

A rapid screening method of mycotoxins in grains by liquid chromatograph tandem mass spectrometry

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Overview

Development of a rapid simple screening method by LC/MS/MS for mycotoxins containing fumonisins.

Introduction

Mycotoxin is a chemical substance produced by mold. In terms of harmful substances to health of human and livestock, regulatory limitation is defined in each country. In recent years, the risk management of mycotoxins has been gaining wider acceptance all over the world. In this report, we demonstrate a simultaneous screening analysis for 18

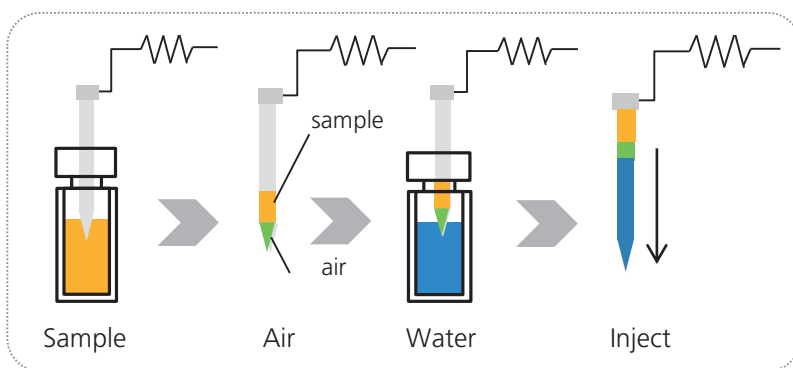
kinds of mycotoxins in grains by LC/MS/MS. For the purification of grain extract, Multitoxin Spin Column (Romer Labs) was utilized in order to perform a simple and rapid clean-up treatment for mycotoxins, which has individual chemical properties, such as aflatoxins, ochratoxin A, trichothecenes, and fumonisins.

Methods and Materials

Analytical conditions

Analysis was performed by a LCMS-8050 which was equipped with a Nexera™ X2 UHPLC. Pentafluorophenyl (PFP) bounded column was used to separate the regioisomeric pair (3-AcDON / 15-AcDON, FB2 / FB3) by gradient elution with a series of mobile phases containing ammonium acetate, acetic acid and methanol. Quantitative limits had been deemed to be less than or equivalent to the minimum values specified in EC/1886/2006. The developed method achieved the simultaneous

determination of mycotoxins such as aflatoxins (B1, B2, G1, G2), fumonisins (B1, B2, B3), ochratoxin A (OTA), trichothecenes [(3-acetyldeoxynivalenol(3-AcDON), 15-acetyldeoxynivalenol (15- AcDON), deoxynivalenol (DON), HT-2, nivalenol (NIV), T-2, zearalenone (ZEN)), Fusarenon-X (FUX) , Diacetoxy- scirpenol (DAS)] and patulin (PAT) in 15 minutes analytical cycle. This analytical method was developed by the modified LC/MS/MS method package for mycotoxin (Shimadzu Corporation, Japan).



Inject method

Improving peak shape of NIV solved in more than 50% of acetonitrile aqueous solution, the sample solutions should be injected with additional water. SIL-30AC autosampler has this useful function shown as above.

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High Speed Mass Spectrometer
Ultra Fast Polarity Switching -5 msec
Ultra Fast MRM -Max.555 transition/sec

Figure 1 LCMS-8050 triple quadrupole mass spectrometer

UHPLC conditions (Nexera™ X2 system)

Column : Mastro PFP 2 (150 mm×2.1 mm, 3 μm)
 Mobile phase A : 10 mmol/L Ammonium acetate-water
 Mobile phase B : Methanol including 2% acetic acid
 Flow rate : 0.4 mL/min
 Time program : B conc. 15%(0 min) -35%(1.51 min) –
 45%(5.50 min) - 60%(5.51 min) –
 95%(9.50-12.00 min) - 15%(12.01-15.00 min)

Column temp. : 40 °C
 Injection vol. : 2.5 μL with 50 μL Water
 Rinse R0 : Mobile phase A
 Rinse R1 : 10 mmol/L Sodium citrate aqueous solution
 Rinse R2 : Water/ Methanol / Acetonitrile/ IPA = 1/1/1/1 including 1% formic acid
 Needle rinse program : inside: R1 → R0 → R2 → R0, outside: R3(1 sec) → R0

