

# MassHunter PFAS MRM Database (for analysis of Per/Polyfluoroalkyl substances)

## Quick Start Guide

What is the MassHunter PFAS MRM Database?	2
Working with MassHunter PFAS MRM Database	2
MassHunter PFAS MRM Database Product Content	3
Optional Products	4
Where to find more information	5
Installation	6
To install the MassHunter PFAS MRM Database files	7
To uninstall MassHunter database files	8
MRM Database Workflow	9
Single Standard Mix Workflow	10
Multiple Standard Mix Workflow	10
Getting Started	11
Step 1. Prepare to use the PFAS database	11
Step 2. Set up LC method parameters	12
Step 3. Set up LC/MS ion source parameters	14
Step 4. Set up the MRM method	17
Step 5. Set up a worklist to run the methods	23
Example Methods	24
System and column	24
G7167B Multisampler	25
G7120A Binary Pump	28
G7116B Column Compartment	30

## What is the MassHunter PFAS MRM Database?

### Working with MassHunter PFAS MRM Database

## What is the MassHunter PFAS MRM Database?

The MassHunter PFAS MRM Database lets you screen 108 native and labeled per- and polyfluoroalkyl substance analytes with up to 4 transitions each with enhanced sensitivity, all in a single LC/MS analysis. The MassHunter PFAS MRM Database and example methods help to minimize development time for your analysis.

The MassHunter PFAS MRM Database contains target compounds for EPA, ASTM, ISO methods, and EU DWD plus emerging PFAS classes methods and regulations.

## Working with MassHunter PFAS MRM Database

The MassHunter PFAS MRM Database helps minimize method development time for your PFAS analysis, when used with the Agilent recommended LC/MS configuration and accessories. It stores up to 4 MRM transitions (a precursor and product ion) per compound of the PFAS analytes included in the database, and their optimized fragmentor and collision energy settings from the Agilent 6400 Series Triple Quadrupole LC/MS and Ultivo LC/TQ instruments. Method development can simply be achieved by importing target compounds from the database to the MassHunter Data Acquisition program.

For some compounds in the MassHunter PFAS MRM Database, several transitions are provided. If you want to convert an MRM method to a dMRM or tMRM method, please refer to the *MassHunter MRM/dMRM/tMRM Database Familiarization Guide*.

# MassHunter PFAS MRM Database Product Content

## MassHunter PFAS MRM Database Quick Start Guide

The Quick Start Guide provides an introduction to the MassHunter PFAS MRM Database.

## MassHunter PFAS MRM Database Media

This media contains:

- PFAS MRM database for 6470B, 6495C, and Ultivo LC/TQ
- Example methods for 6470B, 6495C, and Ultivo LC/TQ
- PFAS example data

## MassHunter MRM Database Familiarization Media

Download the MassHunter MRM Database Familiarization Media from SubscribeNet or get it from your Agilent representative.

This media contains:

- Checkout Mix example data, database, methods, and reports.
- *MassHunter MRM/dMRM/tMRM Database Familiarization Guide*

## Optional Products

### **PFC-free HPLC Conversion Kit (optional)**

Your Agilent Infinity HPLC instrument must be set up to remove the interference of the PFAS compounds leaching from the tubings and solvents. Agilent recommends the use of:

- **PFC-Free HPLC Conversion Kit** (p/n 5004-0006), which includes PTFE-free tubings and a delay column.

### **Checkout Test Mix and Column (optional)**

You can purchase the optional Checkout Test Mix and column to collect your own data to use with the Familiarization exercises. If you purchased the Installation and Familiarization option, these items are included.

- **LC TOF/QTOF/QQQ Pesticide Checkout Test Mix** (p/n 5190-0469), which includes 6 ampoules containing a Pesticides LC/MS Checkout test mix of 20 components in total (3 ampoules of Positive and 3 ampoules of Negative). Instructions to prepare the sample is found in the *MassHunter MRM/dMRM/tMRM Database Familiarization Guide*.
- **Agilent ZORBAX RRHD Eclipse Plus C18, 95Å, 2.1 x 100 mm, 1.8 µm, 1200 bar pressure limit** (p/n 959758-902)

## What is the MassHunter PFAS MRM Database?

Where to find more information

### Where to find more information

**MassHunter MRM/dMRM/tMRM Database Familiarization Guide** Use the exercises in this guide to learn how to use your MassHunter MRM, dMRM, or tMRM Database with MassHunter Data Acquisition, Qualitative Analysis, and Quantitative Analysis programs. You use the example Checkout Mix data, method files, and database to learn how to find and identify compounds in a data file. The Checkout Mix data files, methods, and database are based on the Pesticides Checkout Test Mix, which contains a wide variety of compound classes. Find it in the **\MassHunter\Documents** folder of the Familiarization media, or in the **\MassHunter\Documents** folder on your computer after installation of the Familiarization media.

**Agilent Data Acquisition Familiarization Guide** Use this guide to learn more about tMRM acquisition.

**Agilent web site** Check [www.agilent.com](http://www.agilent.com) for more information. Look for **PFAS MRM Database**.

## Installation

Where to find more information

# Installation

### Before you begin

- Make sure these MassHunter programs are installed.
  - MassHunter Data Acquisition
    - for Ultivo LC/TQ, 1.1 or later
    - for 6470B, 10.1 or later
    - for 6495C, 10.0 SR1 or later
  - MassHunter Quantitative Analysis 10.0 or later.
  - MassHunter Qualitative Analysis 10.0 or later.
- Make sure you know which drive the **MassHunter** folder is installed on your computer. By default, this folder is created on the **D:** drive.

