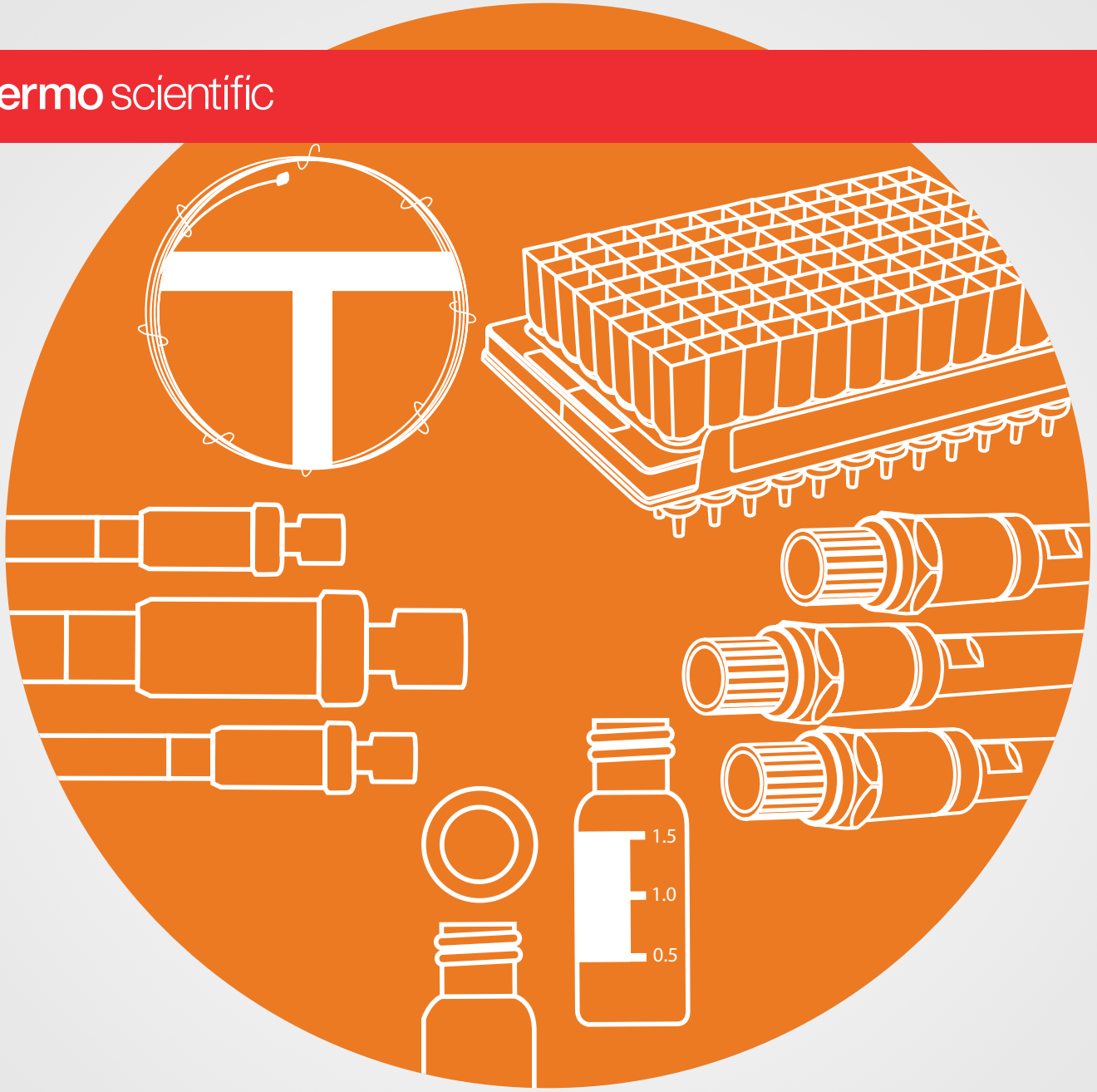
thermo scientific



GC columns and accessories

Technical resources document

ThermoFisher SCIENTIFIC

Contents

GC column selection

When selecting a GC column for your analysis, it can often be difficult to choose the most appropriate column because of the wide range of options. However, the choice can be simplified by considering a number of questions about the planned separation. This section provides useful information to help you determine the most suitable column for your analysis.

Column selection for existing or regulated methods

This section provides a number of tools to aid in selecting the most appropriate Thermo Scientific GC column. The Thermo Scientific GC column phase table lists details for the wide array of phases offered in the Thermo Scientific™ TraceGOLD™, Thermo Scientific™ TRACE™ and Thermo Scientific™ TracePLOT GC™ column ranges. The GC column selection by manufacturer table provides a quick cross reference for Thermo Scientific columns to other GC column manufacturers. If you are following an ASTM, NIOSH or US EPA method, please refer to the column selection by method tables for the best Thermo Scientific product.

Method development considerations

When first developing a method, you should consider these column characteristics to determine the best column for the separation:

A. Column phase
B. Internal diameter
C. Film thickness
D. Column length

A. Column phase

In GC, the separation of two analytes occurs due to differences in their interaction with the stationary phase, therefore a phase must be chosen that matches the properties of the sample. For example, if the components have different boiling points (greater than 2°C), a non-polar column such as the TG-1MS is recommended. If the products differ primarily in their polarities, then a polar column such as the TG-WaxMS will be ideal.

If you know the particular class of your sample, please refer to the column selection by application for a recommended phase (see page 11). Always select the least polar column which will perform the separation.

B. Internal diameter

The selection of the internal diameter is often determined by the instrument or detection method. Most modern GC equipment will accommodate most column sizes. With a larger internal diameter, column sample capacity increases, but resolution and sensitivity decrease. Conversely, a smaller ID column can improve resolution and sensitivity, but with the drawback of reduced sample capacity and a greater need for sample preparation. It is a good idea to find a similar application which gives separation of the desired components and use this as a guide.

C. Film thickness

Increasing the film thickness increases the sample capacity of the column and slows the elution of the peaks which can help when analyzing volatile compounds. A thicker film also reduces the potential of overloading the column, thus improving the resolution. However, a thicker film can be more sensitive to degradation. The same component will elute at a higher temperature on a thick film when compared to a thin film.

Compounds with high boiling points or those with a high molecular weight should be analyzed using a thin film to improve resolution and avoid unnecessarily long analysis times.

Another factor to consider is the phase ratio (β) which is calculated using both the internal diameter and film thickness in the following equation:

$$\beta = \frac{\text{Internal diameter (µm)}}{4 \times \text{Film thickness (µm)}}$$

The phase ratio can be used in two ways:

1. To categorize the best dimensions for an application:
   a. For volatile samples β < 100
   b. For general samples β ~ 250
   c. For high molecular weight samples β > 400
2. To transfer an analysis from a column of one ID to another without changing the method substantially, choose a column with a similar b value as this will have similar retention properties.

D. Column length

Internal Diameter (mm)	Film Thickness (µm)					
	0.1	0.25	0.5	1	1.8	3
0.1	250	100	50	25	14	8
0.25	625	250	125	63	35	21
0.32	800	320	160	80	44	27
0.53	1325	530	265	133	74	44

Phase ratio (β) of common column dimensions

A longer column length will provide greater efficiency and resolution, but this is not a linear relationship. Resolution is proportional to the square root of column length, so doubling the column length will increase resolution by approximately 40%. However, increasing the column length will also increase the retention time. Double column length, twice the analysis time. Generally, it is recommended to use the shortest column which will perform the desired separation.

Additional considerations

Several generalizations regarding GC columns exist that you might rely on when in doubt. First, 95% of all GC columns used are either TG-1MS, TG-5MS or TG-WaxMS type columns. A good starting column is a 30m x 0.25mm ID, 5% Phenyl column with a 0.25µm film thickness, such as the TG-5MS. (Part number 26098-1420).

This is a non-polar column, which separates predominately on boiling point, but has some polar characteristics.

GC column phase information

Range	Column	Phase	Polarity	Max operating temp °C (isothermal / programmable)
TraceGOLD	TG-1MS	100% Methylpolysiloxane	Non-polar	330°C / 350°C
	TG-XLBMS	Proprietary	Non-polar	360°C
	TG-5MS	5% Phenyl Methylpolysiloxane	Non-polar	330°C / 350°C
	TG-SQC	Proprietary	Non-polar	330°C / 350°C
	TG-5MS AMINE	Base Optimised 5% Phenyl Methylpolysiloxane	Non-polar	300°C / 315°C
	TG-5SilMS	Similar to 5% Phenyl Methylpolysiloxane	Non-polar	330°C / 350°C
	TG-5HT	5% Phenyl Methylpolysiloxane	Non-polar	380°C / 400°C
	TG-35MS	35% Phenyl Methylpolysiloxane	Mid-polarity	340°C / 360°C
	TG-35MS AMINE	Base Optimised 35% Phenyl Methylpolysiloxane	Mid-polarity	220°C
	TG-1301MS	6% Cyanopropylphenyl Methylpolysiloxane	Mid-polarity	260°C / 280°C
	TG-624	6% Cyanopropylphenyl Methylpolysiloxane	Mid-polarity	240°C
	TG-624SilMS	Similar to 6% Cyanopropylphenyl Methylpolysiloxane	Mid-polarity	320°C
	TG-VRX	Proprietary		260°C
	TG-VMS	Proprietary		260°C
	TG-1701MS	14% Cyanopropylphenyl Methylpolysiloxane	Mid-polarity	260°C / 280°C
	TG-17MS	50% Phenyl Methylpolysiloxane	Mid-polarity	300°C / 320°C
	TG-17SilMS	Similar to 50% Phenyl Methylpolysiloxane	Mid-polarity	340°C / 360°C
	TG-225MS	50% Cyanopropylmethyl Phenylmethylpolysiloxane	Mid-polarity	220°C / 240°C
	TG-200MS	Trifluoropropyl Methylpolysiloxane	Mid-polarity	320°C / 340°C
	TG-WaxMS	Polyethylene Glycol (PEG)	Polar	240°C / 260°C
	TG-WaxMS A	Acid Optimised Polyethylene Glycol (PEG)	Polar	240°C / 250°C
	TG-WaxMS B	Base Optimised Polyethylene Glycol (PEG)	Polar	200°C / 220°C
	TG-OCP I	Proprietary		340°C
	TG-OCP II	Proprietary		340°C
	TG-OPP I	Proprietary		330°C
	TG-OPP II	Proprietary		330°C
	TG-ALC Plus I	Proprietary		260°C
	TG-ALC Plus II	Proprietary		260°C
	TG-Dioxin	Proprietary		340°C
	TG-PAH	Proprietary		350°C / 360°C
	TG-PBDE	Proprietary		330°C / 360°C
	TG-VVOC	Proprietary		270°C / 290°C
	TG-POLAR	95% Cyanopropyl Phenylpolysiloxane	Polar	275°C
	TG-1MT	100% Methylpolysiloxane	Non-polar	430°C
	TG-5MT	5% Phenyl Methylpolysiloxane	Non-polar	430°C
	TG-WaxMT	Polyethylene Glycol (PEG)	Polar	240°C / 260°C

* Maximum temperature may vary with different column film thickness. Refer to box label for more details.

TraceGOLD
• Offering a leap forward in column performance delivering low bleed and superior inertness

TRACE
• For excellent quality and reproducibility for a wide range of applications