MS conditions (LCMS-8050)

Ionization : ESI, Positive/Negative MRM mode
 DL temp. : 150 °C
 Interface temp. : 200 °C
 Heat block temp. : 400 °C
 Nebulizer gas : 2.5 L/min
 Heating gas : 15 L/min
 Drying gas : 5 L/min

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Table 1 MRM transitions of Mycotoxins

No.	Mycotoxin	Retention Time (min)	Polarity	Precursor Ion (m/z)	Product Ion (m/z)	CE (V)
1	Nivalenol (NIV)	2.261	-	371.10	281.10	15
2	Patulin (PAT)	2.569	-	153.00	109.10	13
3	Deoxynivalenol (DON)	2.998	-	355.10	295.10	10
4	Fusarenon-X (FUX)	3.827	-	413.10	353.10	9
5	15-AcetylDeoxynivareno (15-AcDON)	5.582	+	339.10	261.10	-12
6	3-AcetylDeoxynivareno (3-AcDON)	5.751	+	339.10	231.10	-14
7	Aflatoxin G2 (AF G2)	7.197	+	331.10	245.10	-30
8	Diacetoxy- scirpenol (DAS)	7.480	+	384.20	307.10	-13
9	Aflatoxin G1 (AF G1)	7.433	+	329.10	243.10	-27
10	Aflatoxin B2 (AF B2)	7.669	+	315.10	259.10	-30
11	Fumonisin B1 (FB1)	7.804	+	722.40	334.10	-42
12	Aflatoxin B1 (AF B1)	7.904	+	313.10	241.10	-39
13	HT-2 toxin (HT-2)	8.060	+	442.20	263.10	-13
14	Fumonisin B3 (FB3)	8.107	+	706.40	336.10	-35
15	Fumonisin B2 (FB2)	8.475	+	706.40	336.10	-39
16	T2-toxin (T-2)	8.705	+	484.30	185.10	-23
17	Ochratoxin A (OTA)	8.987	+	404.10	239.10	-24
18	Zearalenone (ZEN)	9.532	-	317.10	131.10	30

Sample preparation

Analytical samples were prepared through the extraction protocol of MycoSpin™400 (Romer Labs), which is a very convenient method without evaporator nor nitrogen purge procedures. The operation of MycoSpin™400 was completed within 5 minutes.

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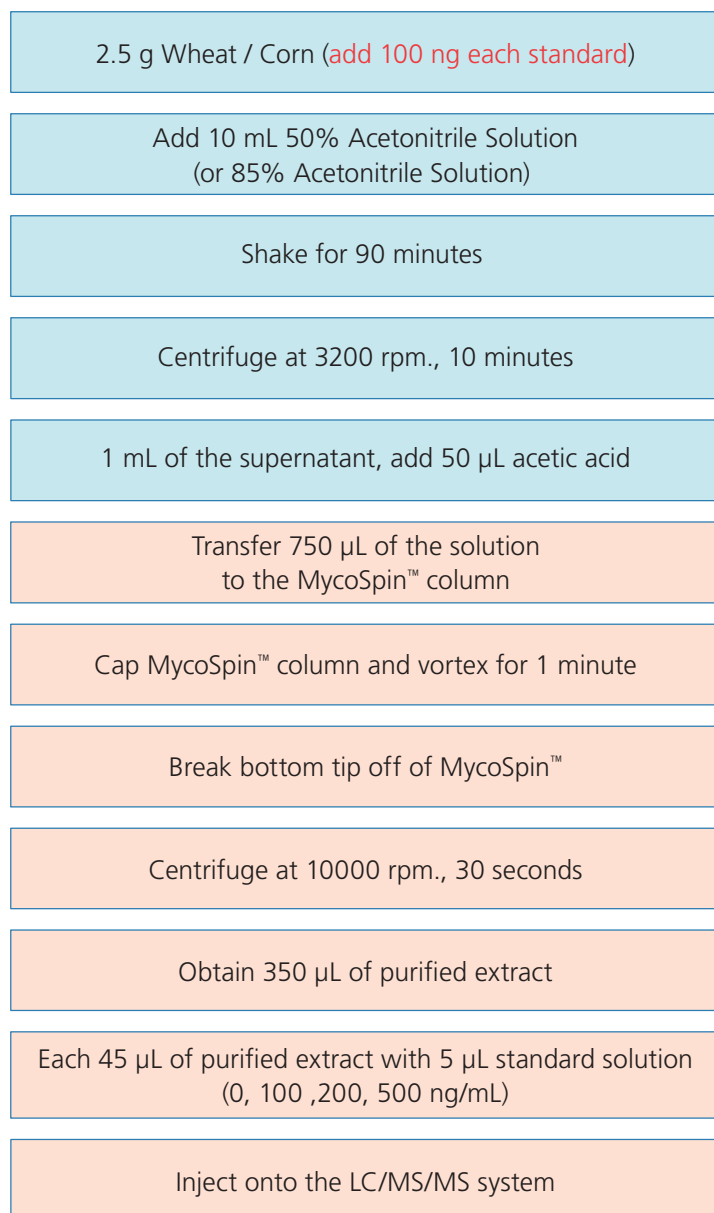