## Installation

To install the MassHunter PFAS MRM Database files

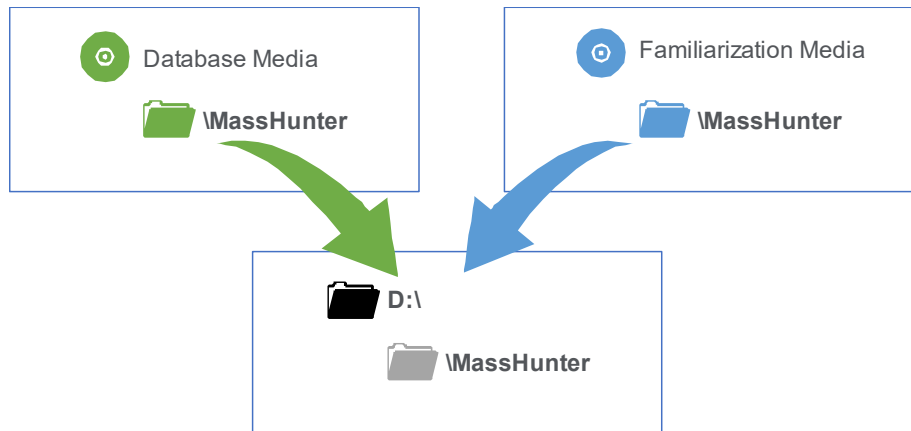
# To install the MassHunter PFAS MRM Database files

- 1 Copy the **\MassHunter** folder into the root folder of your MassHunter drive. By default, the folder is **D:\**.

The **\MassHunter** folder already exists on your computer. Make sure to copy the **\MassHunter** folder from the media to the same location where the **\MassHunter** folder already exists.

- 2 To install the Familiarization files, from the Familiarization media, copy the **\MassHunter** folder into the root folder of your MassHunter drive. By default, the folder is **D:\**.

Check the instructions that come with the Familiarization media for the most current installation instructions.



## Installation

### To uninstall MassHunter database files

## To uninstall MassHunter database files

You need only delete folders and not individual files. The appropriate individual files are removed when the folders are removed.

- To remove the PFAS MRM Database files:
  - a** In the **\MassHunter** folder on your computer, search for **PFAS MRM Database**.
  - b** Delete all folders that contains that term.
- To remove Familiarization files:
  - a** In the **\MassHunter** folder on your computer, search for **Checkout Mix**.
  - b** Delete all folders that contains that term.

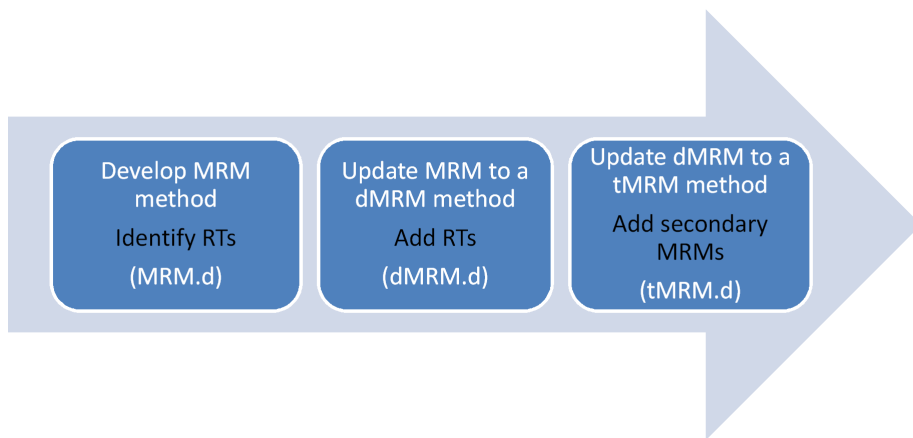


## MRM Database Workflow

To uninstall MassHunter database files

# MRM Database Workflow

**Figure 1** summarizes the workflow, which includes incremental method development from MRM, over to dynamic MRM (dMRM) to triggered MRM (tMRM) methods, including identification of retention times (RT), trigger parameters, and secondary transitions.



**Figure 1.** MRM to dMRM to tMRM Method Development Workflow for single standard mix

Refer to the *MassHunter MRM/dMRM/tMRM Database Familiarization Guide* for more details.

## Single Standard Mix Workflow

You can use this complete workflow to create an MRM, dMRM, or tMRM method to analyze a single standard mix:

- 1 Use the database to create the MRM method for the primary transitions.
- 2 Establish the Retention Times, and then update the MRM method to a dMRM method using the **Update DMRM Method** command. Save as a dMRM method.
- 3 Check the dMRM editor for any overlaps in retention time. If needed, adjust the cycle time settings and/or the Retention Time windows.
- 4 Acquire data to make sure that the dMRM method is valid.
- 5 Update the dMRM method with trigger parameters. Save as a tMRM method.
- 6 Add the secondary transitions.

After you have set up methods to analyze a single standard mix, you can adapt the same procedures for your unique multi-component analysis.

## Multiple Standard Mix Workflow

Some analyses include multiple standard mixes.

To develop a method to analyze multiple compound mixes in one analytical run:

- 1 Create and optimize each dMRM or tMRM method for each standard mix separately. Use the same LC chromatographic method.
- 2 Combine these dMRM or tMRM methods. (Copy and paste transition tables of each dMRM or tMRM method into a single acquisition method.)
- 3 Re-optimize the parameters for overlapping dMRM or tMRM transitions for compounds that co-elute.

For ease of use, optimize no more than 50 compounds at a time in each **MRM -> dMRM -> tMRM** workflow.

