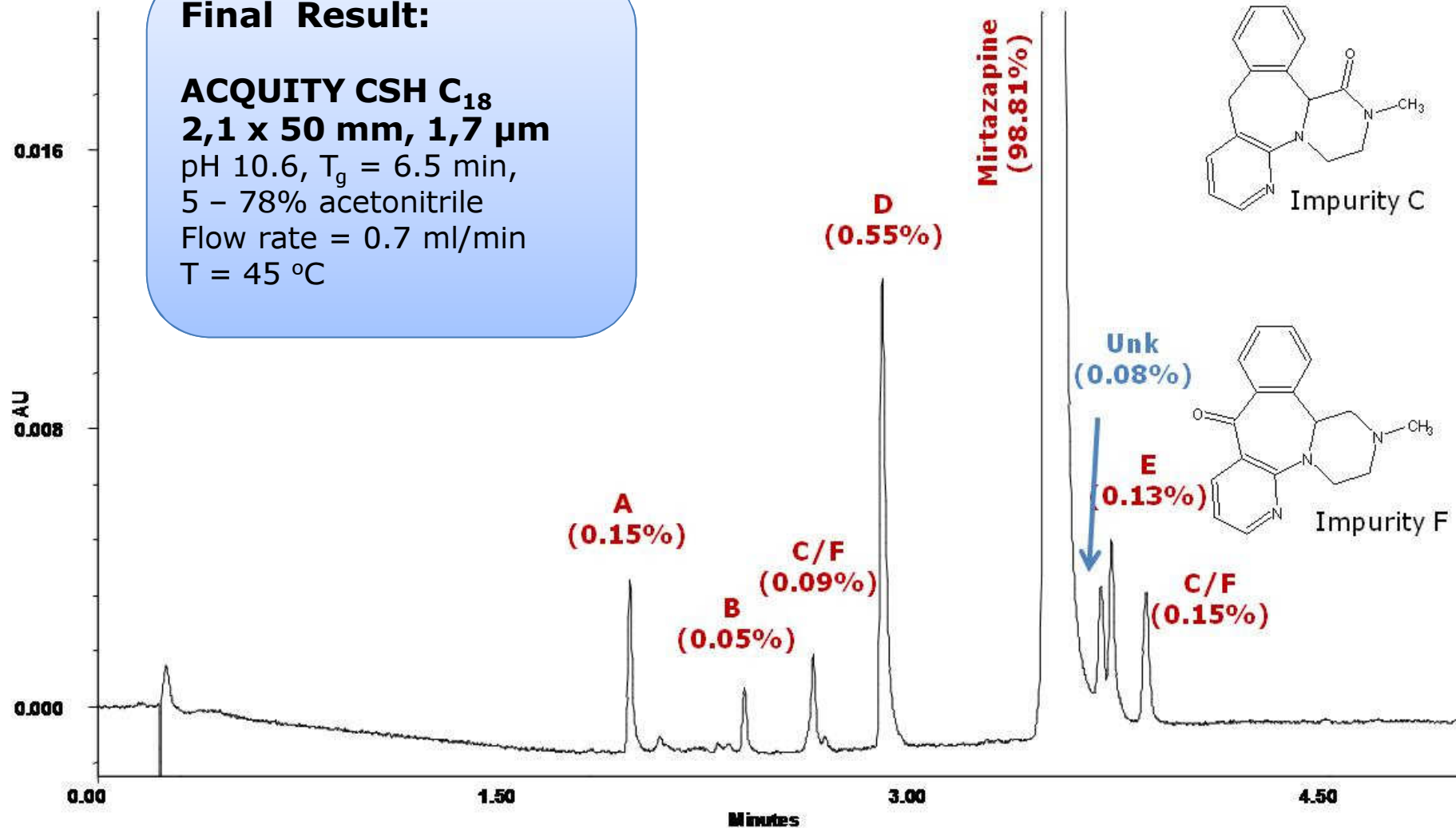
Study Variable Data

Study Variable Name	Optimizer Answer Level Setting
Pump Flow Rate	0.700
Gradient Time	6.50
Final % Organic	77.98
Oven Temperature	45.0

Final Optimized Result: Mirtazapine and Impurities

Final Result:

ACQUITY CSH C₁₈
2,1 x 50 mm, 1,7 μm
pH 10.6, T_g = 6.5 min,
5 – 78% acetonitrile
Flow rate = 0.7 ml/min
T = 45 °C



- **Introduction**
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 - Success Criteria
- **Controlling Selectivity and Retention**
 - Stationary Phase and Particle Substrate Design
 - Organic Modifier
 - Mobile Phase pH
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 - Quality by Design [Q_bD] Approach
- **Implementing the Approach**
 - Case Study
- **Conclusion**

- UPLC Technology facilitates rapid development of robust methods
 - Systematic screening protocol involving pH, organic modifier and column chemistry
 - High resolution sub 2 μm column technology creates high resolution separations, faster
 - Automated column and mobile phase selection
 - Quaternary solvent mixing [ACQUITY UPLC H-Class]
- The principles of method development described here can be implemented for both HPLC and UPLC
 - XSelect HPLC columns and ACQUITY CSH UPLC columns provide a broad range of selectivity [C18, Phenyl-Hexyl and Fluoro-Phenyl] to efficiently develop robust methods
- Combining Fusion AE method development software with UPLC Technology enables a rapid yet comprehensive approach to Q_bD method development
 - Develop robust methods in a matter of days
 - Incorporates robustness modeling into the method development process
 - Aligned with FDA and EMA Q_bD initiatives

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Questions? Comments?