Figure 2 Protocol of sample preparation

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Result

Analysis of Standard Solution

Figure 3 shows MRM chromatograms of the 18 mycotoxin standards (each 10 ng/mL).

At first, we evaluated the solvent for better recovery of the mycotoxin from MycoSpin™ column.

In comparison, with 85% acetonitrile aqueous solution, better recovery of fumonisins was obtained with 50% acetonitrile aqueous solution (Fig.4). Thus, we decided to use 50% acetonitrile aqueous solution for the extraction solvent.

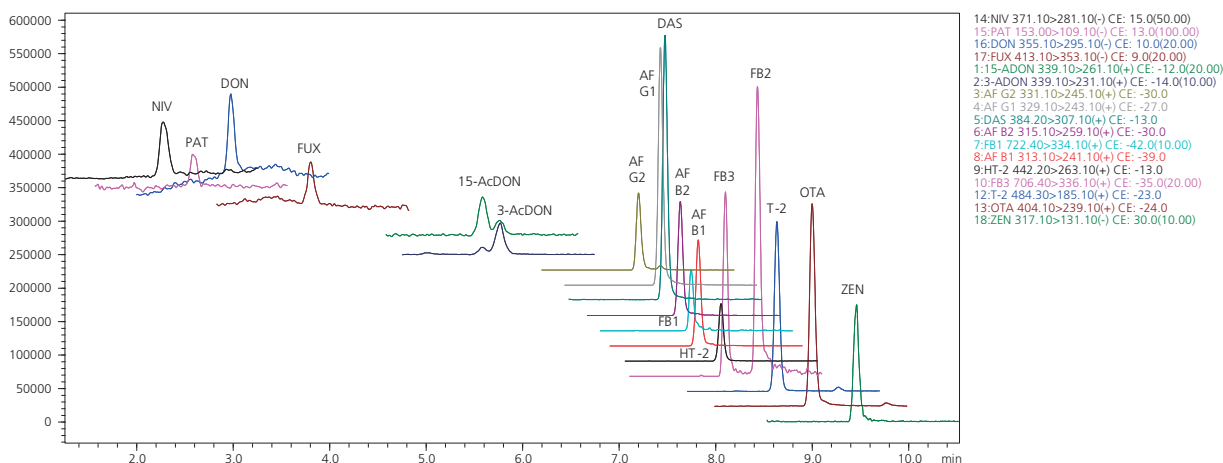


Figure 3 MRM chromatograms of the 18 mycotoxin standards (each 10 ng/mL).