## Getting Started

### Step 1. Prepare to use the PFAS database

# Getting Started

This section describes the steps to get started with the MassHunter PFAS MRM Database. In this section, you prepare to use the MassHunter PFAS MRM Database for the first time, set up your method with content from the MassHunter PFAS MRM Database, and then set up a worklist.

The MassHunter PFAS MRM Database product includes two example methods for each model, as described in “**Example Methods**” on page 24. The example methods give you appropriate results only if you use the same HPLC setup that was used to create the methods. Follow the steps in this section to create your own methods with your existing HPLC setup.

## Step 1. Prepare to use the PFAS database

- 1 Before you use the database or run the Familiarization exercises for the first time, do a Preventative Maintenance on your system.
- 2 Install the **PFC-free HPLC Conversion Kit** to prevent the interference of PFAS compounds possibly leaching from tubings and solvents.

### NOTE

If you plan to use the **LC TOF/QTOF/QQQ Pesticide Checkout Test Mix (p/n 5190-0469)** to collect your own checkout data, Agilent recommends that you collect data before you install the **PFC-free HPLC Conversion Kit**. The Familiarization exercises use a different column than the **PFC-free HPLC Conversion Kit**.

## Getting Started

### Step 2. Set up LC method parameters

## Step 2. Set up LC method parameters

This step assumes a **Agilent ZORBAX RRHD Eclipse Plus C18, 95Å, 2.1 x 100 mm, 1.8 µm, 1200 bar pressure limit (p/n 959758-902)**. Adjust your method as needed.

- 1 Prepare your samples and transfer the sample solutions to 1-mL polypropylene vials for analysis.
- 2 Set up the mobile phases. The mobile phases used to create the **Example Methods** are:
  - Solvent A: 5 mM ammonium formate in water
  - Solvent B: methanol
- 3 In the MassHunter Data Acquisition program, open the method of your choice or create a new method.

You can also use the *model\_PFAS\_BaseMethod.m* found in the **\MassHunter\Methods\PFAS MRM Database 1.0** folder. Note that the Base Methods and the Comprehensive Methods are dMRM methods. These dMRM methods contain Retention Times based on instrument setup as described in **“Example Methods”** on page 24. If your instrument setup is different, expected Retention Times may differ from those in the Base Methods.

If you start with one of the **Example Methods**, you need to save the method to a different name.

- 4 Check that the method is set up to make the appropriate injection volume. The default injection volumes used in the **Example Methods** are:
  - Ultivo LC/TQ: 5 µL
  - 6470B LC/TQ: 3 µL
  - 6495C LC/TQ: 2 µL
- 5 Set up the gradient.

See **Figure 2** on page 13 for the gradient used in the **Example Methods**. The Stop time is 14.5 minutes with a post time of 2.5 minutes.
- 6 Set the Column Compartment temperature. The temperature used to create the **Example Methods** is 55°C.

## Getting Started

### Step 2. Set up LC method parameters

Time [min]	$\Delta$	A [%]	B [%]	Flow [mL/min]	Max. Pressure Limit [bar]
0.00		85.00	15.00	0.400	1000.00
1.00		85.00	15.00	0.400	---
1.50		45.00	55.00	0.400	---
5.50		30.00	70.00	0.400	---
7.00		20.00	80.00	0.400	---
12.00		0.00	100.00	0.400	---
14.40		0.00	100.00	0.400	---
14.50		85.00	15.00	0.400	---

**Figure 2.** Gradient for 1290 Infinity II LC system with Agilent Eclipse Plus C18, 2.1 mm × 100 mm, 1.8- $\mu$ m ZORBAX LC column (p/n 959758-902)

## Getting Started


### Step 3. Set up LC/MS ion source parameters

## Step 3. Set up LC/MS ion source parameters

- Set up the ion source parameters in the MS part of the method tab.  
For a multicomponent method, the ion source parameters shown in the next figures are used to achieve the best overall sensitivity for all compounds in the database for each instrument. You can adjust the method to optimize for individual or selected compounds.

### Ion source parameters for Ultivo QQQ with an Agilent Jet Stream source

#### Source Parameters



##### Global Parameters

	Setpoint	Actuals
Gas Temperature (°C)	<input type="text" value="230"/>	
Gas Flow (L/min)	<input type="text" value="5.0"/>	
Nebulizer (psi)	<input type="text" value="15"/>	
Sheath Gas Temperature (°C)	<input type="text" value="350"/>	
Sheath Gas Flow (L/min)	<input type="text" value="11.0"/>	

#### Per Time Segment

	Positive Setpoint	Negative Setpoint	Actuals
Capillary Voltage (V)	<input type="text" value="0"/>	<input type="text" value="2500"/>	
Nozzle Voltage (V)	<input type="text" value="0"/>	<input type="text" value="0"/>	

## Getting Started

### Step 3. Set up LC/MS ion source parameters

#### Ion source parameters for 6470B with an Agilent Jet Stream source

Source parameters

Gas Temp:	<input type="text" value="230"/>	°C	<input type="text"/>	°C
Gas Flow:	<input type="text" value="6"/>	l/min		
Nebulizer:	<input type="text" value="20"/>	psi	<input type="text"/>	psi
Sheath Gas Temp:	<input type="text" value="375"/>	°C		
Sheath Gas Flow:	<input type="text" value="12"/>	l/min		
	Positive		Negative	
Capillary:	<input type="text" value="0"/>	V	<input type="text" value="2500"/>	V
			<input type="text"/>	V
Nozzle Voltage:	<input type="text" value="0"/>	V	<input type="text" value="0"/>	V

Under **Time segments**, make sure that **Delta EMV (-)** is set to **200**.