Range	Column	Phase	Polarity	Max operating temp °C (isothermal / programmable)
TRACE	TR-1MS	100% Dimethyl Polysiloxane	Non-polar	340°C / 360°C
	TR-5	5% Phenyl Methylpolysiloxane	Non-polar	320°C / 340°C for films ≤ 1.5µm 280°C / 300°C for films > 1.5µm
	TR-5MS	5% Phenyl Polysilphenylene-siloxane	Non-polar	360°C / 370°C for films ≤ 1.5µm 350°C / 360°C for films > 1.5µm
	TR-5HT	5% Phenyl Polycarborane Siloxane	Non-polar	380°C / 400°C
	TR-35MS	35% Phenyl Polysilphenylene-siloxane	Mid-polarity	330°C / 360°C
	TR-1701	14% Cyanopropylphenyl Polysiloxane	Mid-polarity	280°C / 300°C
	TR-50MS	50% Phenyl Polysilphenylene-siloxane	Mid-polarity	360°C / 370°C
	TR-225	50% Cyanopropylphenyl Polysiloxane	Mid-polarity	230°C / 250°C
	TR-Wax	Polyethylene Glycol (PEG)	Polar	260°C / 280°C for films ≤ 1.0µm 240°C / 260°C for films > 1.0µm
	TR-WaxMS	Polyethylene Glycol (PEG)	Polar	260°C / 280°C
	TR-FFAP	TPA Modified Polyethylene Glycol (PEG)	Polar	240°C / 250°C
	TR-SimDist	100% Dimethyl Polysiloxane	Non-polar	400°C for films ≤ 1.0µm 370°C for 2.65µm films
	TR-V1	6% Cyanopropylphenyl Polysiloxane	Mid-polarity	280°C / 300°C
	TR-FAME	70% Cyanopropyl Polysilphenylene-siloxane	Polar	250°C / 260°C
	TR-524	Cyanopropylphenyl Dimethyl Polysiloxane	Mid-polarity	240°C / 260°C
	TR-525	Proprietary	Mid-polarity	340°C / 360°C
	TR-527	5% Phenyl Polysilphenylene-siloxane	Non-polar	330°C / 350°C
	TR-8095	8% Phenyl Polycarborane-siloxane	Mid-polarity	360°C / 370°C
	TR-8270	5% Phenyl Polysilphenylene-siloxane	Non-polar	330°C / 350°C
	TR-PCB 8MS	8% Phenyl Polysilphenylene-siloxane	Mid-polarity	330°C / 350°C
	TR-Dioxin 5MS	5% Phenyl Polysilphenylene-siloxane	Non-polar	330°C / 350°C
	TR-Biodiesel (M)	100% Dimethyl Polysiloxane	Non-polar	300°C / 320°C
	TR-Biodiesel (F)	Polyethylene Glycol (PEG)	Polar	280°C / 300°C
	TR-Biodiesel (G)	5% Phenyl Polysilphenylene-siloxane	Non-polar	380°C / 400°C
	TR-DoA5	5% Phenyl Methylpolysiloxane	Non-polar	330°C / 350°C
	TR-DoA35	35% Phenyl Polysilphenylene-siloxane	Mid-polarity	330°C / 350°C
	TR-Pesticide	5% Phenyl Methylpolysiloxane	Non-polar	330°C / 350°C
	TR-Pesticide II	Proprietary	Non-polar	330°C / 350°C
	TR-Pesticide III	35% Phenyl Methylpolysiloxane	Mid-polarity	300°C / 320°C
	TR-Pesticide IV	35% Phenyl Methylpolysiloxane	Mid-polarity	300°C / 320°C
TracePLOT	TG-Bond Alumina (Na_2SO_4)	Na_2SO_4 Deactivated Aluminium Oxide	Non-polar	200°C
	TG-Bond Alumina (KCl)	KCl Deactivated Aluminium Oxide	Non-polar	200°C
	TG-Bond Msieve 5A	Molecular Sieve (5A)	Non-polar	300°C
	TG-Bond Q	100% Divinylbenzene	Non-polar	280°C / 300°C
	TG-Bond Q+	Porous Divinylbenzene Polymer	Mid-polarity	250°C
	TG-Bond S	Divinylbenzene 4-Vinylpyridine	Mid-polarity	250°C
	TG-Bond U	Divinylbenzene Ethylene Glycol / Dimethylacrylate	Polar	190°C

*** Maximum temperature may vary with different column film thickness. Refer to box label for more details.**

Learn more at **thermofisher.com/tracegold**

GC column selection by manufacturer

Column	Phase	Manufacturer	Recommended Thermo Scientific alternatives
Capillary	007-1(MS)	Quadrex	TG-1MS
	007-17(MPS-50)	Quadrex	TG-17MS
	007-1701	Quadrex	TG-1701MS
	007-2(MP-5)	Quadrex	TG-5MS
	007-2(MPS-5)	Quadrex	TG-5SilMS
	007-23	Quadrex	TR-FAME
	007-5MS	Quadrex	TG-5MS
	007-624	Quadrex	TG-624
	007-CW	Quadrex	TG-WaxMS
	AT-5	Alltech	TR-5
	AT50	Alltech	TG-17MS
	AT-5MS	Alltech	TG-5MS
	AT-624	Alltech	TG-624
	AT-Silar	Alltech	TR-FAME
	AT-Wax	Alltech	TR-WaxMS
	BP10	SGE	TG-1701MS
	BP20	SGE	TG-WaxMS
	BP21	SGE	TG-WaxMS A TR-FFAP
	BP225	SGE	TG-225MS
	BP5	SGE	TG-5MS
	BP624	SGE	TG-624 TG-624SilMS
	BPX1	SGE	TG-1MS TR-SimDist
	BPX5	SGE	TG-5MS
	BPX50	SGE	TG-17MS TG-17SilMS
	BPX608	SGE	TG-35MS
	BPX70	SGE	TR-FAME
	BPX90	SGE	TG-POLAR
	BPX-Volatiles	SGE	TG-624
	CARBOWAX	Agilent	TR-WaxMS
	CP-1301	Agilent	TG-1301MS
	CP-FFAP CB	Agilent	TG-WaxMS A TR-FFAP
	CP-Select624CB	Agilent	TG-624
	CP-Sil 19CB	Agilent	TG-1701MS
	CP-Sil 5CB MS	Agilent	TG-1MS
	CP-Sil 88	Agilent	TR-FAME
	CP-Sil 8CB	Agilent	TR-5MS/TG-5MS
	CP-SimDist	Agilent	TR-SimDist
	CP-Wax 51 (Amines)	Agilent	TG-WaxMS B
	CP-Wax 52CB	Agilent	TG-WaxMS TG-WaxMT
	CP-Wax 58 CB (FFAP)	Agilent	TG-WaxMS A TR-FFAP

Column	Phase	Manufacturer	Recommended Thermo Scientific alternatives
Capillary	DB-1	Agilent	TG-1MS TR-1MS
	DB-1301	Agilent	TG-1301MS
	DB-17	Agilent	TG-17MS
	DB-1701	Agilent	TG-1701MS
	DB-17ht	Agilent	TG-17MS
	DB-17ms	Agilent	TG-17MS TG-17SilMS
	DB-1ms	Agilent	TG-1MS TR-1MS
	DB-200	Agilent	TG-200MS
	DB-225	Agilent	TG-225MS
	DB-225ms	Agilent	TG-225MS
	DB-23	Agilent	TG-225MS
	DB-2887	Agilent	TR-SimDist
	DB-35	Agilent	TG-35MS
	DB-35ms	Agilent	TG-35MS
	DB-5	Agilent	TR-5 TG-5MS
	DB-5.625	Agilent	TG-5MS
	DB-5ht	Agilent	TG-5HT TG-5MT
	DB-5ms	Agilent	TG-5MS TG-5SilMS
	DB-624	Agilent	TG-624 TG-624SilMS
	DB-ALC1	Agilent	TG-ALC1 TG-ALC Plus I
	DB-ALC2	Agilent	TG-ALC2 TG-ALC Plus II
	DB-FFAP	Agilent	TG-WaxMS A TR-FFAP
	DB-HT Sim Dis	Agilent	TR-SimDist
	DB-PETRO	Agilent	TG-1MS
	DB-WAX	Agilent	TG-WaxMS TG-WaxMT
	DB-WAXetr	Agilent	TR-WaxMS TG-WaxMS
	DB-XLB	Agilent	TG-XLBMS
	Elite-1301	PerkinElmer	TG-1301MS
	Elite-17	PerkinElmer	TG-17MS
	Elite-1701	PerkinElmer	TG-1701MS
	Elite-17ms	PerkinElmer	TG-17MS
	Elite-200	PerkinElmer	TG-200MS
	Elite-23	PerkinElmer	TR-FAME
	Elite-35ms	PerkinElmer	TG-35MS
	Elite-5	PerkinElmer	TR-5
	Elite-5ms	Perkin Elmer	TG-5MS
	Elite-5ht	PerkinElmer	TG-5HT
	Elite-624	PerkinElmer	TG-624
	Elite-FFAP	PerkinElmer	TG-WaxMS A TR-FFAP
	Elite-WAX	PerkinElmer	TG-WaxMS
	Elite-WAX ETR	PerkinElmer	TG-WaxMS

GC column selection by manufacturer (continued)

Column	Phase	Manufacturer	Recommended Thermo Scientific alternatives
Capillary	HP-1	Agilent	TG-1MS TR-1MS
	HP-17	Agilent	TG-17MS TG-17SilMS
	HP-1701	Agilent	TG-1701MS
	HP-1MS	Agilent	TG-1MS TG-1MT
	HP20M	Agilent	TG-WaxMS
	HP-35	Agilent	TG-35MS
	HP-35MS	Agilent	TG-35MS
	HP-5	Agilent	TR-5
	HP-50+	Agilent	TG-17MS
	HP-5MS	Agilent	TG-5MS TG-5SilMS
	HP5-TA	Agilent	TG-5MS
	HP-88	Agilent	TR-FAME
	HP-FFAP	Agilent	TG-WaxMS A TR-FFAP
	HP-INNOWax	Agilent	TG-WaxMS TR-WaxMS
	HP-VOC	Agilent	TG-624 TG-624SilMS
	HP-Wax	Agilent	TG-WaxMS TR-WaxMS
	HT5	SGE	TG-5HT
	HT8	SGE	TR-PCB 8MS
	MDN-1	Sigma Aldrich	TG-1MS
	MDN-35	Sigma Aldrich	TG-35MS
	MDN-5	Sigma Aldrich	TR-5 TG-5MS
	MDN-5S	Sigma Aldrich	TG-5SilMS
	Nukol	Sigma Aldrich	TG-WaxMS
	OV-17	Ohio Valley	TG-17MS
	OV-1701	Ohio Valley	TG-1701MS
	OV-5	Ohio Valley	TR-5
	OV-624	Ohio Valley	TG-624
	Petrocol 2887	Sigma Aldrich	TR-SimDist
	Petrocol DH	Sigma Aldrich	TG-1MS
	Petrocol EX2887	Sigma Aldrich	TR-SimDist
	MXT-1	Restek	TG-1MT
	MXT-5	Restek	TG-5MT
	MXT-WAX	Restek	TG-WaxMT
	Rtx-1301	Restek	TG-1301MS
	Rtx-1701	Restek	TG-1701MS
	Rtx-1MS	Restek	TG-1MS
	Rtx-200	Restek	TG-200MS
	Rtx-200MS	Restek	TG-200MS
	Rtx-225	Restek	TG-225MS
	Rtx-2330	Restek	TG-POLAR
	Rtx-2560	Restek	TR-FAME
	Rtx-2887	Restek	TR-SimDist
	Rtx-35	Restek	TG-35MS
	Rtx-35 Amine	Restek	TG-35MS AMINE

Column	Phase	Manufacturer	Recommended Thermo Scientific alternatives
Capillary	Rtx-35MS	Restek	TG-35MS
	Rtx-5	Restek	TG-5MS TR-5
	Rtx-5 Amine	Restek	TG-5MS AMINE
	Rtx-50	Restek	TG-17MS
	Rtx-5SilMS	Restek	TG-5SilMS
	Rtx-624	Restek	TG-624
	Rtx-CLPesticides	Restek	TG-OCP I
	Rtx-CLPesticides2	Restek	TG-OCP II
	Rtx-OPPesticides	Restek	TG-OPP I
	Rtx-OPPesticides2	Restek	TG-OPP II
	Rtx-Dioxin 2	Restek	TG-Dioxin
	Rtx-VMS	Restek	TG-VMS
	Rtx-Volatiles	Restek	TG-624
	Rtx-VRX	Restek	TG-VRX
	Rtx-Wax	Restek	TG-WaxMS
	Rxi-17	Restek	TG-17MS
	Rxi-17SilMS	Restek	TG-17SilMS
	Rxi-1ms	Restek	TG-1MS
	Rxi-5HT	Restek	TG-5HT
	Rxi-5MS	Restek	TG-5MS
	Rxi-5SilMS	Restek	TG-5SilMS
	Rxi-624SilMS	Restek	TG-624SilMS
	Rxi-XLB	Restek	TG-XLBMS
	SE-30	Agilent	TG-1MS
	SE-52	Agilent	TG-5MS
	SE-54	Agilent	TG-5MS
	SolGel-Wax	SGE	TG-WaxMS
	SP-2100	Supelco	TG-1MS
	SP-2250	Supelco	TG-17MS
	SP-2330	Supelco	TR-FAME
	SP-2380	Supelco	TR-FAME
	SPB-1	Supelco	TG-1MS
	SPB-17	Supelco	TG-17MS
	SPB-35	Supelco	TG-35MS
	SPB-5	Supelco	TR-5 TG-5MS
	SPB-50	Supelco	TG-17MS
	SUPELCOWAX-10	Supelco	TG-WaxMS TR-WaxMS
	Stabilwax	Restek	TG-WaxMS
	Stabilwax-DA	Restek	TG-WaxMS A TR-FFAP
	Stabilwax-DB	Restek	TG-WaxMS B
	SUPELCOWAX-10	Supelco	TG-WaxMS
	VF-17ms	Agilent	TG-17MS TG-17SilMS
	VF-1ms	Agilent	TG-1MS TR-1MS
	VF-200ms	Agilent	TG-200MS
	VF-23ms	Agilent	TG-POLAR
	VF-35ms	Agilent	TG-35MS
	VF-5ht	Agilent	TG-5HT
	VF-5ms	Agilent	TG-5MS
	VF-Xms	Agilent	TG-XLBMS

GC column selection by manufacturer (continued)