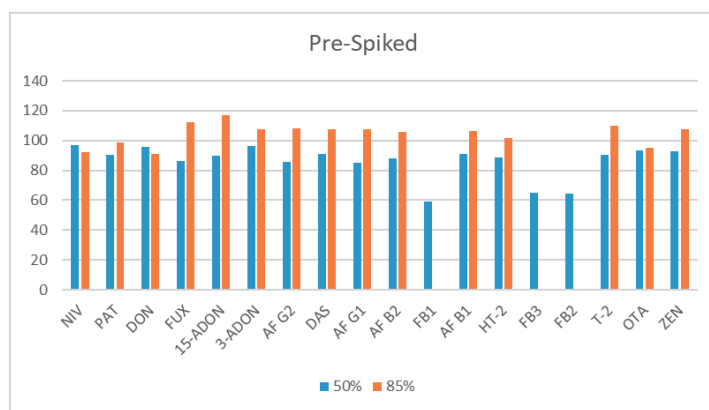


Figure 4 Recovery (%) of the mycotoxin standard from MycoSpin™ column. (Each 10 ng/mL standard mixture was applied on the column.)

Evaluation of the matrix effect

Figure 5 shows recovery (%) of the mycotoxin standards in the four kinds of extraction as wheat, corn powder, peanut powder, and almond powder. MycoSpin™ protocol was convenient in short timescale. However, even after the clean-up, many matrix compounds remained and was

affected.(Fig. 5). Although under this situation, it usually requires each labeled internal standard for target compound, we tried to investigate the quantify by the standard additive method.

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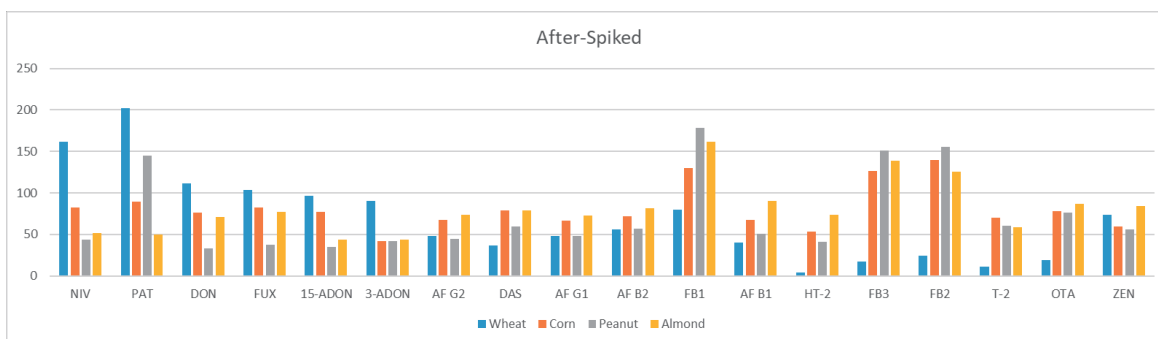


Figure 5 Recovery (%) of the mycotoxins under the four kinds of extractions. The each standard was spiked after clean-up procedure (final each 10 ng/mL).

Quantitative analysis

The results of quantitative analysis from wheat and corn powder using the sample preparation protocol (2.2) is summarized in table 2 through using the standard additive method instead of the internal standard method. The results indicates that the standard addition calibration

method could help correct and improve the recovery rate even under the influence of the matrix effect. Using this method, only a small amount of mycotoxins were detected in corn powder below the regulation value.

Table 2 The results of wheat and corn powder

	Wheat			Corn		
	Result (mg/Kg)	Recovery (%)	%RSD (n=2)	Result (mg/Kg)	Recovery (%)	%RSD (n=2)
NIV	N.D.	93	0.86	N.D.	134	2.00
PAT	0.0350	94	5.36	N.D.	79	15.60
DON	0.0148	128	6.34	0.1376	114	10.66
FUX	N.D.	101	0.30	N.D.	149	0.02
15-AcDON	N.D.	119	10.73	0.0213	114	14.30
3-AcDON	N.D.	106	7.64	N.D.	117	8.26
AF G2	N.D.	104	4.27	N.D.	115	1.80
DAS	N.D.	118	4.24	N.D.	137	2.25
AF G1	N.D.	101	3.45	N.D.	119	1.11
AF B2	N.D.	110	0.07	N.D.	125	5.95
FB1	N.D.	60	18.33	0.0407	94	15.16
AF B1	N.D.	104	2.06	0.0011	124	2.32
HT-2	N.D.	107	9.21	0.0008	132	3.91
FB3	N.D.	70	19.66	0.0078	83	7.19
FB2	N.D.	66	6.23	0.0094	79	5.80
T-2	N.D.	105	4.79	0.0005	119	1.80
OTA	N.D.	97	12.29	N.D.	109	4.38
ZEN	N.D.	97	0.90	0.0120	103	3.37