## Getting Started

### Step 3. Set up LC/MS ion source parameters

#### Ion source parameters for 6495C QQQ with an Agilent Jet Stream source

Source parameters			
Gas Temp:	<input type="text" value="250"/>	°C	<input type="text"/>
Gas Flow:	<input type="text" value="11"/>	l/min	<input type="text"/>
Nebulizer:	<input type="text" value="25"/>	psi	<input type="text"/>
Sheath Gas Temp:	<input type="text" value="375"/>	°C	
Sheath Gas Flow:	<input type="text" value="11"/>	l/min	
	Positive	Negative	
Capillary:	<input type="text" value="0"/> V	<input type="text" value="3000"/> V	<input type="text"/>
Nozzle Voltage:	<input type="text" value="0"/> V	<input type="text" value="0"/> V	<input type="text"/>

iFunnel parameters			
	Positive	Negative	
High Pressure RF	<input type="text" value="100"/> V	<input type="text" value="90"/> V	
Low Pressure RF	<input type="text" value="110"/> V	<input type="text" value="80"/> V	

Under **Time segments**, make sure that **Delta EMV (-)** is set to **200**.

For the 6495C, the **Source** tab includes iFunnel parameters. These iFunnel parameters ensure the best overall sensitivity for PFAS compounds. You can use the **Source and iFunnel Optimizer** program to optimize for individual or selected compounds. Refer to the MassHunter Data Acquisition *online Help*.



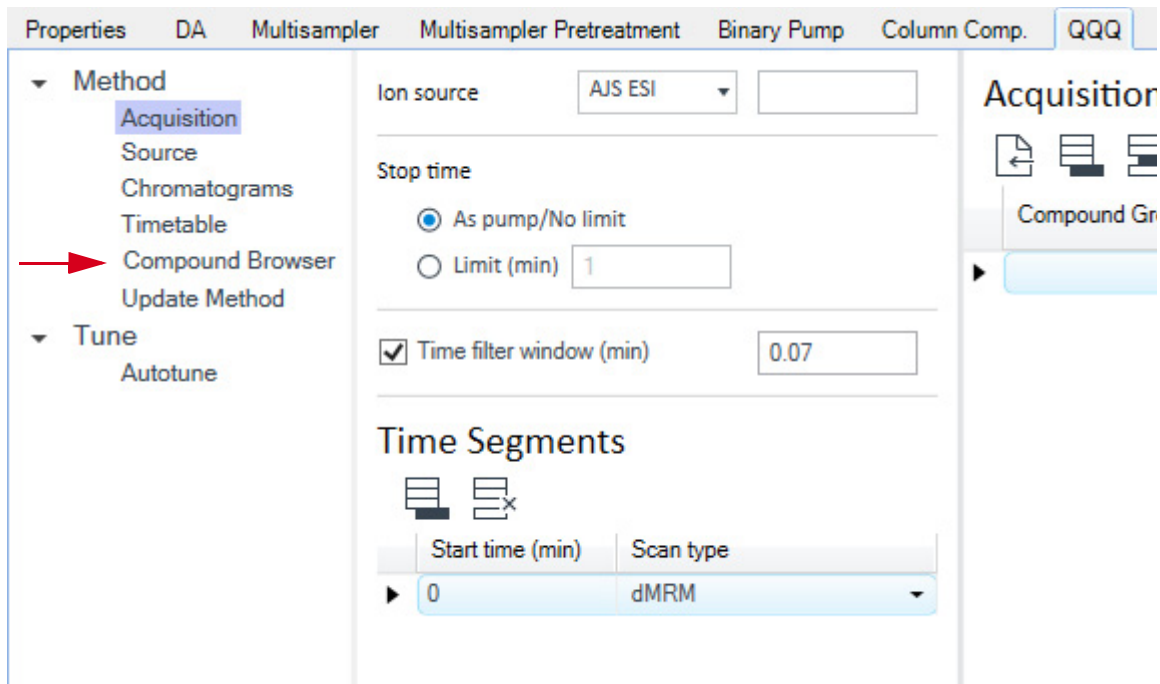
## Getting Started

### Step 4. Set up the MRM method

## Step 4. Set up the MRM method

### Ultivo LC/TQ

- 1 In the Method Editor window, click **QQQ > Acquisition**.



- 2 Select **Compound Browser**.
- 3 In the Compound Browser, click **File > Open Database**.
- 4 From the folder **\\MassHunter\Databases\PFAS MRM Database 1.0**, open **PFAS\_MRM\_Database\_Ultivo\_1.1**. Click **OK**.

## Getting Started

### Step 4. Set up the MRM method

- 5 Select the list of transitions from the database:
  - a Check the box next to each **Compound Name** of interest.
  - b Click **Add to Import List**.


<input type="checkbox"/>	Compound Name	IUPAC Name	Other Names	CAS	Formula	ChemSpider	Groups	Species
<input checked="" type="checkbox"/>	PFBA	Heptafluorobutan	PFBA / Perfluorob	375-22-4	C4HF7O2	9394	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFPeA	Nonafluoropentan	PFPeA / Perfluoro	2706-90-3	C5HF9O2	68426	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHxA	Undecafluorohex	PFHxA / Perfluoro	307-24-4	C6HF11O2	60864	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHxA	Undecafluorohex	PFHxA / Perfluoro	307-24-4	C6HF11O2	60864	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHpA	Tridecafluorohept	PFHpA / Perfluor	375-85-9	C7HF13O2	61135	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHpA	Tridecafluorohept	PFHpA / Perfluor	375-85-9	C7HF13O2	61135	PFCA	(M-H)-
<input type="checkbox"/>	PFOA	Pentadecafluoroo	PFOA / Perfluoro	335-67-1	C8HF15O2	9180	PFCA	(M-H)-
<input type="checkbox"/>	PFOA	Pentadecafluoroo	PFOA / Perfluoro	335-67-1	C8HF15O2	9180	PFCA	(M-H)-

Current Database : D:\MassHunter\Databases\PFAS MRM Database Ultivo (ReadOnly) Add to Import List

- c Click **Import**.