Column	Phase	Manufacturer	Recommended Thermo Scientific alternatives
Capillary	ZB-1/ZB-1plus/ZB-1MS	Phenomenex	TR-1MS/TG-1MS
	ZB-1HT		TG-1HT
	ZB-5/ZB-5plus/ZB-5MS/ ZB-5MSplus	Phenomenex	TR-5/TR-5MS/TG-5MS/TG-5MS Amine
	ZB-5MSi		TG-5SilMS
	ZB-5HT		TR-5HT/TG-5HT
	ZB-XLB/ZB-XLB-HT	Phenomenex	TG-XLBMS
	ZB-35/ZB-35HT	Phenomenex	TR-35MS
	TR-35MS/TG-35MS		TG-35MS(TG-35SilMS)
	ZB-50	Phenomenex	TR-50MS/TG-17MS/TG-17SilMS
	ZB-624	Phenomenex	TG-624/TG-624SilMS
	ZB-1701/ZB-1701P	Phenomenex	TR-1701/TG-1701MS
	ZB-WAX/ZB-WAXplus	Phenomenex	TR-Wax/TR-WaxMS/TG-WaxMS
	ZB-FFAP	Phenomenex	TR-FFAP/TG-WaxMS A
PLOT	Alumina-PLOT	Supelco	TG-BOND Alumina (Na_2SO_4)
	AT-Alumina	Alltech	TG-BOND Alumina (Na_2SO_4)
	AT-Molsieve	Alltech	TG-BOND Msieve 5A
	AT-Q	Alltech	TG-BOND Q
	CP-Al2O3/KCl	Agilent	TG-BOND Alumina (KCl)
	CP-Al2O3/Na_2SO_4	Agilent	TG-BOND Alumina (Na_2SO_4)
	CP-Molsieve 5A	Agilent	TG-BOND Msieve 5A
	CP-PoraPLOT Q	Agilent	TG-BOND Q
	CP-PoraPLOT S	Agilent	TG-BOND S
	CP-PoraPLOT U	Agilent	TR-BOND U
	GS-Alumina	Agilent	TG-BOND Alumina (Na_2SO_4)
	GS-Alumina KCl	Agilent	TG-BOND Alumina (KCl)
	GS-Molsieve	Agilent	TG-BOND Msieve 5A
	GS-Q	Agilent	TG-BOND Q+
	HP PLOT M	Agilent	TG-BOND Alumina (Na_2SO_4)
	HP PLOT Molsieve	Agilent	TG-BOND Msieve 5A
	HP PLOT S	Agilent	TG-BOND Alumina (Na_2SO_4)
	HP-UPLOT	Agilent	TG-BOND U
	PoraBond Q	Agilent	TG-BOND Q
	PoraBond U	Agilent	TG-BOND U
	Molsieve 5A PLOT	Supelco	TG-BOND Msieve 5A
	PLT-5A	Quadrex	TG-BOND Msieve 5A
	Rt-Alumina Bond (KCl)	Restek	TG-BOND Alumina (KCl)
	Rt-Alumina Bond (Na_2SO_4)	Restek	TG-BOND Alumina (Na_2SO_4)
	Rt-Msieve 5A	Restek	TG-BOND Msieve 5A
	Rt-Q-BOND	Restek	TG-BOND Q
	Rt-QS-BOND	Restek	TG-BOND Q+
	Rt-S-BOND	Restek	TG-BOND S
	Rt-U-BOND	Restek	TG-BOND U
	Supel-Q-PLOT	Supelco	TG-BOND Q

GC column selection by application

- Recommended
- Alternative

	TG-1MS, TR-1MS	TG-5MS, TG-5SilMS, TG-5MS AMINE, TG-5MT, TR-5, TR-5MS	TG-35MS, TG-35MS AMINE, TR-35MS	TG-17MS, TG-17SilMS	TG-1301MS	TG-1701MS, TR-1701	TG-WaxMS, TG-WaxMT, TR-Wax, TR-WaxMS	TG-WaxMS A	TG-WaxMS B	TG-PAH	TG-POLAR	TG-624, TG-624SilMS	TG-200MS	TG-225MS	TG-5HT, TR-5HT	TG-XLBMS	TG-VRX, TG-VMS	TG-OCP I, TG-OCP II	TG-OPP I, TG-OPP II	TG-ALC Plus I, TG-ALC Plus II	TR-FFAP	TR-V1	TR-FAME	TR-Simdist	TR-524	TR-525	TG-PBDE	TR-8270	TR-DoA5Q	TR-DoA35	TR-Biodiesel (M)	TR-Biodiesel (F)	TR-Biodiesel (G)	TG-Dioxin TR-Dioxin 5MS/TG-Dioxinß	TR-Pesticide, TR-Pesticide II, TR-Pesticide III, TR-Pesticide IV	TR-PCB 8MS	TR-8095
Acids		•					•	•													•																
Acid/neutral drugs		•	•																																		
Alcohols		•			•		•	•	•		•	•	•								•	•															
Alcohols in beverages					•		•	•	•			•									•	•															
Aldehydes							•	•	•		•										•																
Alditol acetates (sugars)		•					•	•	•					•							•		•														
Amines – aliphatic		•	•		•		•		•			•										•														
Amines – aromatic		•	•		•		•		•			•										•														
Antidepressants		•	•																																	
Benzenes, substituted															•																					
Biodiesel – methanol																															•					
Biodiesel – fames											•																					•				
Biodiesel – glycerine																																	•			
Blood alcohols											•									•																
Brominated flame retardants		•																								•										
Butter fat		•													•																					
Carboxylic acids								•																													
Cigarette lighter fuel		•			•							•										•														
Chlorinated aromatics	•	•	•			•							•																							•	
Dioxins		•		•																															•			
Drugs of abuse			•																										•								
Drugs of Abuse – THC																														•							
Essential oils							•	•	•												•																
Explosives																																					•	
Fames											•			•	•								•														
Glucose – methylated																								•													
Herbicides		•	•	•		•	•	•	•		•		•					•			•														•		
Ketones				•	•	•	•	•	•		•	•	•	•							•	•	•														
Monomers					•							•										•															
Nitroaromatics		•	•	•		•	•	•	•		•										•																
Organic acids								•																													
Organochlorine pesticides	•	•	•	•		•							•					•																	•		
Organophosphorous pesticides	•	•	•	•															•																	•		
Pahs	•	•	•	•		•		•							•																						
Paraffins	•	•																																			
Pcbs		•														•																				•	
Pesticides		•														•																				•		
Petroleum															•									•													
Phenols		•	•				•	•	•		•		•																								
Phthalates		•	•													•					•																
Plant sterols		•	•	•																																•		
Polyethylene															•																						
Polymers	•																																				
Polywax	•	•													•																						
Pyrethroids	•	•	•	•		•																														•		
Sedatives		•	•																																		
Semivolatiles	•	•																								•		•								
Silicon oil															•																						
Solvents					•		•	•	•		•	•									•	•															
Terpenes		•																																		•		
Triglycerides		•													•																						
Trph	•	•																																			
Volatiles		•			•		•	•	•		•	•					•				•	•			•												
Xylenes	•	•					•	•	•												•			•													

GC column selection by U.S. pharmacopeia specifications

The USP specifications are listed below with the appropriate Thermo Scientific GC column offerings included for your convenience. In some cases, there is more than one phase that matches the phase description. When in doubt, it is recommended that you consult the original complete method or contact our technical support team for additional information or help in choosing the correct column for your application.

USP Code	Description	Recommended Thermo Scientific phases
G1	Dimethylpolysiloxane oil	TG-1MS
		TG-1MT
		TR-1MS
G2	Dimethylpolysiloxane gum	TG-1MS
		TG-1MT
		TR-1MS
G3	50% Phenyl-50% Methylpolysiloxane	TG-17MS
		TR-50MS
		TG-17SilMS
G5	3-Cyanopropylpolysiloxane	TR-FAME
G6	Trifluoropropyl Methylpolysiloxane	TG-200MS
G7	50% Cyanopropyl Phenylmethyl Polysiloxane	TG-225MS
G16	Polyethylene Glycol Compound (ave. mol. wt. ~15,000) with Diepoxide Linker	TG-WaxMS
		TG-WaxMT
		TR-WaxMS
		TR-Wax
G19	50% Cyanopropyl 50% Phenylmethyl Polysiloxane	TG-225MS
G20	Polyethylene Glycol (ave. mol. wt. of 380 – 420)	TG-WaxMS
		TG-WaxMT
		TR-WaxMS
		TR-Wax
G27	5% Phenyl-95% Methylpolysiloxane	TG-5MS
		TG-5MT
		TR-5MS
		TR-5
G36	1% Vinyl-5% Phenylmethylpolysiloxane	TR-5MS
		TR-5
G38	Phase G1 containing a small percentage of tailing inhibitor	TG-5MS
		TG-5MT
		TR-5MS
		TR-5
G42	35% Phenyl-65% Dimethylpolysiloxane (percentages refer to molar substitution)	TG-35MS
		TR-35MS
G43	6% Cyanopropylphenyl-94% Dimethylpolysiloxane (percentages refer to molar substitution)	TG-624
		TR-V1
		TG-624SilMS
G46	14% Cyanopropylphenyl-86% Methylpolysiloxane	TG-1701MS
		TR-1701
G48	90% Biscyanopropyl 10% Cyanopropyl Phenyl Polysiloxane	TG-POLAR

GC column selection by ASTM method

Selected ASTM methods are listed below with the appropriate Thermo Scientific GC column offerings. In some cases, there is more than one phase or column dimension that can be used. When in doubt, it is recommended that you consult the original complete method or contact our technical support team for additional information or help in choosing the correct column for your application.

Method	Title	Recommended Thermo Scientific phases	Cat. no.
D1983	Fatty acid methyl ester composition	TG-WaxMS	**26088-1420**
D2245	Oils and oil acids in solvent-reducible paints	TR-FAME	**260M154P**
D2268	High-purity n-heptane and isooctane	TG-1MS	**Inquire**
D2306	C8 aromatic hydrocarbons	TG-WaxMS	**26088-1540**
D2360	Trace impurities in monocyclic aromatic hydrocarbons	TG-WaxMS	**26088-1550**
D2456	Polyhydric alcohols in alkyd resin	TG-WaxMS	**26088-2980**
D2580	Phenols in water	TG-5MS	**26098-2230**
D2753	Oil and oil acids	TR-FAME	**260M154P**
D2800	FAME analysis	TR-FAME	**260M154P**
D2804	Purity of methyl ethyl ketone	TG-WaxMS	**26088-2980**
D2887	Boiling range distribution of petroleum fractions	TR-SimDist	**260S348P**
D2998	Polyhydric alcohols in alkyd resin	TG-1MS	**26099-2970**
D2999	Monopentaerythritol in commercial pentaerythritol	TG-1MS	**Inquire**
D3009	Composition of turpentine	TG-WaxMS	**26088-2240**
D3054	Cyclohexane	TG-1MS	**Inquire**
D3168	Polymers in emulsion paints	TG-1MS	**26099-2970**
D3257	Aromatics in mineral spirits	TG-624	**26085-3960**
D3271	Solvent analysis in paints	TG-WaxMS	**26088-2980**
D3304	PCBs in environmental materials	TG-5MS TR-PCB 8MS	**26098-1540** **26AJ148P**
D3329	Purity of methyl isobutyl ketone	TG-WaxMS TG-624	**26088-2980** **26085-3960**
D3432	Unreacted toluene diisocyanates in urethane prepolymers and coating solutions	TG-1MS	**26099-3090**
D3447	Purity of halogenated organic solvents	TG-624	**26085-3960**
D3452	Identification of rubber	TG-1MS	**26099-3090**
D3457	FAME analysis	TR-FAME	**260M154P**
D3534	PCBs in water	TG-5MS TR-PCB 8MS	**26098-3360** **26AJ148P**
D3545	Alcohol content and purity of acetate esters	TG-624	**26085-3960**
D3687	Alcohol content and purity of acetate esters	TG-WaxMS	**26088-2980**
D3695	Volatile alcohols in water by direct aqueous-injection GC	TG-WaxMS	**26088-2980**
D3710	Boiling range distribution of gasoline and gasoline fractions	TR-SimDist	**260S348P**
D3725	Fatty acids in drying oils	TR-FAME	**Inquire**
D3760	Isopropylbenzene (cumene)	TG-WaxMS TG-1MS	**26088-1550** **Inquire**
D3797	o-Xylene	TG-WaxMS	**26088-2360**
D3798	p-Xylene	TG-WaxMS	**26088-2360**
D3871	Purgeable organic compounds in water using headspace sampling	TG-624	**26085-4080**
D3893	Purity of methyl amyl ketone and methyl isoamyl ketone	TG-624	**26085-3960**
D3973	Low molecular weight halogenated hydrocarbons in water	TG-624	**26085-3960**
D4059	PCBs in insulating liquids	TG-5MS TR-PCB 8MS	**26098-1540** **26AJ148P**
D4415	Dimer in acrylic acid	TG-WaxMS	**26088-1430**

GC column selection by ASTM method (continued)