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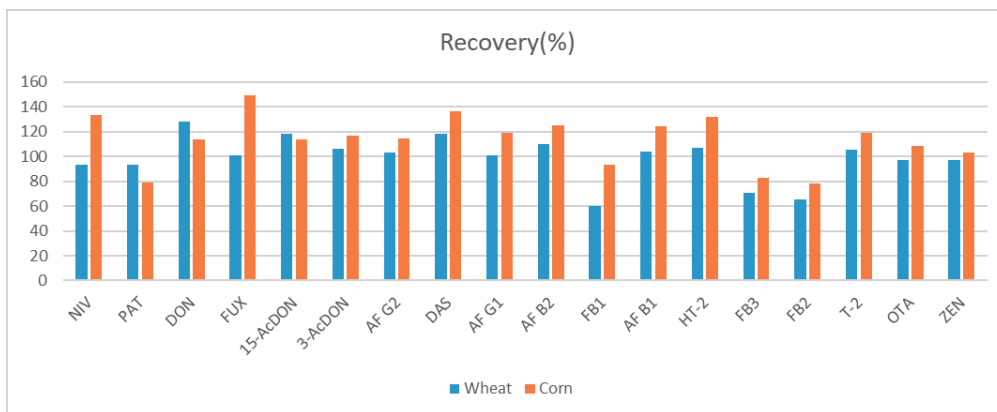


Figure 6 Recovery (%) of the mycotoxin standards from wheat and corn powder (Each 100 ng standard was spiked)

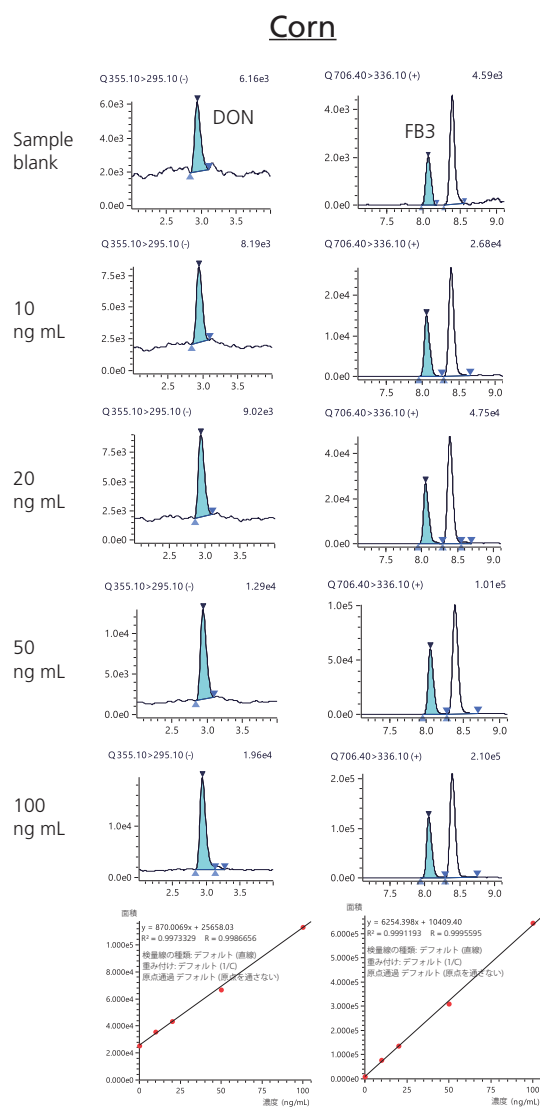


Figure 7 Chromatograms and calibration curves of the corn extraction (DON and FB3).

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Conclusions

- A rapid screening method for mycotoxins had been established.
- The LC/MS/MS method package for mycotoxin (Shimadzu Corporation, Japan) is useful tools for this type of analysis.
- The extraction solvent to improve the recovery rate of fumonisins was optimized.
- We investigated the standard addition method in order to compensate for the effect of the matrix.
- We plan to continue to evaluate multi-function columns or ion exchange columns for further improvement of the clean-up.

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