Compound Browser

File Edit View



Search/Filter Import List

Compound Name	Formula	MW	Polarity	Species	Precursor	Product	Frag	CE	Primary	Trigger	RT	RT Wind
▶ PFBA	C4HF7O2		Negative	(M-H)-	213	169	60	4	<input checked="" type="checkbox"/>	<input type="checkbox"/>	3.05	1
PFPeA	C5HF9O2		Negative	(M-H)-	263	219	60	4	<input checked="" type="checkbox"/>	<input type="checkbox"/>	3.5	1
PFHxA	C6HF11O2		Negative	(M-H)-	313	269	80	4	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.02	1
PFHxA	C6HF11O2		Negative	(M-H)-	313	119	80	24	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.02	1
PFHpA	C7HF13O2		Negative	(M-H)-	363	319	80	4	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.71	1
PFHpA	C7HF13O2		Negative	(M-H)-	363	169	80	16	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.71	1

Import

- 6 Save the method.

If you started with a PFAS example Base Method, save the method to a different name.

Refer to the *MassHunter MRM/dMRM/tMRM Database Familiarization Guide* for more details.

## Getting Started

### Step 4. Set up the MRM method

#### 6470B

- 1 In the Method Editor window, click **QQQ > Acquisition**.
- 2 Right-click the scan segment table and click **Import from Database Browser**.

The screenshot shows the Agilent MassHunter Acquisition Method Editor interface. The 'Acquisition' tab is selected, and the 'Scan segments' table is visible. A context menu is open over the table, with 'Import from Database Browser...' highlighted. The table contains three rows of scan segments.

#	Start Time	Scan Type	Div Valve	Delta EMV (+)	Delta EMV (-)	Stored
1	0	Dynamic MRM	To Waste	0	0	<input checked="" type="checkbox"/>
2	0.65	Dynamic MRM	To MS	0	200	<input checked="" type="checkbox"/>
3	12	Dynamic MRM	To Waste	0	0	<input checked="" type="checkbox"/>

- 3 In the Database Browser, click **File > Open Database**.
- 4 From the folder **\\MassHunter\Databases\PFAS MRM Database 1.0**, open **PFAS\_MRM\_Database\_6470B\_10.1**. Click **OK**.

## Getting Started

### Step 4. Set up the MRM method

- 5 Select the list of transitions from the database:
  - a Check the box next to each **Compound Name** of interest.
  - b Click **Add to Import List**.

<input type="checkbox"/>	Compound Name	IUPAC Name	Other Names	CAS	Formula	ChemSpider	Groups	Species
<input checked="" type="checkbox"/>	PFBA	Heptafluorobutan	PFBA / Perfluorob	375-22-4	C4HF7O2	9394	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFPeA	Nonafluoropentan	PFPeA / Perfluoro	2706-90-3	C5HF9O2	68426	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHxA	Undecafluorohex	PFHxA / Perfluoro	307-24-4	C6HF11O2	60864	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHxA	Undecafluorohex	PFHxA / Perfluoro	307-24-4	C6HF11O2	60864	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHpA	Tridecafluorohept	PFHpA / Perfluor	375-85-9	C7HF13O2	61135	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHpA	Tridecafluorohept	PFHpA / Perfluor	375-85-9	C7HF13O2	61135	PFCA	(M-H)-

Current Database : D:\MassHunter\Databases\PFAS MRM Database 6470 (ReadOnly) Add to Import List

- c Click **Import**.

Search/Filter

Compound Name	Formula	MW	Polarity	Species	Precursor	Product	Frag	CE	Primary	Trigger	RT	RT
PFBA	C4HF7O2		Negative	(M-H)-	213	169	72	8	<input checked="" type="checkbox"/>	<input type="checkbox"/>	3.06	1
PFPeA	C5HF9O2		Negative	(M-H)-	263	219	72	4	<input checked="" type="checkbox"/>	<input type="checkbox"/>	3.51	1
PFHxA	C6HF11O2		Negative	(M-H)-	313	269	72	8	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.01	1
PFHxA	C6HF11O2		Negative	(M-H)-	313	119	72	24	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.01	1
PFHpA	C7HF13O2		Negative	(M-H)-	363	319	72	8	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.68	1
PFHpA	C7HF13O2		Negative	(M-H)-	363	169	72	16	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.68	1

Import Close

- 6 Save the method.

If you started with a PFAS example Base Method, save the method to a different name.

Refer to the *MassHunter MRM/dMRM/tMRM Database Familiarization Guide* for more details.

## Getting Started

### Step 4. Set up the MRM method

#### 6495C

- 1 In the Method Editor window, click **QQQ > Acquisition**.
- 2 Right-click the scan segment table and click **Import from Database Browser**.

The screenshot shows the Agilent MassHunter Acquisition Method Editor interface. The 'Acquisition' tab is selected, and the 'Scan segments' table is visible. A context menu is open over the table, with 'Import from Database Browser...' highlighted. The table contains three rows of scan segments.