Method	Title	Recommended Thermo Scientific phases	Cat. no.
D4443	Residual vinyl chloride monomer content in ppb range in homo- and co-polymers by headspace GC	TG-624	**26085-3960**
D4735	Trace thiophene in refined benzene	TG-WaxMS	**26088-2250**
D4773	Propylene gycol monomethyl ether, dipropylene glycol monomethyl ether, and propylene glycol monomethyl ether acetate	TR-5	**260E470P**
D4806	Denatured fuel ethanol for blending with gasoline for use as automotive spark-ignition engine fuel	TG-1MS	**Inquire**
D4864	Traces of methanol in propylene concentrates	TG-5MS	**Inquire**
D4947	Chlordane and heptachlor in indoor air	TG-5MS	**26098-3360**
D5060	Impurities in high-purity ethylbenzene	TG-WaxMS	**26088-2360**
D5075	Nicotine in indoor air	TG-5MS	**26098-2970**
D5134	Petroleum naphthas through n-nonane	TG-1MS	**Inquire**
D5135	Styrene	TG-WaxMS	**26088-2360**
D5399	Boiling point distribution of hydrocarbon solvents	TR-SimDist	**260S348P**
D5441	Methyl t-butyl ether	TG-1MS	**Inquire**
D5442	Petroleum waxes	TG-1MS TG-5MS	**26099-1430** **26098-1300**
D5480	Motor oil volatility	TG-5MS	**Inquire**
D5501	Ethanol content of denatured fuel ethanol	TG-1MS	**Inquire**
D5599	Oxygenates in gasoline by oxygen selective FID	TG-1MS	**26099-3080**
D5623	Sulfur compounds in light petroleum liquids using sulfur selective detection	TG-1MS	**Inquire**
D5713	High purity benzene for cyclohexane feedstock	TG-1MS	**Inquire**
D5739	Oil spill source identification using positive ion electron impact low resolution MS	TG-5MS	**26098-1420**
D5769	Benzene, toluene and total aromatics in finished gasolines	TG-1MS TG-624 TG-624SilMS	**26099-3080** **26085-3330** **26059-3330**
D5790	Purgeable organic compounds in water	TG-5MS	**26098-1420**
D5812	Organochlorine pesticides in water	TG-1701MS TG-17MS TG-WaxMS	**26090-1420** **26089-1420** **26088-1550**
D5917	Trace impurities in monocyclic aromatic hydrocarbons	TR-FAME	**260M154P**
D5974	Fatty and rosin acids in tall oil fraction products	TG-1MS	**Inquire**
D5986	Oxygenates, benzene, toluene, C8-C12 aromatics and total aromatics in finished gasoline by GC/FTIR	TG-5MS	**26098-1420**
D6160	PCBs in waste materials	TR-SimDist	**260S250P**
D6352	Boiling range distribution of petroleum fractions	TG-1MS TR-SimDist	**Inquire** **260S250P**
D6417	Engine oil volatility	TG-1MS	**Inquire**
D6584	Free and Total Glycerin in B-100 Biodiesel	TR-BioDiesel (G)	**26AF024P**
D6729	Individual components in spark ignition engine fuels	TG-1MS	**Inquire**
D6730	Individual components in spark ignition engine fuels using precolumn	TG-5MS TG-624	**26098-2960** **26085-4080**
E202	Ethylene glycols and propylene glycols	TR-5	**260E470P**
E475	Di-tert-butyl peroxide	TG-1MS	**Inquire**
E1616	Acetic anhydride	TG-WaxMS	**26088-3090**
E1863	Acrylonitrile	TR-SimDist	**260S250P**

GC column selection by U.S. EPA drinking water test method

Selected EPA drinking water methods are listed below with the appropriate Thermo Scientific GC column offerings. In some cases, there is more than one phase or column dimension that can be used. When in doubt, it is recommended that you consult the original complete method or contact our technical support team for additional information or help in choosing the correct column for your application.

Method	Title	Recommended Thermo Scientific phases	Cat. no.
501.3	Trihalomethanes	TG-624	**26085-3960**
502.1	Volatile halogenated compounds	TG-624 TR-5MS	**26085-4080** **260F396P**
502.2	Volatile organic compounds	TG-624 TG-624	**26085-4080** **26085-3320**
503.1	Volatile aromatic and unsaturated organics	TG-624 TR-5MS	**26085-4080** **260F396P**
504	EDB and DBCP	TR-5MS TG-5MS	**260F396P** **26098-2240**
504.1	EDB and DBCP	TR-5MS TG-5MS	**260F396P** **26098-2240**
506	Phthalates and adipates	TG-1MS TG-5MS	**26099-1430** **26098-1430**
507	Organonitrogen and organophosphorus pesticides	TG-5MS TG-5MT TG-17MS TG-17SilMS	**26098-1420** **26M98-1420** **26089-1420** **26072-1420**
509	Ethylene thiourea	TG-1701MS TG-WaxMS	**26090-1420** **26088-1300**
513	Dioxin	TG-5MS TG-5MT	**26098-1540** **26M98-1540**
515.2	Chlorinated herbicides	TG-5MS TG-17MS	**26098-1430** **26089-1430**
524.1	Volatile organic compounds	TR-524 TG-624 TG-624 TG-624SilMS TG-624SilMS	**26RV495P** **26085-4080** **26085-3320** **26059-4080** **26059-3320**
524.2	Volatile organic compounds	TR-524 TG-624 TG-624 TG-VMS	**26RV495P** **26085-4080** **26085-3320** **26080-4950**
525.1	Semi-volatile organic compounds	TR-525 TG-5MS TG-624SilMS TG-624SilMS	**26RX142P** **26098-1420** **26059-4080** **26059-3320**
525.2	Semi-volatile organic compounds	TR-525 TG-5MS	**26RX142P** **26098-1420**
527	Selected pesticides and flame retardants	TR-527 TG-5MS	**26RF142P** **26098-1420**
548.1	Endothall	TG-1MS TG-5MS	**26099-1430** **26098-1420**
551	Chlorinated disinfection by-products/chlorinated solvents	TG-5MS TG-1701MS	**26098-1420** **26090-2240**
552	Haloacetic acids	TG-1701MS TG-35MS	**26090-1430** **26094-1430**
552.1	Haloacetic acids and dalapon	TG-1701MS TG-35MS	**26090-1430** **26094-1430**

GC column selection by U.S. EPA waste water test method

Selected EPA waste water methods are listed below with the appropriate Thermo Scientific GC column offerings. In some cases, there is more than one phase or column dimension that can be used. When in doubt, it is recommended that you consult the original complete method or contact our technical support team for additional information or help in choosing the correct column for your application.

Method	Title	Recommended Thermo Scientific phases	Cat. no.
601	Purgeable halocarbons	TG-624 TG-624	**26085-4080** **26085-3320**
602	Purgeable aromatics	TG-624 TG-5MS TG-5MT	**26085-4080** **26098-2960** **26M98-2960**
603	Acrolein and acrylonitrile	TG-624 TG-5MS TG-5MT	**26085-4080** **26098-2960** **26M98-2960**
604	Phenols	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**
606	Phthalate ester	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**
607	Nitrosamines	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**
608.1	Organochlorine pesticides in industrial and municipal water	TG-5MS	**26098-2240**
608.2	Organochlorine pesticides in wastewater	TG-5MS	**26098-2240**
609	Nitroaromatics and isophorone	TG-5MS TG-35MS	**26098-1430** **26094-1430**
610	Polynuclear aromatic hydrocarbons	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
611	Haloethers	TG-5MS TG-35MS	**26098-1430** **26094-1430**
612	Chlorinated hydrocarbons	TG-5MS TG-35MS	**26098-1430** **26094-1430**
613	Dioxin	TG-5MS TG-5MT	**26098-1540** **26M98-1540**
614	Organophosphorous pesticides in industrial and municipal water	TG-5MS TG-5MT TG-17MS TG-17SilMS	**26098-1420** **26M98-1420** **26089-1420** **26072-1420**
614.1	Organophosphorous pesticides in wastewater	TG-5MS TG-5MT TG-17MS	**26098-1420** **26M98-1420** **26089-1420**
615	Chlorinated herbicides in industrial and municipal water	TG-5MS TG-5MT TG-17MS	**26098-1420** **26M98-1420** **26089-1420**
616	C, H, and O compounds	TG-1MS TG-5MS TG-5MT	**26099-1420** **26098-1420** **26M98-1420**
617	Organohalide pesticides and PCBs in industrial and municipal water	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
618	Volatile pesticides in industrial and municipal water	TG-1MS TG-5MS	**26099-2240** **26098-2240**
619	Triazines, pesticides and PCBs in industrial and municipal water	TG-35MS	**26094-1430**
620	Diphenylamine in industrial and municipal water	TG-1MS TG-5MS	**26099-1430** **26098-1430**
622	Organophosphorous pesticides in industrial and municipal water	TG-5MS TG-5MT TG-17MS	**26098-1420** **26M98-1420** **26089-1420**

GC column selection by U.S. EPA waste water test method (continued)

Method	Title	Recommended Thermo Scientific phases	Cat. no.
622.1	Thiophosphate pesticides	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**
624	Purgeables	TG-624 TG-624 TG-624SilMS TG-624SilMS TG-VMS	**26085-4080** **26085-3320** **26059-4080** **26059-3320** **26080-4950**
625	Base/neutrals and acids	TG-5MS TG-5MT TG-5MS	**26098-1420** **26M98-1420** **26098-1430**
627	Dinitroaniline pesticides in industrial and municipal water	TG-5MS TG-35MS	**26098-1430** **26094-1430**
630.1	Dithiocarbamate pesticides such as carbon disulfide	TG-5MS TG-5MS TG-5MT	**26098-1420** **26098-1430** **26M98-1420**
633	Organonitrogen pesticides	TG-5MS TG-5MT TG-17MS TG-17SilMS	**26098-1420** **26M98-1420** **26089-1420** **26072-1420**
633.1	Neutral nitrogen-containing pesticides	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**
634	Thiocarbamate pesticides	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**
645	Amine pesticides and lethane in industrial and municipal water	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**
646	Dinitro aromatic pesticides	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**

GC column selection by U.S. EPA solid waste test method

Selected EPA solid waste methods are listed below with the appropriate Thermo Scientific GC column offerings. In some cases, there is more than one phase or column dimension that can be used. When in doubt, it is recommended that you consult the original complete method or contact our technical support team for additional information or help in choosing the correct column for your application.

Method	Title	Recommended Thermo Scientific phases	Cat. no.
8010B	Halogenated volatile organics	TG-624 TG-624	**26085-4080** **26085-3320**
8011	EDB and DBCP	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
8015B	Non-halogenated volatile organics	TG-624 TG-5MS TG-5MT	**26085-4080** **26098-2960** **26M98-2960**
8020A	Aromatic volatile organics	TG-624 TG-5MS TG-5MT	**26085-4080** **26098-2960** **26M98-2960**
8021A	Halogenated and aromatic volatile organics	TG-624 TG-5MS TG-5MT	**26085-4080** **26098-2960** **26M98-2960**
8030A	Acrolein and acrylonitrile	TG-624	**26085-4080**
8031	Acrylonitrile	TG-624	**26085-3390**
8032	Acrylamide	TG-624	**26085-3390**

GC column selection by U.S. EPA solid waste test method (continued)

Method	Title	Recommended Thermo Scientific phases	Cat. no.
8040A	Phenols	TG-5MS TG-5MT TG-35MS	**26098-1420** **26M98-1420** **26094-1420**
8060	Phthalate esters	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
8061	Phthalate esters	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
8070	Nitrosamines	TG-5MS	**26098-1430**
8081	Organochlorine pesticides and PCBs	TG-5MS TG-5MT TG-17MS TG-17SilMS TG-OCP I/II	**26098-2230** **26M98-2q230** **26089-1420** **26072-1420** **26078-5760/26077-1430**
8090	Nitroaromatics and cyclic ketones	TG-5MS	**26098-1430**
8095	Explosives	TR-8095	**260P123P**
8100	Polynuclear aromatic hydrocarbons	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
8110	Haloethers	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
8120A	Chlorinated hydrocarbons	TG-5MS	**26098-1430**
8121	Chlorinated hydrocarbons	TG-5MS	**26098-1430**
8140	Organophosphorous pesticides	TG-5MS TG-5MT TG-17MS	**26098-1420** **26M98-1420** **26089-1420**
8141A	Organophosphorous pesticides	TG-OPP/TG OPP II	**26098-1420** **26M98-1420** **26089-1420**
8150B	Chlorinated herbicides	TG-5MS TG-5MT TG-17MS TG-OCP I/TG-OCP II	**26098-1420** **26M98-1420** **26089-1420** **26076-2240/26075-5760**
8151	Chlorinated herbicides	TG-5MS TG-5MT TG-17MS TG-OCP I/TG-OCP II	**26098-1420** **26M98-1420** **26089-1420**
8240B	Volatile organic compounds	TG-624 TG-624 TG-624SilMS TG-624SilMS	**26085-4080** **26085-3320** **26059-4080** **26059-3320**
8250A	Semi-volatile organic compounds	TG-5MS TG-5MS TG-5MT	**26098-1420** **26098-1430** **26M98-1420**
8260A	Volatile organic compounds	TG-624 TG-624 TG-VMS	**26085-4080** **26085-3320** **26080-4950**
8270B	Semi-volatile organic compounds	TG-5MS TG-5MS TG-5MT	**26098-1420** **26098-1430** **26M98-1420**
8270C	Semi-volatile organic compounds	TR-8270	**26RF296P**
8280	Polychlorinated dioxins and furans	TG-5MS TG-5MT	**26098-1540** **26M98-1540**
8290	Polychlorinated dioxins and furans	TG-5MS TG-5MT	**26098-1540** **26M98-1540**

GC column selection by NIOSH method

Selected NIOSH methods are listed below with the recommended Thermo Scientific GC column offerings included for your convenience. There may be more than one phase or column dimension that can be used. When in doubt, it is recommended that you consult the original complete method or contact our technical support team for additional information or help in choosing the correct column for your application.

Method	Title	Recommended Thermo Scientific phases	Cat. no.
801	Aerobic bacteria	TR-FAME	**Inquire**
1001	Methylene chloride	TG-1MS	**26099-1430**
1002	Chloroprene	TG-1MS TG-1MT	**26099-2960** **26M99-2960**
1003	Halogenated hydrocarbons	TG-624 TG-624SilMS	**26085-3390** **26059-3390**
1004	Dichloroethyl ether	TG-1MS	**Inquire**
1005	Methylene chloride	TG-WaxMS	**26088-1430**
1010	Epichlorohydrin	TG-WaxMS	**Inquire**
1011	Ethyl bromide	TG-WaxMS	**26088-2240**
1013	Propylene dichloride	TG-WaxMS	**Inquire**
1015	Vinylidene chloride	TG-624	**Inquire**
1016	1,1,2,2-Tetrachloro-2,2-difluoroethane and 1,1,2,2-tetrachloro-1,2-difluoroethane	TG-WaxMS	**26088-2240**
1018	Dichlorodifuoromethane, 1,2-dichlorotetrafluoroethane and chlorodifluoromethane	TG-1MS	**26099-2970**
1020	1,1,2-Trichloro-1,2,2-trifluoroethane	TG-WaxMS	**26088-1430**
1300	Ketones 1	TG-WaxMS	**26088-2240**
1301	Ketones 2	TG-WaxMS	**26088-2240**
1302	N-Methyl-2-pyrrolidone	TG-5MS	**26098-2970**
1400	Alcohols 1	TG-WaxMS	**26088-2240**
1401	Alcohols 2	TG-WaxMS	**26088-2240**
1402	Alcohols 3	TG-WaxMS	**26088-2240**
1403	Alcohols 4	TG-WaxMS	**26088-1430**
1450	Esters 1	TG-WaxMS	**26088-2240**
1451	Methyl cellosolve acetate	TG-5MS	**26098-2970**
1453	Vinyl acetate	TG-5MS	**26098-2970**
1454	Isopropyl acetate	TG-1MS	**26099-2970**
1457	Ethyl acetate	TG-WaxMS	**26088-2970**
1458	Methyl acetate	TG-WaxMS	**26088-2970**
1501	Aromatic hydrocarbons	TG-WaxMS	**26088-2970**
1550	Naphthas	TG-1MS TG-1MT	**26099-1540** **26M99-1540**
1551	Turpentine	TG-1MS TG-1MT	**26099-1540** **26M99-1540**
1552	Terpenes	TG-WaxMS	**26088-3100**
1601	1,1-Dichloro-1-nitroethane	TG-1MS	**Inquire**
1602	Dioxane	TG-5MS	**26098-2970**
1604	Acrylonitrile	TG-WaxMS	**26088-2240**
1606	Acetonitrile	TG-WaxMS	**26088-2970**
1608	Glycidol	TG-WaxMS	**Inquire**
1609	Tetrahydrofuran	TG-WaxMS	**26088-2240**
1610	Ethyl ether	TG-1MS	**26099-2970**
1611	Methylal	TG-WaxMS	**Inquire**
1612	Propylene oxide	TG-5MS	**26098-2970**
1613	Pyridine	TG-5SilMS	**26096-2970**
1614	Ethylene oxide	TG-WaxMS	**Inquire**
1615	Methyl-tert-butyl ether	TG-1MS	**26099-2240**
2000	Methanol	TG-35MS	**26094-2980**
2004	Dimethylacetylamide and dimethylformamide	TG-WaxMS	**26088-2240**

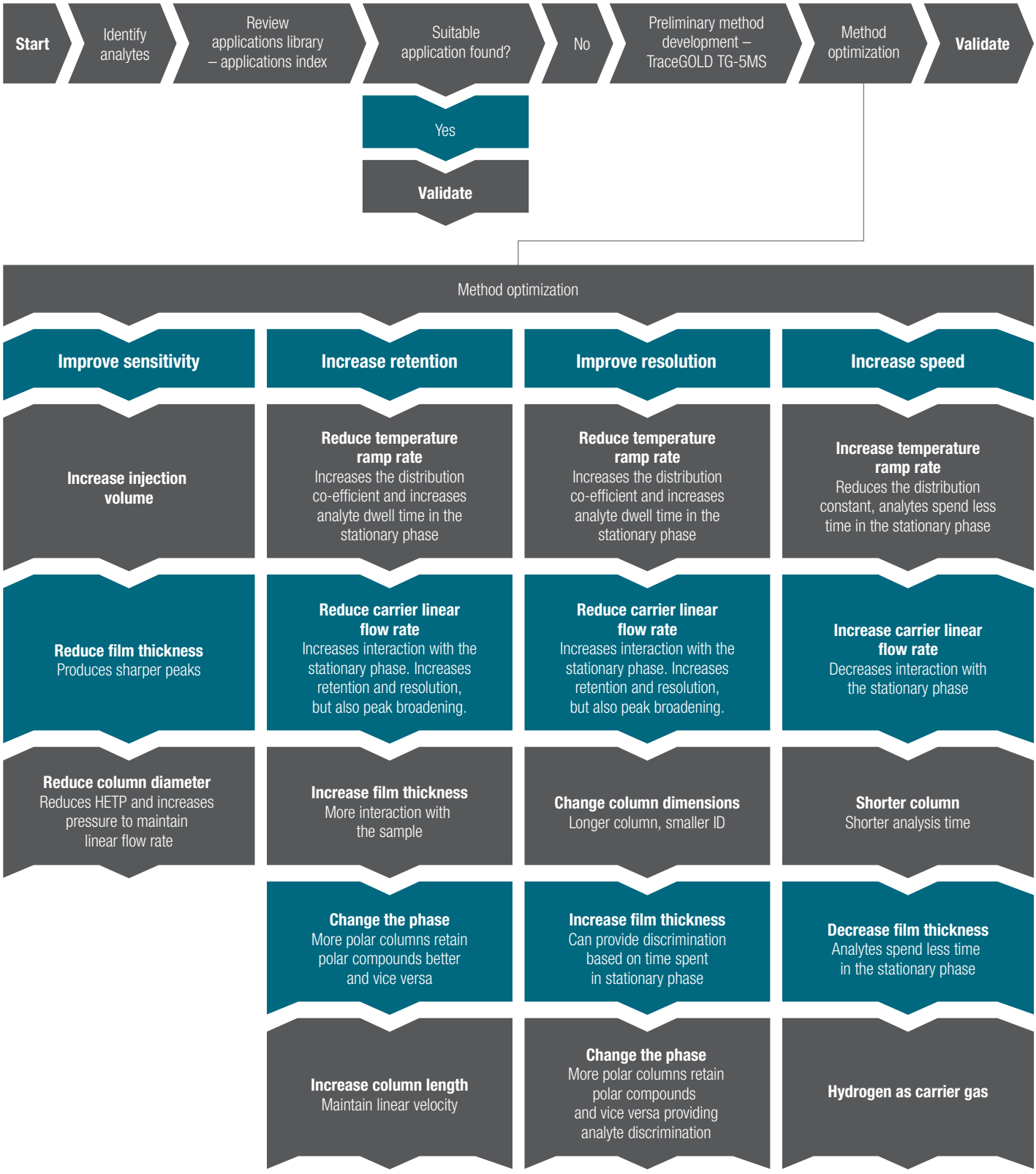
GC column selection by NIOSH method (continued)

Method	Title	Recommended Thermo Scientific phases	Cat. no.
2005	Nitroaromatics	TG-5MS TG-5MT	**26098-2250** **26M98-2250**
2007	Aminoethanol compounds 1	TG-5MS	**Inquire**
2010	Aliphatic amines	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
2012	n-Butylamine	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
2017	Aniline, o-toluidine and nitrobenzene	TG-5MS	**26098-2970**
2500	Methyl ethyl ketone	TG-1MS	**26099-2970**
2505	Furfuryl alcohol	TG-1MS TG-1MT	**26099-1420** **26M99-1420**
2520	Methyl bromide	TG-1MS	**26099-2970**
2529	Furfural	TG-5MS TG-5MT	**26098-2960** **26M98-2960**
2536	Valeraldehyde	TG-5MS	**26098-1310**
2537	Methyl methacrylate	TG-35MS	**26094-2980**
2541	Formaldehyde	TG-WaxMS	**26088-2240**
2542	Mercaptans	TG-1MS TG-1MT	**26099-2960** **26M99-2960**
2546	Cresols and phenol	TG-WaxMS	**26088-1430**
2549	Volatile organic compounds (screening)	TG-1MS TG-1MT	**26099-2960** **26M99-2960**
2550	Benzothiazole in asphalt fume	TG-1MS	**26099-2970**
2551	Nicotine	TG-5MS	**26098-2970**
3511	Monomethylaniline	TG-5MS TG-5MT	**26098-1420** **26M98-1420**
3513	Tetranitromethane	TG-1MS TG-1MT	**26099-1420** **26M99-1420**
5020	Dibutyl phthalate and di(2-ethylhexyl) phthalate	TG-1MS TG-1MT	**26099-1300** **26M99-1300**
5515	Polynuclear aromatic hydrocarbons	TG-1MS	**26099-3090**
5519	Endrin	TG-1MS	**26099-3090**
5523	Glycols	TG-35MS	**26094-2980**
5600	Organophosphorus pesticides	TG-5MS	**26098-2970**
5602	Chlorinated organonitrogen herbicides (air sampling)	TG-17MS TG-17SilMS	**26089-1420** **26072-1420**
5701	Resorcinol	TG-1MS TG-1MT	**26099-1420** **26M99-1420**
9200	Chlorinated organonitrogen herbicides (hand wash)	TG-17MS TG-17SilMS	**26089-1420** **26072-1420**
9201	Chlorinated organonitrogen herbicides (dermal patch)	TG-17MS TG-17SilMS	**26089-1420** **26072-1420**

GC technical information

GC method selection and optimization

The following flow chart briefly describes the common steps in GC method development and optimization.



GC troubleshooting

Before you start any troubleshooting, it is essential to observe safe laboratory practices. Know the chemical and physical properties of any solvents used and have the appropriate Material Safety Data Sheets (MSDSs) readily available. All electrically powered instruments should be shut down and unplugged before starting. Eye protection should also be worn.

The following table lists common GC problems encountered, the possible causes and solutions for your quick reference.

Symptom	Cause	Recommended solutions
Baseline related problems		
Baseline drifting	Accumulation of stationary phase	Remove the end section of the column
	Carrier gas cylinder pressure too low to allow control	Replace the carrier gas cylinder. Increase the pressure
	Drifting carrier gas or combustion gas flows	Check the gas controllers
	Accumulation of impurities in the column	Check impurity levels in the gas source. Use correct gas purity. Replace or install appropriate Gas Filters
Baseline falling	Carrier gas leak in the system	Perform a leak test. Check the tightness of the connections on the carrier gas line
	Column is baking out	Allow enough time for the column to stabilize
Baseline falling away slowly after a high initial value	Purge valve left closed during acquisition	Alter the GC program. See your GC user manual for details
	Inadequate purge flow rate	Increase the purge flow rate
	Purge valve left closed for too long	Shorten the purge time
	Solvent tail peak	Increase the solvent delay. Shorten the purge time
	Pre-filters are dirty. (when using a quadrupole MS detector)	Contact your service representative
Baseline rising	Accumulation of impurities in the column	Check impurity levels in the gas source. Use correct gas purity. Replace or install appropriate Gas Filters
	Contaminated detector	Check the detector and clean it
	There is bleeding from the GC column	Condition column. Change the column
	Air is leaking into the system	Trace and repair the leak.
Baseline rising under temperature program control	Column contaminated	Recondition the column
Baseline high standing current	Carrier gas flow rate too high	Reduce the carrier gas flow
	Column contaminated	Recondition the column
	Contaminated gases	Replace gas cylinders. Replace the gas filters
	Excessive column stationary phase bleeding	Check the oven temperature, ensuring that it doesn't exceed the column upper limit. Recondition the column. Replace the column
	Loose connections	Ensure that all interconnections and screw connections are tight
Baseline irregular shape: dip after solvent peak	Detector contaminated	Bake out the detector. Clean the detector
Baseline irregular shape: s-shaped	Excessive column bleed during column temperature programming.	Reduce the upper column temperature. Bake out the column. Install a high temperature column
	Oxygen contamination is decomposing the stationary phase	Install oxygen filters in the carrier gas line. Check the pneumatic and inlet systems for leaks. Use correct gas purity with low oxygen content
Baseline high frequency noise	Contaminated detector	Isolate the detector from the electronics. If noise disappears, clean the collector
	Combustion gas flow too low or too high	Check the detector gas flows
	Column contaminated	Condition the column
	Contaminated detector gas supply	Check the gas purity and install appropriate filters
	Detector temperature higher than column maximum temperature	Reduce the detector temperature to the column temperature upper limit
	Loose column fittings	Tighten fittings accordingly
Baseline spiking	Column too close to flame. (when using an FID)	Lower the column to the correct position (2-3mm below the tip of the jet)
	Dirty jet or detector	Isolate the detector from the electronics. If the spiking disappears, clean the jet and the collector
	FID temperature too low. (when using an FID)	Increase the FID temperature to at least 150°C