#	Start Time	Scan Type	Div Valve	Delta EMV (+)	Delta EMV (-)	Stored
1	0	Dynamic MRM	To Waste	0	0	<input checked="" type="checkbox"/>
2	0.65	Dynamic MRM	To MS	0	200	<input checked="" type="checkbox"/>
3	12	Dynamic MRM	To Waste	0	0	<input checked="" type="checkbox"/>

- 3 In the Database Browser, click **File > Open Database**.
- 4 From the folder **\\MassHunter\Databases\PFAS MRM Database 1.0**, open **PFAS\_MRM\_Database\_6495C\_10.0**. Click **OK**.

## Getting Started

### Step 4. Set up the MRM method

- 5 Select the list of transitions from the database:
  - a Check the box next to each **Compound Name** of interest.
  - b Click **Add to Import List**.

<input type="checkbox"/>	Compound Name	IUPAC Name	Other Names	CAS	Formula	ChemSpider	Groups	Species
<input checked="" type="checkbox"/>	PFBA	Heptafluorobutan	PFBA / Perfluorob	375-22-4	C4HF7O2	9394	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFPeA	Nonafluoropentan	PFPeA / Perfluoro	2706-90-3	C5HF9O2	68426	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHxA	Undecafluorohex	PFHxA / Perfluoro	307-24-4	C6HF11O2	60864	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHxA	Undecafluorohex	PFHxA / Perfluoro	307-24-4	C6HF11O2	60864	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHpA	Tridecafluorohept	PFHpA / Perfluor	375-85-9	C7HF13O2	61135	PFCA	(M-H)-
<input checked="" type="checkbox"/>	PFHpA	Tridecafluorohept	PFHpA / Perfluor	375-85-9	C7HF13O2	61135	PFCA	(M-H)-

Current Database : D:\MassHunter\Databases\PFAS MRM Database 6495 (ReadOnly) Add to Import List

- c Click **Import**.

Search/Filter		Import List											
Compound Name	Formula	MW	Polarity	Species	Precursor	Product	Frag	CE	Primary	Trigger	RT	RT	
PFBA	C4HF7O2		Negative	(M-H)-	213	169	166	7	<input checked="" type="checkbox"/>	<input type="checkbox"/>	3.08	1	
PFPeA	C5HF9O2		Negative	(M-H)-	263	219	166	7	<input checked="" type="checkbox"/>	<input type="checkbox"/>	3.53	1	
PFHxA	C6HF11O2		Negative	(M-H)-	313	269	166	7	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.01	1	
PFHxA	C6HF11O2		Negative	(M-H)-	313	119	166	23	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.01	1	
PFHpA	C7HF13O2		Negative	(M-H)-	363	319	166	7	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.7	1	
PFHpA	C7HF13O2		Negative	(M-H)-	363	169	166	19	<input checked="" type="checkbox"/>	<input type="checkbox"/>	4.7	1	

Import Close

- 6 Save the method.

If you started with a PFAS example Base Method, save the method to a different name.

Refer to the *MassHunter MRM/dMRM/tMRM Database Familiarization Guide* for more details.

## Getting Started

### Step 5. Set up a worklist to run the methods

## Step 5. Set up a worklist to run the methods

- Set up the worklist as needed for your samples. An example is shown in **Figure 3**.

	✓	Sample Name	Sample Position	Method	Data File	Sample Type	Level Name	Inj Vol (µl)
1	✓	No_Injection	No Injection	6470B_PFAS_Comprehensive.m	No_Injection_01.d	Blank		As Method
2	✓	Solvent_Blank	P1-A1	6470B_PFAS_Comprehensive.m	Solvent_Blank.d	Blank		As Method
3	✓	CAL 1	P1-A2	6470B_PFAS_Comprehensive.m	CAL 1.d	Calibration	1	As Method
4	✓	CAL 2	P1-A3	6470B_PFAS_Comprehensive.m	CAL 2.d	Calibration	2	As Method
5	✓	CAL 3	P1-A4	6470B_PFAS_Comprehensive.m	CAL 3.d	Calibration	3	As Method
6	✓	CAL 4	P1-A5	6470B_PFAS_Comprehensive.m	CAL 4.d	Calibration	4	As Method
7	✓	CAL 5	P1-A6	6470B_PFAS_Comprehensive.m	CAL 5.d	Calibration	5	As Method

Figure 3. Worklist

The Base Methods are dMRM methods, but if you start with your own MRM method, then you can create dMRM and tMRM methods from the MRM method and optimize the parameters. Follow the steps in the *MassHunter MRM/dMRM/tMRM Database Familiarization Guide*.

Refer to the *MassHunter Data Acquisition Familiarization Guide* or *online Help* for more information.

### NOTE

If needed, increase the Delta RT for compounds that have isomers or a shift in retention time.

In a standard method development workflow, the trigger parameters in the tMRM methods such as Threshold, Trigger Entrance, Trigger Delay and Trigger Window are created for analysis of standards in solvent. These trigger parameters must be rechecked for matrix samples as signal abundances and noise characteristics of some MRM transitions can be different in samples with complex matrix compared to solvent standards.

## Example Methods

The MassHunter PFAS MRM Database includes two example methods, the PFAS Base Method and the PFAS Comprehensive Method.

The **PFAS Base Method** includes empty scan segment table to allow import of selected analytes from the database.

The **PFAS Comprehensive Method** includes the MRM settings for all the analytes from the database.

You can find these methods in the **\\MassHunter\\Methods\\PFAS MRM Database 1.0** folder on your computer (post-installation) and the MassHunter PFAS MRM Database media.

Use these methods as a basis for your own analysis, or create your own methods using the steps in **“Getting Started”** on page 11.

For more detailed instructions, see the *MassHunter MRM/dMRM/tMRM Database Familiarization Guide*, and the *MassHunter Data Acquisition Familiarization Guide* and *online Help*.