GC troubleshooting (continued)

Symptom	Cause	Recommended solutions
Peak-Related Problems		
Peaks broadening	Column flow too high	Reduce the flow to slightly above optimum
	Column flow too low	Increase the flow to slightly above optimum
	Split flow too low in split injection	Increase the flow to 40-50mL/min
	Column performances degraded	Test the column at the optimum flow rate
	Dirty injector	Clean or replace the liner
	Stationary phase accumulated in the outlet	Remove the last two coils from the column
	Detector base body temperature too low	Increase the temperature to 5°C below the column maximum
	The sample is overloading the column	Reduce the amount and/or concentration of the sample
Double peaks	Injection speed too low	Inject more rapidly in a smooth motion
	Wrong autosampler injection speed or mode	Use a higher speed
Peak fronting	Column or detector overloaded	Decrease the injected amount. Decrease the analyte concentrations. Increase the split ratio
	Column temperature too low	Increase the temperature
	Stationary phase too thin	Use a thicker-film column
	Poor injection technique	Repeat, with better injection technique
Ghost peaks	Contaminated carrier gas	Replace the cylinder. Replace the filter
	Contamination from laboratory glassware	Ensure the glassware is clean and contamination-free
	Decomposition of injected sample	Decrease the injection port temperature. Use the on-column injection technique
	Dirty injection solution	Carry out adequate clean-up of sample prior to injection
Broad ghost peaks	Contaminated inlet or pneumatics	Remove the column and bake out the inlet. Use a high-quality septum. Replace the split vent filter. Install an in-line filter between the pneumatics and the inlet
	Incomplete elution of previous sample	Increase the final oven program temperature or total run time. Increase the column flow rate
Irregular, chair-shaped peaks	Solvent flooding of column	Increase the initial oven temperature. Reduce the injection volume (On-column). Install a retention gap (On-column)
No peaks after solvent peak	Carrier gas flow too high	Reduce the carrier gas flow rate
	Combustion gas flow incorrect	Check the combustion gas flow
	Detector contaminated	Bake out or clean the detector
	FID flame extinguished by solvent peak	Check the detector temperature and that flame is lit
	Too much sample injected	Inject less sample
	Incorrect column position in S/SL injector (too high)	Check the column position
No peaks at all	Clogged syringe needle	Replace or repair the syringe
	Column broken or disconnected	Check the column and connections
	Defective electrometer or amplifier	Check electrometer or amplifier and associated connections. Replace if required
	Defective recording device	Replace the recording device
	FID flame is out	Clean FID jet, check detector gas flows and re-light flame
	Incorrect column position in S/SL injector (too high)	Check the column position
Sample peak tailing	Column degradation causing activity	Inject a test mixture and evaluate the column
	Column/oven temperature too low	Increase the column/oven temperature. Do not exceed the recommended maximum temperature for the stationary phase.
	Column contaminated at inlet	Trim first 10-20cm from column and re-install in injector
	Glass wool or inlet liner causing activity	Replace with fresh silanized wool and a clean inlet liner
	Inlet temperature too low	Increase the inlet temperature
	Poor or obstructed column connections	Remake the column inlet connection
	Wrong stationary phase	Replace the column according to the column manufacturer's literature
Solvent peak tailing	Incorrect column position in inlet	Reinstall the column
	Initial oven temperature too high (On Column)	Reduce the initial oven temperature
	Septum purge flow too low and/or split/splitless vent flow too low	Check and adjust the septum purge and vent flows
	Too large injection size	Reduce the injection size

Symptom	Cause	Recommended solutions
Unresolved peaks	Carrier gas flow rate too high	Reduce the carrier gas flow rate
	Column deteriorated	Replace the column
	Column temperature too high	Lower the column oven temperature
	Column too short	Use a longer column
	Incorrect column choice	Install a suitable column
	Injection technique is not adequate	Choose a correct injection technique
Discrete high-intensity contaminant peaks	Bleed from the GC column	Condition or change the column
	Bleed from the septum	Replace the septum
	Sample vial septa are contaminating the sample	Discard sample. Store samples upright, in a refrigerator. Use Teflon™ faced septa, with the Teflon facing downwards (i.e. towards the sample)
Results-related problems		
Low reproducibility of peak area	Concentration not compatible with the dynamic range of the detection system	Ensure that the sample concentration is suitable for the detection system
	Inappropriate injection technique	Try a different injection technique
	Injection parameters inappropriate	Check the injection temperature. Check the flow rates
	Non reproducible sample injection technique	Evaluate the sample preparation sequences. Compare the results with a series of standard injections
	Leaking syringe or septum	Check and replace the syringe at regular intervals. Check and replace septum at regular intervals
	Leaks at the injection	Check the column connections. Run a leak check
	Poor injection technique	Carefully meter the injected amount. Use a clean, good-quality syringe
	Poor split flow or ratio control	Monitor the flow. Replace the in-line filter
Poor sensitivity increased retention time	Carrier gas flow rate too low	Increase the carrier gas flow rate. Locate and remove possible obstructions in the carrier gas line. Check the injector/column ferrules
Poor sensitivity with normal retention time	Oven or injector parameters are not optimized	Adjust the oven parameters. Adjust the injector parameters
	Leaks in the GC carrier gas line	Run a leak test and correct leaks
	Syringe leaks during injection	Replace syringe or piston seals, if applicable
	Split injection temperature too low	Increase the temperature of the injector
	Column is in poor condition, or wrong column type used	Condition the columns. Change the column
Retention times decreasing	Stationary phase deteriorated by oxygen and/or water	Use a carrier gas free of oxygen and water. Replace or install appropriate gas filters
	Stationary phase loss due to column bleeding	Reduce the column temperature
Retention times increasing	Increasing carrier leakage	Check the septum and column connections
	Carrier gas supply running out	Replace the bottle
Low reproducibility of retention times	Drifting or unstable pneumatic controller	Monitor the column pressure or flow. Check and replace the controller if necessary
	Poor injection technique	Start the run at consistent time after injection
	Sample size is too large	Reduce the injected amount and/or volume
	Unstable column temperature	Check the main oven door and cooling flap. Monitor the column temperature
Retention times are inconsistent	GC column is in poor condition	Condition the column. Change the column
	Insufficient equilibration time set on GC	Increase equilibration time
	Poor injection	Repeat with better injection technique
	Oven temperature programmed to rise too quickly	Reduce oven temperature ramp rate
	Air is leaking into the system at the injector seal or the carrier gas manifold	Trace and repair the leak

GC equations

Adjusted retention time (t_R')

An analyte's retention time (t_R) minus the elution time of an unretained peak (t_m).

$$t_R' = t_R - t_m$$

Adjusted retention time is also equivalent to the time the analyte spends in the stationary phase.

Capacity factor (k)

Expression that measures the degree of retention of an analyte relative to an unretained peak, where t_R is the retention time for the sample peak and t_m is the retention time for an unretained peak. A measurement of capacity will help determine whether retention shifts are due to the column (capacity factor is changing with retention time changes) or the system (capacity factor remains constant with retention time changes).

$$k = \frac{t_R - t_m}{t_m}$$

Thus, the higher the capacity factor, the longer the retention time.

Effective theoretical plates (N_{eff})

A measure of a column performance that accounts for the effects of unretained elution time, where t_R' is the adjusted retention time and s is the standard deviation of the peak.

$$N_{eff} = \left(\frac{t_R}{\sigma}\right)^2$$

This value also remains constant as retention gaps and guards are used. Depending on the method of peak width calculation, different efficiencies can be reported. This leads to two popular measures:

$$N_{eff} = 16\left(\frac{t_R}{W}\right)^2$$

Where W is the tangential peak width (13.4% peak height).

$$N_{eff} = 5.54\left(\frac{t_R}{W}\right)^2$$

Where W is the width measured at half height (50% peak height).

HEEP (H_{eff})

Height Equivalent to an Effective Plate.

$$H_{eff} = L/N_{eff}$$

Where L is the column length. The smaller the N_{eff}, the more efficient the column's performance.

HETP (H)

Height Equivalent to a Theoretical Plate is a measure of column efficiency where L is the column length and N is the number of theoretical plates.

$$H = L/N$$

HETP is based on actual (t_R) rather than adjusted retention times (t_R').

Linear velocity (u)

Mobile phase flow rate expressed in cm/s and is expressed as:

$$u = L/t_m$$

Where L is the column length and t_m is the breakthrough time of an unretained peak.

Phase ratio (β)

The ratio of the volume of mobile phase to the stationary phase. An important value when changing the column dimensions in a method.

$$\beta = \frac{\text{column ID (µm)}}{4 \times \text{film thickness (µm)}}$$

Resolution

A measure of the separation of two peaks taking into account both the difference in elution time and the peak widths.

$$R_s = \frac{(t_2 - t_1)}{0.5\,(W_1 + W_2)}$$

Where t_2 and t_1 are the two retention times, and W_1 and W_2 are baseline peak widths.

Selectivity (α)

The relative retention of two adjacent peaks. Selectivity can be calculated using capacity factor.

$$\alpha = \frac{k_2}{k_1}$$

Trennzahl number

A value to describe a separation. The Trennzahl number is calculated from the resolution between two consecutive homologous hydrocarbons. The Trennzahl number represents the number of peaks that can be included between the two hydrocarbon peaks.

$$T_Z = \left(\frac{t_{R2} - t_{R1}}{(W_h)_1 + (W_h)_2}\right) - 1$$

Where t_R equals analyte retention time and W_h equals peak width at half height.

van Deemter equation

This is a relationship that considers the effect of linear velocity on the HETP or H, where

A accounts for eddy diffusion, B describes the molecular diffusion of the vapor in the direction of the column axis, C refers to the resistance to transfer from the stationary to mobile phase and u is the linear velocity of the mobile phase.

$$H = A + \frac{B}{u} + Cu$$

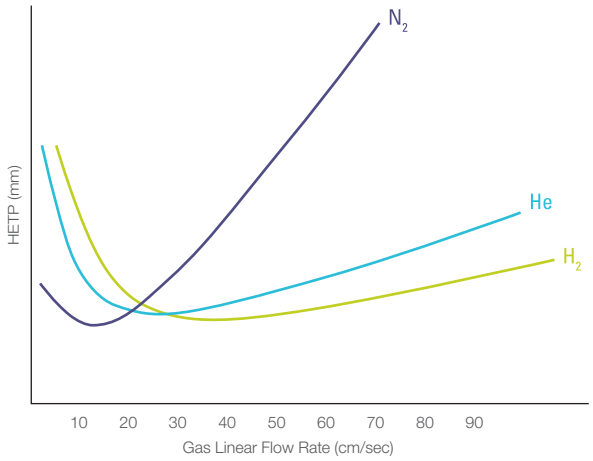
Carrier gas choice

The choice of carrier gas is a compromise between a number of considerations, among them, efficiency and speed as well as availability, safety and cost. The three most common carrier gases used are nitrogen, helium and hydrogen.

Nitrogen shows the lowest HETP, making it the most efficient of the gases. High quality nitrogen is readily available and inexpensive compared to other options. However, the optimum flow rate to achieve nitrogen's very low HETP leads to long analysis times (see figure).

Helium has a slightly lower efficiency than nitrogen, but the optimum flow rate is higher. Also small changes in flow rate of helium around the optimum will not affect efficiency as greatly as with nitrogen.

For many, hydrogen is the carrier gas of choice. It shows higher efficiency than helium and at a higher flow rate. The variation in HETP with changes in flow rate is also far lower, making it more forgiving and reproducible. There is, however, a slight risk of an explosive atmospheric build-up in the oven.



A van Deemter plot of efficiency against linear flow rate for three carrier gases.

Recommended flow rates and velocities for capillary columns

Carrier gas	0.25mm ID		0.32mm ID		0.53mm ID	
	mL/min	cm/min	mL/min	cm/min	mL/min	cm/min
He	1	35	1.7	35	6	35
H_2	1.6	50	2.6	50	7.5	50
N_2	0.4	14	0.5	11	0.9	7

Recommended detector gas flow rates

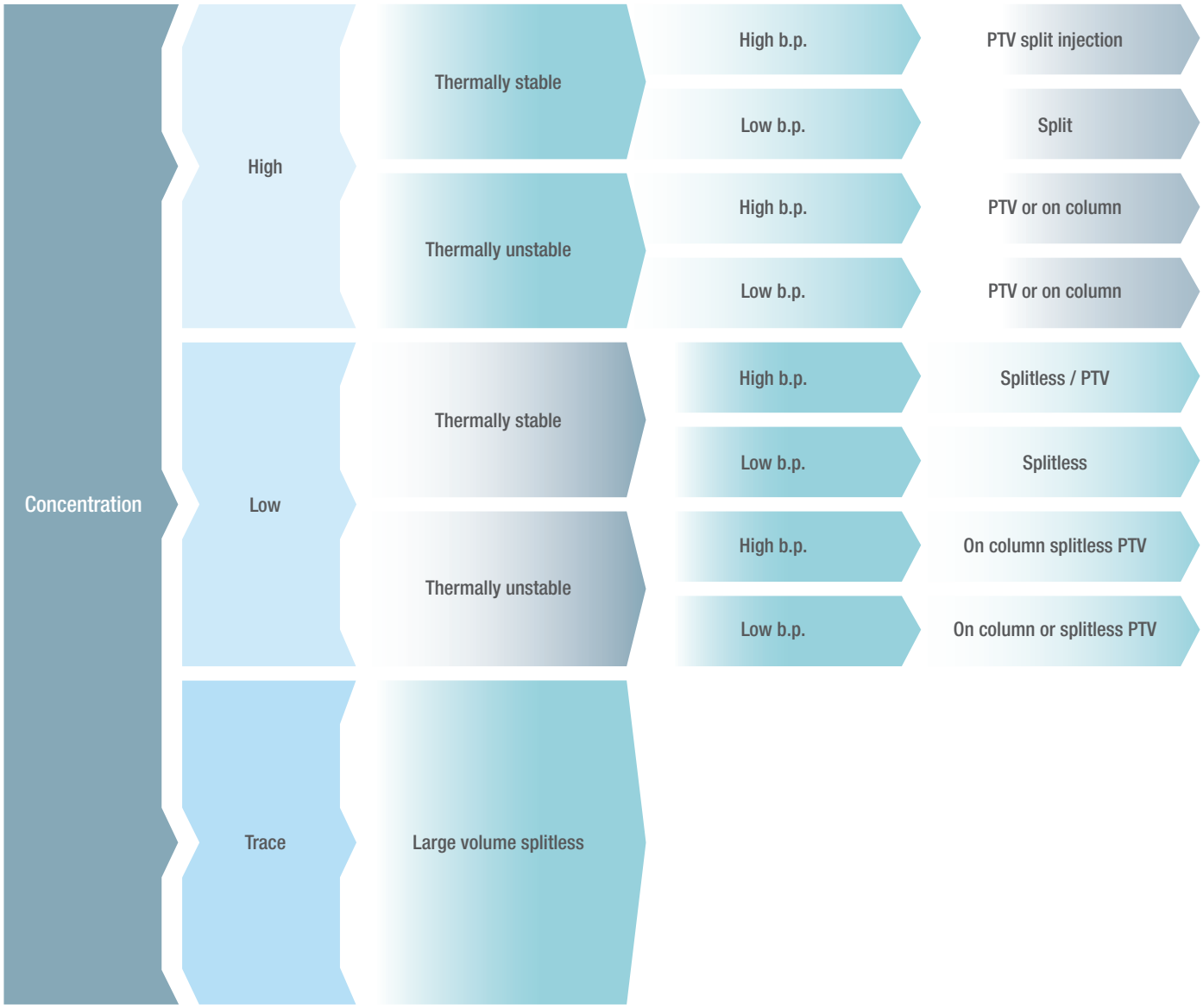
Detector	Air (mL/min)	H_2 (mL/min)	Make Up (mL/min)
ECD	–	–	35-40
FID	350	35	30
NPD	60	2.5	15
FPD	100	75	30

Unretained compounds

Detector	Analyte
FID	Methane
ECD	Methylene Chloride
NPD	Acetonitrile
TCD, MS	Methane, Butane
PID, ELCD	Vinyl Chloride

Selection of injection method

The identification of the most appropriate injection method relies on the sample type and the boiling point to be used in the separation. The diagram below summarizes this selection process:



Column conditioning (All Columns except TG-WaxMS, TRACE TR-1MS and TR-WaxMS)

It is recommended that before the column is subjected to any thermal gradients, all oxygen has been removed because the presence of oxygen in the system can shorten the column lifetime. Removal of oxygen can be achieved by purging the columns with oxygen-free carrier gas for a minimum of 20 minutes at 40°C using an approximate head pressure of 100kPa.

Although all Thermo Scientific columns have been pre-conditioned, we recommend that they are conditioned after installation by following these steps:

1. Heat the column from 50 °C at 5 °C/min to a temperature 20 °C above the highest operating temperature of the method, or the column's maximum isothermal temperature, whichever is lower and hold for 1 hour. The column's maximum isothermal temperatures are provided below.
2. Monitor the detector signal during conditioning until a stable baseline is reached. Due to the factory pre-conditioning of the column, this should be achieved in approximately one hour. This duration may be longer in the case of thick films and polar phases.

Maximum operating temperatures for TraceGOLD and TRACE GC Columns

Column	* Max Temp °C (isothermal/programmable)
TG-1MS	330°C / 350°C
TG-XLBMS	360°C
TG-5MS	330°C / 350°C
TG-SQC	330°C / 350°C
TG-5MS AMINE	300°C / 315°C
TG-5SILMS	330°C / 350°C
TG-5HT	380°C / 400°C
TG-35MS	340°C / 360°C
TG-35MS AMINE	220°C
TG-17MS	300°C / 320°C
TG-17SilMS	340°C / 360°C
TG-1301MS	260°C / 280°C
TG-624	240°C
TG-624SilMS	320°C
TG-VRX	260°C
TG-VMS	260°C
TG-1701MS	260°C / 280°C
TG-225MS	240°C
TG-200MS	320°C / 340°C
TG-POLAR	275°C
TG-WaxMS	260°C
TG-WaxMS A	250°C
TG-WaxMS B	220°C
TG-Dioxin	340°C
TG-OCP I / TG-OCP II	340°C
TG-OPP I / TG-OPP II	330°C
TG-ALC I / TG-ALC II	260°C
TG-1MT	430°C
TG-5MT	430°C
TG-WaxMT	260°C

Column	* Max Temp °C (isothermal/programmable)
TR-1MS	340°C / 360°C
TR-5	320°C / 340°C for films ≤ 1.5µm 280°C / 300°C for films > 1.5µm
TR-5MS	360°C / 370°C for films ≤ 1.5µm 350°C / 360°C for films > 1.5µm
TR-5HT	380°C / 400°C
TR-35MS	330°C / 360°C
TR-1701	280°C / 300°C
TR-50MS	360°C / 370°C
TR-225	230°C / 250°C
TR-Wax	260°C / 280°C for films ≤ 1.0µm 240°C / 260°C for films > 1.0µm
TR-WaxMS	260°C / 280°C
TR-FFAP	240°C / 250°C
TR-SimDist	400°C for films ≤ 1.0µm 370°C for 2.65µm films
TR-V1	280°C / 300°C
TR-FAME	250°C / 260°C
TR-524	240°C / 260°C
TR-525	340°C / 360°C
TR-527	330°C / 350°C
TR-8095	360°C / 370°C
TR-8270	330°C / 350°C
TR-PCB 8MS	330°C / 350°C
TR-Dioxin 5MS	330°C / 350°C
TR-Biodiesel (M)	300°C / 320°C
TR-Biodiesel (F)	280°C / 300°C
TR-Biodiesel (G)	380°C / 400°C
TR-DoA5	330°C / 350°C
TR-DoA35	330°C / 350°C
TR-Pesticide	330°C / 350°C
TR-Pesticide II	330°C / 350°C
TR-Pesticide III	300°C / 320°C
TR-Pesticide IV	300°C / 320°C

*** Maximum temperature may vary with different column film thickness. Refer to box label for more details.**

Column conditioning for the TraceGOLD, TG-WaxMS, TRACE TR-WaxMS and TR-1MS columns

This procedure will ensure an ultra low bleed for the column's entire lifetime and is only required once. Once performed, future installation of the column need only be followed by a 30-minute hold at the maximum temperature limit.

After installing the column according to the instrument manufacturer's instructions, follow the procedure below.

Steps	TG-WaxMS/TR-WaxMS	TR-1MS
1	Equilibrate the column at 40°C with carrier gas flow for 20 minutes, purging air content.	Equilibrate the column at 40°C with carrier gas flow for 20 minutes, purging air content.
2	Raise the temperature to 100°C at 5°C/min.	Raise the temperature to 100°C at 5°C/min.
3	Hold for 30 minutes.	Hold for 30 minutes.
4	Raise to 150°C at 5°C/min.	Raise to 150°C at 5°C/min.
5	Hold for 30 minutes.	Hold for 30 minutes.
6	Raise to 200°C at 5°C.	Raise to 250°C at 5°C.
7	Hold for 40 minutes.	Hold for 40 minutes.
8	Raise to 250°C at 5°C/min.	Raise to 300°C at 5°C/min.
9	Hold for 40 minutes.	Hold for 40 minutes.
10	Raise to 280°C at 5°C/min.	Raise to 360°C at 5°C/min.
11	Hold for 30 minutes.	Hold for 30 minutes.

Although quite a long procedure, it will result in longer lifetimes and lower bleed for your column.

Performance recovery

The performance of the column may exhibit signs of deterioration over time as a result of many different causes. Some of these, such as contamination by high boiling or strongly retained compounds, can be cleared by repeating the column-conditioning until a stable baseline is achieved.

Other contamination such as non-volatile compounds, pieces of septa or ferrule metal can result in poor peak shape due to band broadening at the injection step. This can be cured by the removal of a section from the front end of the column. The amount removed is dependent on the degree of contamination, the size of injection and the ID of the column, but generally 50 cm should be sufficient. As the efficiency of the column is proportional to the square root of its length, the removal of the front end will not lower the separation effectiveness by the same ratio as 50 cm/column length. A last resort in column regeneration is column washing. Column washing uses a pressurized vessel to force solvent through the column in a reverse direction. The selection of the solvent is dependent on the nature of the samples that have been analyzed and therefore the contamination. It is also dependent on the stationary phase. Generally, 2 mL of pentane is suitable for non-polar contamination with methanol used for more polar samples.

GC reagents

Derivatization

Chemical literature contains an abundance of data on derivatization, most of which is relevant to particular compounds, classes of compounds and derivatization reagents. Two books are recognized as standards in the field of analytical derivatization. The first book, Handbook of Analytical Derivatization Reactions by Daniel R. Knapp[1], provides a general collection of analytical derivatization methods for chromatography and mass spectrometry (MS) that involves formation of covalent derivatives prior to analysis. The second book, Silylation of Organic Compounds by Alan F. Pierce,[2] "was a significant factor in the transfer of silylation reactions from the relatively esoteric field of organosilicon chemistry to the status of perhaps the most widely practiced of derivatization methods."[3]

Compounds or compound mixtures are derivatized before analysis for the following reasons:

1. To make a compound that otherwise could not be analyzed by a particular method suitable for analysis.[4]
2. To improve the analytical efficiency of the compound.[5,6]
3. To improve the detectability of the compound.[7]

Suitability

Often compounds cannot be analyzed because they are not in a form that is suitable for the particular analytical technique. Examples include nonvolatile compounds for GC analysis,[8,9,10] insoluble compounds for HPLC analysis and materials that are not stable using the conditions of the technique.[11] The derivatization procedure modifies the chemical structure of the compounds, allowing analysis by a desired technique.[12]

Efficiency

Direct analysis can be difficult when compounds interact with each other or with the column. These interactions can lead to poor peak resolution and/or asymmetrical peaks that make proper peak integration difficult or impractical. This interference can be reduced with conversion to derivatized products.[13,14] Compounds that exhibit co-elution can often be separated by using the appropriate derivatization methods.

Detectability

As demand increases for the analysis of increasingly smaller amounts of materials, it becomes important to extend the detectability range of the materials in question. This increased sensitivity can be accomplished by improved detector design that is directed toward specific atoms or functional groups.

Another popular approach to increase detectability is the use of derivatization. Enhanced detectability can be achieved by increasing the bulk of the compound, or by introducing atoms or functional groups that strongly interact with the detector.[16,17] This technique is performed in gas chromatographic applications, with the addition of halogen atoms for electron capture detectors,[18,19] and with the formation of TMS derivatives to produce readily identifiable fragmentation patterns and mass ions.[20]

Types of derivatization

Compounds containing functional groups with active hydrogens (-COOH, -OH, -NH and -SH) are usually derivatized prior to analysis by gas chromatography. These functional groups have a tendency to form intermolecular hydrogen bonds that affect the volatility, their tendency to interact deleteriously with column packing materials and their thermal stability. Silylation, acylation and alkylation are derivatization techniques used to alter these functional groups to improve their thermal and chromatographic character.

The ideal derivatization procedure will:

1. Accomplish the desired modification.
2. Proceed quantitatively, or at least reproducibly.
3. Produce products that are readily distinguishable and separable from the starting materials.
4. Proceed rapidly with simple and straight-forward laboratory techniques that will be both selective and applicable to a number of similar compounds.
5. Involve reagents and reactions that present no unusual hazards.