This topic describes the LC parameters that were used to create the example methods. The MS parameters are described in **“Step 3. Set up LC/MS ion source parameters”** on page 14.

## System and column

Component	Description
HPLC System	1290 Infinity II LC System
Conversion kit	PFC-free HPLC Conversion Kit (p/n 5004-0006)
LC column	ZORBAX RRHD Eclipse Plus C18, 95Å, 2.1 x 100 mm, 1.8 µm, 1200 bar pressure limit (p/n 959758-902)
Guard column	ZORBAX RRHD Eclipse Plus C18, 2.1 mm, 1.8 µm, 1200 bar pressure limit, UHPLC guard (p/n 821725-901)



## G7167B Multisampler

Table 1. G7167B Multisampler parameters

Parameter	Value																									
Thermostat temperature	5°C																									
Injection volume	Ultivo LC/TQ: 5 µL 6470B LC/TQ: 3 µL 6495C LC/TQ: 2 µL																									
Draw speed	200 µL/min																									
Eject speed	400 µL/min																									
Wait time after draw	2.0 s																									
Needle height offset	0.0 mm																									
Use vial/well bottom sensing	No																									
Needle wash	Multi-wash																									
Wash solvent 1 (S1)	15:85 methanol:water																									
Wash solvent 2 (S2)	1:1 acetonitrile:2-propanol																									
Multi-wash program	<table border="1"> <thead> <tr> <th>Step</th> <th>Solvent</th> <th>Time (s)</th> <th>Seat Back Flush</th> <th>Needle wash</th> </tr> </thead> <tbody> <tr> <td>1</td> <td>S1</td> <td>20</td> <td><input checked="" type="checkbox"/></td> <td><input checked="" type="checkbox"/></td> </tr> <tr> <td>2</td> <td>S2</td> <td>20</td> <td><input checked="" type="checkbox"/></td> <td><input checked="" type="checkbox"/></td> </tr> <tr> <td>3</td> <td>Off</td> <td>—</td> <td><input type="checkbox"/></td> <td><input type="checkbox"/></td> </tr> <tr> <td>4</td> <td>S1</td> <td>—</td> <td><input checked="" type="checkbox"/></td> <td><input type="checkbox"/></td> </tr> </tbody> </table>	Step	Solvent	Time (s)	Seat Back Flush	Needle wash	1	S1	20	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	2	S2	20	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	3	Off	—	<input type="checkbox"/>	<input type="checkbox"/>	4	S1	—	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Step	Solvent	Time (s)	Seat Back Flush	Needle wash																						
1	S1	20	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>																						
2	S2	20	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>																						
3	Off	—	<input type="checkbox"/>	<input type="checkbox"/>																						
4	S1	—	<input checked="" type="checkbox"/>	<input type="checkbox"/>																						
Stop time	As Pump/No Limit																									
Post time	Off																									

These parameters are shown in the Method Editor window in the next two images.

Note that the **Injection volume** shown is for the 6470B. The volume varies with the instrument model, as shown in [Table 1](#).

## Example Methods

### G7167B Multisampler

Agilent MassHunter Acquisition Method Editor

File Tools Method Help

6470B\_PFAS\_Comprehensive.m

Properties DA **Multisampler** Multisampler Pretreatment Binary Pump Column Comp. QQQ

### Multisampler (G7167B)

**Injection**

Injection volume: 3.00 µL

**Needle Wash**

Multi-wash

**Stop/Posttime**

As Pump/No Limit  Off

1.00 min  1.00 min

**Advanced**

**Sampling Speed**

Draw Speed: 200.0 µL/min

Eject Speed: 400.0 µL/min

Wait Time After Draw: 2.0 s

**Needle Height Position**

Offset: 0.0 mm

Use Vial/Well Bottom Sensing

**High Throughput**

Sample Flush-Out Factor: 5.0

Injection Valve to Bypass for Delay Volume Reduction

Enable Overlapped Injection

When Sample is Flushed Out

After Period of Time

0.00 minutes after injection

**Injection Path Cleaning**

## Example Methods

### G7167B Multisampler

Agilent MassHunter Acquisition Method Editor

File Tools Method Help

6470B\_PFAS\_Comprehensive.m

Properties DA **Multisampler** Multisampler Pretreatment Binary Pump Column Comp. QQQ

### Multisampler (G7167B)

**Injection**

Injection volume: 3.00  $\mu$ L

**Needle Wash**

Multi-wash

**Stoptime** **Posttime**

As Pump/No Limit       Off  
 1.00 min       1.00 min

**Advanced**

**Injection Path Cleaning**

**Standard Wash**

Mode: Flush Port  
 Time: 3 s  
 Location:  
 Repeat: 3

**Multi-wash**

Step	Solvent	Time [s]	Seat Back Flush	Needle Wash	Comment
1	S1	20	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	15:85/MeOH:Water
2	S2	20	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	1:1/ACN:IPA
3	Off		<input type="checkbox"/>	<input type="checkbox"/>	
Start Cond.	S1		<input checked="" type="checkbox"/>	<input type="checkbox"/>	

## G7120A Binary Pump

Table 2. G7120A Binary Pump parameters

Parameter	Value		
Flow rate	0.4 mL/min		
Solvent A	5 mM ammonium acetate in water		
Solvent B	methanol		
Minimum pressure limit	0 bar		
Maximum pressure limit	1000 bar		
Stop time	14.5 min		
Post time	2.5 min		
Timetable	Time (min)	A (%)	B (%)
	0.00	85	15
	1.00	85	15
	1.50	45	55
	5.50	30	70
	7.00	20	80
	12.00	0	100
	14.40	0	100
	14.50	85	15

These parameters are shown in the Method Editor window in the next image.