Thermo Scientific silylation reagents

Silyl derivatives are the most widely used derivatives for gas chromatographic applications. Usually they are formed by the replacement of the active hydrogens from acids, alcohols, thiols, amines, amides and enolizable ketones and aldehydes with the trimethylsilyl group. A variety of reagents is available for the introduction of the trimethylsilyl group. These reagents differ in their reactivity, selectivity and side reactions and the character of the reaction products from the silylation reagent itself. Considerable literature is available to assist you in the selection of the most suitable silylation reagent for your particular compounds or systems.[1,2]

Silylation reagents and trimethylsilyl derivatives are hydrolytically unstable and must be protected from moisture. However, the rate of hydrolysis for various reagents and derivatives is different, and sometimes it is possible to prepare derivatives in the presence of small amounts of moisture,[21] or to isolate and purify derivatives by extraction in an organic solvent, followed by washing with aqueous solutions.[22] Reagents that introduce a t-butyldimethylsilyl group instead of the trimethylsilyl group were developed for greater hydrolytic stability.[23] These derivatives provide improved stability against hydrolysis and provide distinctive fragmentation patterns, making them useful in GC-MS applications.[24]

Most trimethylsilyl and t-butyldimethylsilyl derivatives offer excellent thermal stability and are suitable for a wide range of injector and column conditions. However, as the silylation reagents will derivatize nearly all active hydrogens, it is important that they are not injected onto any column in which the stationary phase contains these functional groups. Examples of packings that are not compatible with silylating reagents are polyethylene glycols (TG-WaxMS) and free fatty acid phases (TG-WaxMS A).

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Thermo Scientific acylation reagents

Acylation is the conversion of compounds (through the action of a carboxylic acid or a carboxylic acid derivative) that contain active hydrogens such as -OH, -SH and -NH to esters; thioesters; and amides.[1] In chromatographic applications, the acylation reaction is used primarily for converting the above classes of compounds into derivatives that are better suited for chromatography[2] or that give a greater response to the chromatographic detection system than the parent compound.[3]

An important example of this application is the insertion of perfluoroacyl groups into a molecule to enhance the detectability of the substance by electron capture. The presence of a carbonyl group adjacent to the halogenated carbons enhances the electron capture detector (ECD) response.

Acyl derivatives are also useful in MS applications in which they influence the fragmentation patterns of the compounds to be studied.[4]

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Thermo Scientific alkylation reagents

When used in derivatization for gas chromatography, alkylation represents the substitution of an active hydrogen by an aliphatic or aliphatic-aromatic[1] (benzyl) group. This technique is used to modify those compounds containing acidic hydrogens, such as carboxylic acids and phenols. The principal chromatographic use of this reaction is the conversion of organic acids into esters, which produce better chromatograms than the free acids.

In addition, alkylation reactions can be used to prepare ethers, thioethers and thioesters; N-alkylamines; and amides.[2] As the acidity of the active hydrogen decreases, the strength of the alkylating reagent must be increased. As the reagents and conditions become harsher, the selectivity and applicability of the methods become more limited.

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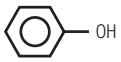
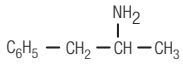
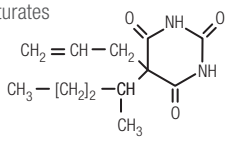
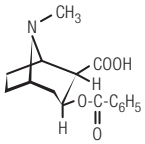
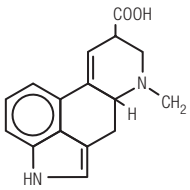
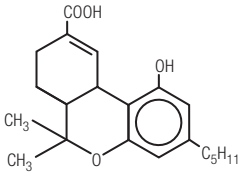
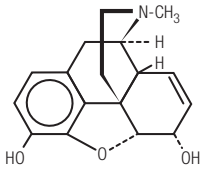
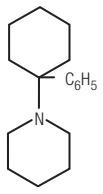
Derivatization reagents for specific functional groups

Functional group	Procedure	Reagent	Derivative	Notes
Amides O II –C–NH_2 Primary	Silylation	BSA	TMS Amides	Difficult to form due to steric hindrance
		BSTFA	TMS Amides	
		BSTFA+TMCS	TMS Amides	TMCS used as a catalyst
		MSTFA	TMS Amides	Reaction byproducts more volatile
		MSTFA+TMCS	TMS Amides	
		Tri-Sil Reagents	TMS Amides	
		MTBSTFA	TBDMCS Amides	Difficult to form; very stable
		MTBSTFA+TBDMCS	TBDMCS Amides	TBDMCS aids derivatization
O II –C–NHR Secondary	Acylation	MBTFA	Trifluoroacetamides	
		TFAA	Trifluoroacetamides	
		PFAA	Pentafluoropropionamides	Good for ECD detection
		HFBI	Heptafluorobutyamides	
	Alkylation	MethElute Reagent (TMPAH)	Methyl Amides	On-column derivatization especially for drugs
Amines H I –C–NH_2 I H Primary	Silylation	BSTFA	TMS	
		BSTFA	TMS	
		BSTFA+TMCS	TMS	TMCS aids derivatization
		MSTFA	TMS	
		MSTFA+TMCS	TMS	TMCS aids derivatization
		Tri-Sil® Reagents	TMS	
	Silylation	MTBSTFA	TBDMS	Difficult to form, but more stable
		MTBSTFA+TBDMCS	TBDMS	TBDMCS aids derivatization
–C–NHR H I –C–NHR I H Secondary	Acylation	MBTFA	Trifluoroacetamides	Good for trace analysis with ECD
		TFAA	Trifluoroacetamides	Good for trace analysis with ECD
		TFAI	Trifluoroacetamides	Good for trace analysis with ECD
		PFAA	Pentafluoropropionamides	
		PFPI	Pentafluoropropionamides	
		HFAA	Heptafluorobutyamides	
		HFBI	Heptafluorobutyamides	
	Alkylation	MethElute Reagent (TMPAH)	Methyl Amides	On-column derivatization for specific drugs
Carbohydrates $(CH_2OH)_n$	Silylation	MSTFA	TMS	
		TMSI	TMS	Can be used with some syrups
		Tri-Sil Reagents	TMS	
	Acylation	MBTFA	Trifluoroacetates	Volatile derivatives of mono-, di- and trisaccharides
		TFAI	Trifluoroacetates	
Carboxyl O II –C–OH	Silylation	BSA	TMS	Easily formed, generally not stable, analyze quickly
		BSTFA	TMS	
		BSTFA+TMCS	TMS	
		MSTFA	TMS	
		TMCS	TMS	Can be used with some salts
		TMSI	TMS	
		Tri-Sil Reagents	TMS	
		MTBSTFA	TBDMS	More stable than TMS derivatives
		MTBSTFA+TBDMCS	TBDMS	TBDMCS aids derivatization
	Alkylation	PFBBr	Pentafluorobenzyl Esters	Used in EC detection and UV, MS
		BF_3-Methanol	Methyl Esters	Best for large samples of fatty acids
		Methylate Reagent (DMFDMA)	Methyl Esters	Fatty acids and amino acids
		MethElute Reagent (TMPAH)	Methyl Esters	On-column derivatization
		PFAA+Pentafluoropropanol	Pentafluoropropyl Ester	Drug analysis
Hydroxyl-OH R–OH Alcohols  Phenols	Silylation	BSA	TMS	Most often used derivatives
		BSTFA	TMS	Good thermal stability
		BSTFA+TMCS	TMS	Poor hydrolytic stability
		HMDS	TMS	Weak donor usually used with TMCS
		MSTFA	TMS	
		MSTFA+TMCS	TMS	
		TMCS	TMS	Weak donor usually used with HMDS; can be used with salts
		TMSI	TMS	
		Tri-Sil Reagents	TMS	Can be used with syrups
		MTBSTFA	TBDMS	More stable than TMS, good MS fragmentation patterns
		MTBSTFA+TBDMCS	TBDMS	TBDMCS aids derivatization
	Acylation	MBTFA	Trifluoroacetates	Good for trace analysis with EDC
		TFAA	Trifluoroacetates	Good for trace analysis with EDC
		TFAI	Trifluoroacetates	Good for trace analysis with EDC
		PFAA	Pentafluoropropionates	Good for trace analysis with EDC
		HFBI	Heptafluorobutyrates	Good for trace analysis with EDC
		HFAA	Heptafluorobutyrates	Good for trace analysis with EDC
	Alkylation	PFBBr	Pentafluorobenzyl Ethers	With alkoxides only

Derivatization reagents for drugs of abuse

Drug	Form	Reagent
Amphetamines 	Amphetamines Amphetamines Amphetamines Amphetamines Amphetamines Methamphetamine	BSTFA HFAA HFAA/PFAA MSTFA with TMCS TFAA TFAA
Barbiturates 		BSTFA MethElute Reagent (TMPAH) Methylate Reagent (DMFDMA) PFBBr
Cocaine 	Benzoylecgonine	BSTFA/Butyl Iodine/TMPAH BSTFA MTBSTFA PFAA/PFPOH
LSD 		BSA BSTFA MSTFA TFAI
Marijuana 	THC metabolites	BSA BSTFA/BSTFA+1% TMCS BSTFA/TMCS/TMSI MSTFA MSTFA/MSTFA+1% TMCS MTBSTFA PFBBr PFAA/HFIOH PFAA/PFPOH TFAA and BF_3/MeOH MethElute Reagent (TMPAH) TMSI
Opiates 	Morphine Morphine/Codeine	BSTFA+1% TMCS MBTFA PFAA TFAA BSTFA BSTFA+1% TMCS BSTFA/TFAI HFBA MBTFA PFAA PFAA/HFAA PFAA/PFPOH TFAA Trimethylsilyl
PCP 	PPC/PCHP/PCP	BSTFA+1% TMCS HFAA

See references on following page.

† Reagent names correspond to product names as listed in this catalog, except PFPOH (pentafluoropropanol).

HFIOH (heptafluoro-isopropanol) is not offered by Thermo Fisher Scientific. PFAA (PentaFluoropropionic Acid Anhydride) and HFAA (HeptaFluorobutyric Acid Anhydride) are sometimes incorrectly referred to as PFPA and HFBA (respectively), which are the appropriate abbreviations for the free acid.

Derivatization reagents for drugs of abuse (continued)

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Drugs of abuse derivatization applications

Cocaine Metabolites

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Derivatization of Opiates

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Troubleshooting reagents

Derivatization problem	Possible cause	Recommended solution
Low Yield	Carrier, air, detector (FID) hydrogen or make-up gas flow set incorrectly	Measure flows using a Thermo Scientific GFM Pro Gas Flow Meter and set accordingly using instrument manufacturer's recommendations
	Reagent deteriorated	Store reagent properly to prevent oxygen/water contamination, temperature damage (refer to product specification sheet)
	Rate of reaction too slow	Re-evaluate reagent concentration, time, temperature and consider heating the reaction mix (consider the thermal stability of the analytes and reagents)
	Water in reaction mix	Remove water by adding sodium sulfate to sample. Store reagent properly to prevent oxygen/water contamination
	Improper handling technique: (e.g. Low boiling components could be lost during sample concentration); sample too dilute; wrong solvent	Re-evaluate technique, if possible eliminate steps in which analyte could be adsorbed or otherwise lost (unnecessary transfers etc.)
	Wrong reagent	Re-evaluate reagent selection and select more appropriate reagent
	Impurities in solvent, starting material, catalysts, or extract interfering with derivatization (e.g. Plasticizers from vial, inorganics used in sample synthesis, preservatives or antioxidants in solvents)	Use only highest purity material at all steps in the sample preparation process
	Reagent: sample ratio too low	Use more reagent for same amount of sample
	Sample adsorbed to glassware	Deactivate glassware, inlet sleeve and column by silanizing
No sample separation after adding reagent and heating	Septum in reaction vial not sealed	Prepare a new sample and derivatize. Be sure that the vial is sealed
Detector response low	Sample components absorbed by inlet liner or column	Inject standard on column known to be performing well. If results are good, remove inlet liner and check cleanliness. Use new, deactivated liner or replace glass wool and packing. Rinse bonded phase column or remove a few cm from inlet end of non-bonded column. If performance is not restored, replace column
	Low yield of derivative – reaction did not go to completion	Add more reagent, increase temperature or heating time or add catalyst. Water may be present; add sodium sulfate to sample
	Detector (FID) dirty	Clean FID as per instrument manual
Extra peak(s)	Derivative reacting with solvent	Use a solvent that does not have an active hydrogen, alcohol or enolizable ketone group (e.g. Hexane, toluene etc.)
	Impurities from sample solvent, reagents, sample vial, other labware	Inject solvent and reagents blanks, solvent rinse from unused vial etc. Isolate sources of impurities
	Reagents interacting with column	Verify that reagent is compatible with analytical column
	Derivative undergoing hydrolysis	Remove water by adding sodium sulfate to sample. Store reagent properly to prevent oxygen/water contamination
Missing peaks or solvent peak only	Wrong reagent	Re-evalaute reagent selection
	Reagent deteriorated	Store reagent properly to prevent oxygen/water contamination, temperature damage (refer to product specification sheet)
	Rate of reaction too slow	Re-evaluate reagent concentration, time, temperature and consider heating the reaction mix (consider the thermal stability of the analytes and reagents)
	Impurities in solvent, starting material, catalysts, or extract interfering with derivatization (e.g. Plasticizers from vial, inorganics used in sample synthesis, preservatives or antioxidants in solvents)	Use only highest purity material at all steps in the sample preparation process
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