# Example Methods

## G7120A Binary Pump

Agilent MassHunter Acquisition Method Editor

File Tools Method Help

6470B\_PFAS\_Comprehensive.m

Properties DA Multisampler Multisampler Pretreatment **Binary Pump** Column Comp. QQQ

### Binary Pump (G7120A)

Flow: 0.400 mL/min

Solvents

A: 1 85.00 % 100.0 % Water V.03 +5 mM ammo  
 2 10.0 % ACN in Water V.02

B: 15.00 % 1 100.0 % Methanol V.03  
 2 50.0 % Acetonitrile in Meth

Pressure Limits  
 Min: 0.00 bar Max: 1,000.00 bar

Stoptime Posttime  
 As Injector/No Limit  Off  
 14.50 min  2.50 min

Advanced

Timetable (14/100 events)

function centric view

Time [min]	A [%]	B [%]	Flow [mL/min]	Max. Pressure Limit [bar]
0.00	85.00	15.00	0.400	1000.00
1.00	85.00	15.00	0.400	---
1.50	45.00	55.00	0.400	---
5.50	30.00	70.00	0.400	---
7.00	20.00	80.00	0.400	---
12.00	0.00	100.00	0.400	---
14.40	0.00	100.00	0.400	---
14.50	85.00	15.00	0.400	---

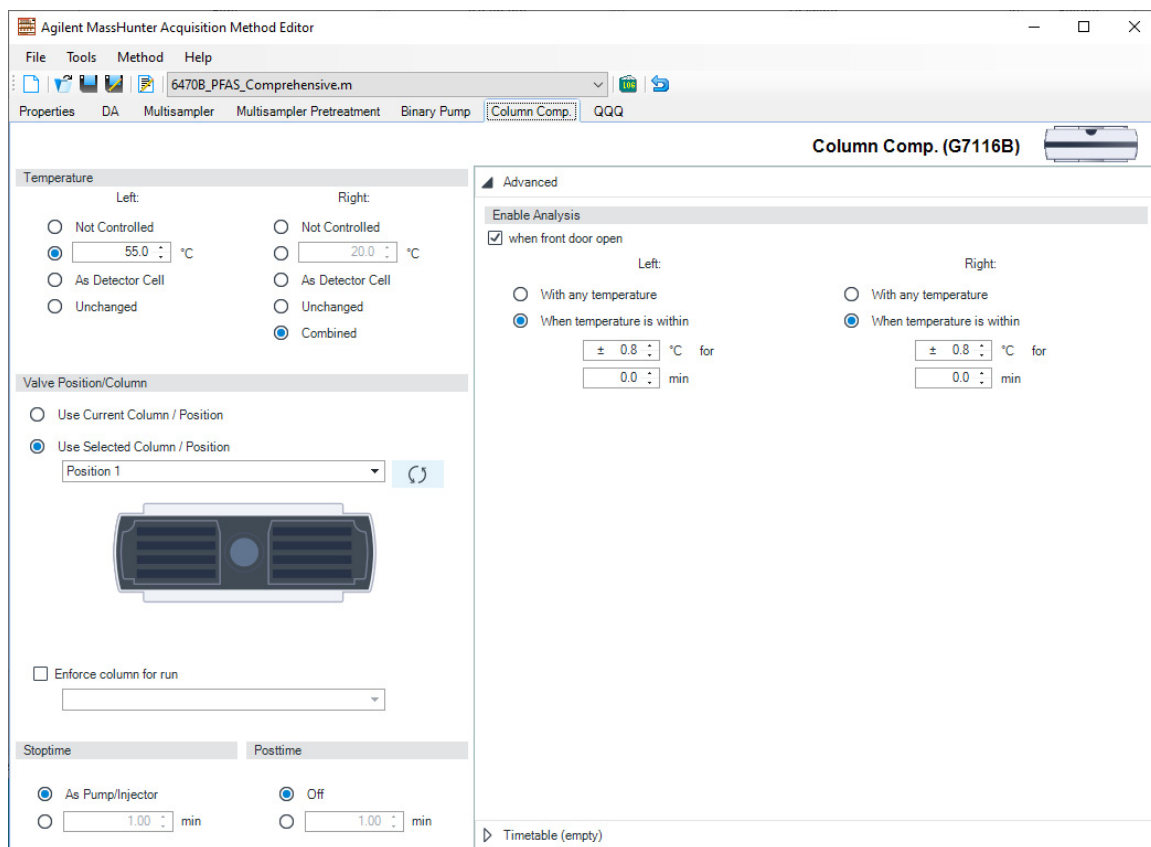
Add Remove Clear All Clear Empty  
 Cut Copy Paste Shift Times 0.00 min

## G7116B Column Compartment

Table 3. G7116B Column Compartment parameters

Parameter	Value
Temperature	Left: 55°C Right: Combined
Temperature allowance	Left and Right: ± 0.8°C

These parameters are shown in the Method Editor window in the next image.



This page intentionally left blank.

## In This Guide

This Quick Start Guide provides an introduction to the MassHunter PFAS MRM Database. This guide applies to PFAS MRM Database 1.0, and includes these database versions:

- PFAS\_MRM\_Database\_6470B\_10.1
- PFAS\_MRM\_Database\_6495C\_10.0
- PFAS\_MRM\_Database\_Ultivo\_1.1

[www.agilent.com](http://www.agilent.com)

© Agilent Technologies, Inc. 2020



G1736-90000

Printed in USA

December 2020 Revision A.01

